

MAGNESIUM IN MAN: IMPLICATIONS FOR HEALTH AND DISEASE

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De Baaij JHF, Hoenderop JGJ, Bindels RJM. Magnesium in Man: Implications for Health and Disease. *Physiol Rev* 95: 1–46, 2015; doi:10.1152/physrev.00012.2014.—Magnesium (Mg^{2+}) is an essential ion to the human body, playing an instrumental role in supporting and sustaining health and life. As the second most abundant intracellular cation after potassium, it is involved in over 600 enzymatic reactions including energy metabolism and protein synthesis. Although Mg^{2+} availability has been proven to be disturbed during several clinical situations, serum Mg^{2+} values are not generally determined in patients. This review aims to provide an overview of the function of Mg^{2+} in human health and disease. In short, Mg^{2+} plays an important physiological role particularly in the brain, heart, and skeletal muscles. Moreover, Mg^{2+} supplementation has been shown to be beneficial in treatment of, among others, preeclampsia, migraine, depression, coronary artery disease, and asthma. Over the last decade, several hereditary forms of hypomagnesemia have been deciphered, including mutations in *transient receptor potential melastatin type 6 (TRPM6)*, *claudin 16*, and *cyclin M2 (CNNM2)*. Recently, mutations in *Mg²⁺ transporter 1 (MagT1)* were linked to T-cell deficiency underlining the important role of Mg^{2+} in cell viability. Moreover, hypomagnesemia can be the consequence of the use of certain types of drugs, such as diuretics, epidermal growth factor receptor inhibitors, calcineurin inhibitors, and proton pump inhibitors. This review provides an extensive and comprehensive overview of Mg^{2+} research over the last few decades, focusing on the regulation of Mg^{2+} homeostasis in the intestine, kidney, and bone and disturbances which may result in hypomagnesemia.

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I. INTRODUCTION

Magnesium (Mg^{2+}) is an essential ion for health. Mg^{2+} plays an important role in the physiological function of the brain, heart, and skeletal muscles. Mg^{2+} has anti-inflammatory properties and acts as Ca^{2+} antagonist. The United States Food and Nutrition Board recommends a daily intake of 420 mg for men and 320 mg for women (1). However, recent reports estimate that at least 60% of Americans do not consume the recommended daily amount of Mg^{2+} (281). Part of the problem stems from the soil used for agriculture, which is becoming increasingly deficient in essential minerals. Over the last 60 years the Mg^{2+} content in fruit and vegetables decreased by 20–30% (570). Moreover, the Western diet contains more refined grains and processed food. Estimates are that 80–90% of Mg^{2+} is lost during food processing. As a result, a significant number of people are Mg^{2+} deficient, which may comprise up to 60%

of critically ill patients (84, 145). Mg^{2+} deficiency is commonly determined by measuring total serum Mg^{2+} concentrations, which ranges between 0.7 and 1.05 mM in a healthy person (323). However, serum Mg^{2+} values reflect only 1% of the body Mg^{2+} content, since most of the body's Mg^{2+} is stored in bone, muscle, and soft tissues. Therefore, although serum values are within the normal range, the body can be in a severely Mg^{2+} -depleted state. Consequently, the clinical impact of Mg^{2+} deficiency may be largely underestimated.

The first use of Mg^{2+} in human medicine can be traced back to 1697 when Dr. Nehemiah Grew identified magnesium sulfate ($MgSO_4$) as the major ingredient of Epsom salt (195). Epsom salt was extracted from a well in Epsom, England and was used over the years to treat abdominal pain, constipation, sprains, muscle strains, hyaline membrane disease, and cerebral edema. Subsequently, Mg^{2+} was recognized as an element (Mg) by Joseph Black in 1755 and first isolated by Sir Humphrey Davy from magnesite [$Mg_3SO_4O_{10}(OH)_2$] and mercury in 1808 (102). The role of Mg^{2+} in the human body emerged once Mg^{2+} was described in blood plasma by Willey Glover Denis in 1920 (113). In 1926, Jehan Leroy demonstrated that Mg^{2+} is essential for life in mice (309). These findings were trans-

lated to humans, and the first report of Mg^{2+} deficiency in humans was by Arthur Hirschfelder and Victor Haury in 1934 (231). Since then, Mg^{2+} has been implicated in and used for treatment of a variety of diseases, including migraines, cardiovascular diseases, and diabetes. Although the importance of Mg^{2+} is widely acknowledged, serum Mg^{2+} values are not generally determined in clinical medicine. Therefore, Mg^{2+} is often referred to as the “forgotten” cation in human health.

This review provides an overview of the role of Mg^{2+} in human health and disease. Mg^{2+} has important cellular functions in enzymatic reactions and in the synthesis and structure of proteins and polynucleotides, which are described in section II. The important regulation of Mg^{2+} homeostasis is discussed in depth in section III. The role of Mg^{2+} in organ function and related diseases is discussed in section IV. An overview is presented of the most important diseases in which Mg^{2+} disturbances have been implicated or in which Mg^{2+} has been considered as a potential treatment. In last part of the review, special attention is awarded to disturbances of intestinal Mg^{2+} uptake and renal Mg^{2+} excretion (sect. V). All together, this review emphasizes the importance of a controlled Mg^{2+} balance in the human body. Increasing the awareness and understanding of Mg^{2+} homeostasis may give more clinical attention to the important role of Mg^{2+} in health and disease.

II. MAGNESIUM IN CELLULAR PHYSIOLOGY

Within the periodic table of elements, Mg has the atomic number 12 and is classed as an alkaline earth element (group 2). Mg occurs in three stable isotopes: ^{24}Mg , ^{25}Mg , and ^{26}Mg . ^{24}Mg is the most common isotope (78,99%) and has a relative atomic mass of 24.305 Da, a melting point of 648.8°C, and a boiling point of 1,090°C (350). Mg^{2+} is highly soluble and the second most abundant cation in seawater (95). In the dissolved state, Mg^{2+} has two hydration shells, making its hydrated radius ~400 times larger than its dehydrated radius, larger than that of other cations like Na^+ , K^+ , and even Ca^{2+} (95). Consequently, Mg^{2+} needs to be dehydrated before passing through channels and transporters, a process that requires a lot of energy. Mg^{2+} is a powerful Ca^{2+} antagonist, despite both having similar charge and chemical properties.

Mg^{2+} is the second most abundant intracellular cation with typical concentrations of ~10–30 mM. However, since most of the intracellular Mg^{2+} is bound to ribosomes, polynucleotides and ATP, the concentration of freely available Mg^{2+} falls within the low millimolar range (0.5–1.2 mM) (133). In contrast to other abundant ions, for which cells maintain considerable transmembrane gradients, the free Mg^{2+} concentrations in the cell and in the extracellular fluid are comparable. Mg^{2+} is a versatile ion that is involved

in practically every major metabolic and biochemical process within the cell. Although it extends beyond the purpose of this review to give a comprehensive overview of all biochemical reactions and structural processes involving Mg^{2+} , the following paragraphs will highlight the most prominent cellular processes in which Mg^{2+} is involved.

A. Nucleotide Binding

Mg^{2+} forms an essential component of the RNA and DNA tertiary structures, as it binds the negatively charged O and N molecules within the polynucleotide chains. Polynucleotide binding is a complex biophysical process that mainly depends on the level of Mg^{2+} dehydration and the electrostatic potential at the binding site (for extensive review, see Ref. 347). The most studied Mg^{2+} -RNA interaction is tRNA, where Mg^{2+} stabilizes the structure. The role of Mg^{2+} became evident in 1966, when it was shown that Mg^{2+} could restore denatured tRNA molecules (319). Crystallographic structures of tRNAs from yeast identified five Mg^{2+} -binding sites, three in the core region around the bend of the L-shaped molecule and two additional sites in the major groove of the anticodon stem (412, 556). Additionally, there may be a few dozen Mg^{2+} in close vicinity of tRNA molecules that may bind weakly to the exterior of the structure (347). Still, the importance of Mg^{2+} binding for the tRNA tertiary structure has been contested over the years. This discussion was mainly triggered by studies showing the importance of nonspecific diffuse binding of Mg^{2+} and other divalent and monovalent cations, questioning the specificity of the Mg^{2+} interactions. However, the role of Mg^{2+} in RNA structure extends beyond tRNAs. For instance, Mg^{2+} is also crucial to the interactions that stabilize the pseudoknot conformation (191), tertiary RNA structures that are present in mRNA, ribosomal RNA, transfer-messenger RNA, catalytic self-splicing RNA, and viral genomic RNA.

In DNA, Mg^{2+} forms hydrogen bonds with the electronegative elements (O, N) to stabilize the natural DNA conformation, referred to as B-DNA (85, 549). Moreover, Mg^{2+} plays a role in the secondary and tertiary structure of DNA by competing with monovalent ions (394). Mg^{2+} binds the minor groove of B-DNA structures, thus protecting it. In Mg^{2+} -deficient conditions, DNA is more accessible to free oxygen radicals and more prone to oxidative stress (406). However, at higher Mg^{2+} concentrations, Mg^{2+} may covalently bind DNA, locally distorting the double helix (22). Therefore, maintaining the cellular Mg^{2+} concentration within the physiological range is essential for DNA stability.

B. Enzymatic Activity

In medical textbooks and scientific literature, Mg^{2+} is often described as a cofactor for ~300 enzymes. Theodor

Günther introduced the number 300 as a rough estimate in 1980 and this has been in use ever since (133). However, in the decades after 1980 many new Mg^{2+} -dependent enzymes have been described, and the number 300 is, therefore, an underestimation. Currently, enzymatic databases list over 600 enzymes for which Mg^{2+} serves as cofactor, and an additional 200 in which Mg^{2+} may act as activator (32, 73). An overview of these Mg^{2+} -dependent enzymes can be found at MetaCyc (<http://www.metacyc.org>; Ref. 73). Many of the enzymes that require Mg^{2+} as coactivator are vital for life.

Mg^{2+} is necessary for the proper structure and activity of DNA and RNA polymerases (56, 500). DNA polymerases have two Mg^{2+} binding sites, which are hypothesized to play a key role in the conformational changes in the polymerase enzyme during the catalytic reaction (56). This model was further enhanced by studies reporting that the release of one of the Mg^{2+} ions is necessary for opening the catalytic site for new nucleotides (577). In addition, Mg^{2+} is an important factor in DNA repair mechanisms within the cell, including nucleotide excision repair (NER), base excision repair (BER), and mismatch repair (MMR). Mg^{2+} acts as cofactor for almost every enzyme involved in basically every step of NER (68). In BER, Mg^{2+} is elemental for the activity of endonucleases, which incise the DNA after DNA damage, and the DNA polymerases and ligases, which repair the gap (41, 473). The third repair pathway, MMR, is also affected by Mg^{2+} availability since several enzymes involved require Mg^{2+} and ATP for activity (33). Other enzymes requiring Mg^{2+} are topoisomerases, helicases, exonucleases, protein kinases, cyclases, and large groups of ATPases, meaning that Mg^{2+} is an essential component of

DNA replication, RNA transcription, amino acid synthesis, and protein formation. Altogether, Mg^{2+} is a key factor in the maintenance of genomic and genetic stability. The consequences of low Mg^{2+} availability on the development of cancer is discussed in section IIE1.

Mg^{2+} is also an important regulator of many enzymes involved in glycolysis, because it is a cofactor for adenine nucleotides. Mg -ATP is required for the activity of hexokinase, phosphofructokinase, aldolase, phosphoglycerate kinase, and pyruvate kinase (166, 550). Consequently, Mg^{2+} availability is of major importance for glucose metabolism, which may explain its role in diabetes mellitus type 2 (see sect. IVE1).

The role of Mg^{2+} , however, extends far beyond DNA and protein synthesis, DNA repair, and glycolysis. Since kinases, ATPases, guanylyl cyclases, and adenylyl cyclases all depend on Mg -ATP for proper function, Mg^{2+} plays a role in virtually every process in the cell.

C. Cellular Mg^{2+} Handling

Since protein and DNA synthesis are highly dependent on intracellular Mg^{2+} availability, intracellular Mg^{2+} concentrations are tightly regulated. Over the last 20 years, elucidating the molecular identities of the transporters involved in Mg^{2+} homeostasis has been the main focus of research within the Mg^{2+} field. Genetic screenings on human diseases and microarray-based expression studies have resulted in the identification of numerous Mg^{2+} -transporting proteins (TABLE 1 and FIGURE 1). Although the exact role of many of these proteins needs further investigation, re-

Table 1. Mg^{2+} Transporters

Name	Membrane	Expression	Permeability	Mechanism	Disease	Reference Nos.
<i>General Mg^{2+} transporters</i>						
TRPM7	Plasma membrane	Ubiquitous	Ba>Ni>Mg>Ca	Channel		314, 349
MagT1	Plasma membrane	Ubiquitous	Mg>Ba>Fe=Cu	Channel	X-MEN syndrome	187, 311
SLC41A1	Plasma membrane	Ubiquitous	Mg>Sr>Fe>Ba>Cu	Exchanger	Nephronophthisis-like	185, 251, 289
SLC41A2	Golgi membrane	Ubiquitous	Mg>Ba>Ni>Ca	Exchanger		442
CNNM3	Plasma membrane	Ubiquitous	Mg>Fe>Cu>Co	Transporter?		545
MRS2	Mitochondrial membrane	Ubiquitous	Mg>Ni	Channel		399
<i>Tissue-specific Mg^{2+} transporters</i>						
TRPM6	Apical plasma membrane	Kidney, intestine	Ba>Ni>Mg>Ca	Channel	Hypomagnesemia secondary hypocalcemia	314
CNNM1	?	Brain	Cu>Mg?	?		13, 545
CNNM2	Basolateral plasma membrane	Kidney	Mg>Sr>Zn>Cd	Transporter? Sensor?	Hypomagnesemia with seizures and mental retardation	184, 497
CNNM4	Basolateral plasma membrane	Intestine	Mg	Exchanger?	Jallili syndrome	387, 402, 575

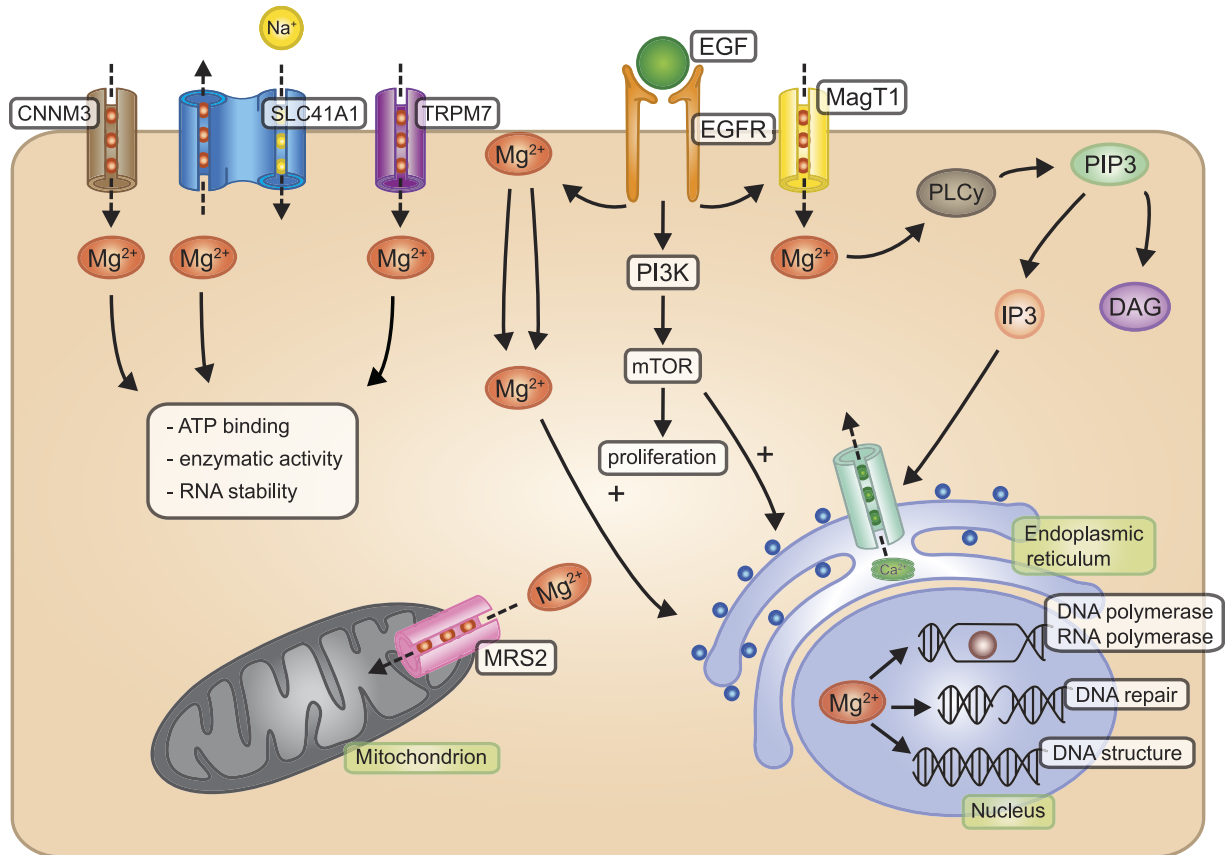


FIGURE 1. Magnesium in cellular physiology. Cellular Mg^{2+} homeostasis is regulated by the combined action of TRPM7, SLC41A1, MagT1, and CNNM3 Mg^{2+} transporters. MRS2 transporters regulate intramitochondrial Mg^{2+} concentrations. In the nucleus, Mg^{2+} is involved in DNA stability and DNA repair and regulates the activity of the DNA and RNA polymerases. Within the cell cytosol, Mg^{2+} regulates ATP binding, enzymatic activity of more than 600 enzymes, proliferation, and tRNA and mRNA stability. Activation of growth factor receptors, such as the EGFR, will increase Mg^{2+} uptake and release of membrane-bound Mg^{2+} resulting in mTOR activation and Ca^{2+} release from the ER. These mechanisms are essential for cell growth and proliferation. TRPM7, transient receptor potential melastatin type 7; CNNM3, cyclin M3; SLC41A1, solute carrier family 41 type 1; MagT1, magnesium transporter 1; MRS2, mitochondrial RNA splicing 2; EGF, epidermal growth factor; EGFR, epidermal growth factor receptor; PI3K, phosphoinositide 3-kinase; mTOR, mammalian target of rapamycin; PLC γ , phospholipase C- γ ; PIP $_3$, phosphatidylinositol 3,4,5-trisphosphate; IP $_3$, inositol trisphosphate.

searchers have identified several proteins critical to cellular Mg^{2+} homeostasis. Within this part of the review, we will focus on the ubiquitous transporters transient receptor potential melastatin type 7 (TRPM7), Mg^{2+} transporter 1 (MagT1), and solute carrier family 41 member 1 (SLC41A1). Tissue-specific Mg^{2+} transporters such as transient receptor potential melastatin type 6 (TRPM6; kidney, colon), cyclin M2 (CNNM2; kidney) and cyclin M4 (CNNM4; colon) are discussed in section III.

1. TRPM7

TRPM7 is a ubiquitously expressed divalent cation channel that is responsible for much of the Mg^{2+} flux in the cell. TRPM7 activity is generally regarded as a prerequisite for cell viability (360, 455). However, recent reports with tissue-specific TRPM7 KO mice suggest that TRPM7-deficient T cells are still viable and have normal intracellular

Mg^{2+} concentrations (266). TRPM7 constitutes a tetrameric channel, where each subunit consists of six transmembrane regions with a pore region between the fifth and sixth transmembrane domain (42). The intracellular COOH terminus contains a kinase domain that regulates autophosphorylation of the channel, although its mechanism is poorly understood, as TRPM7 channel function is not dependent on its kinase activity (504). Initially, the kinase was reported to exist as separate entity (438), and indeed, recently it was shown that the kinase is cleaved from TRPM7 by caspase-8, although the exact function of the cleaved kinase remains unknown (115).

2. MagT1

Originally identified in MDCT cells, Mg^{2+} transporter 1 (MagT1) has been described as a ubiquitously expressed Mg^{2+} channel (187). Survival and growth of TRPM7-defi-

cient cells can be partially rescued by MagT1 overexpression (110). Although identification of MagT1 dates back almost 10 years, the functional characteristics of MagT1 are still undetermined. MagT1 mediates highly specific Mg^{2+} currents in *Xenopus laevis* oocytes, but these results could not be reproduced in mammalian cells (187, 589). Recent studies in T cells suggest that MagT1 mediates a rapid Mg^{2+} influx upon receptor activation (79, 311). Since T cells do not require TRPM7 for maintaining normal intracellular Mg^{2+} concentrations (266), this suggests that MagT1 has a similar function as TRPM7 in certain cell types.

3. Extrusion

In 1984, Theodor Gunther et al. (214) proposed that the main route of Mg^{2+} efflux from the cell is Na^+ dependent. A large body of evidence obtained in a wide range of cell types supports this notion (reviewed in Ref. 424). Over the last decades, the mechanism has been further characterized in a variety of cell types, demonstrating inhibition by Na^+ channel blockers such as amiloride, imipramine, and quinidine. The stoichiometry of this exchange mechanism is still not fully elucidated; Na^+ -dependent Mg^{2+} extrusion is activated by cAMP in several cell models and conditions (424), but Na^+ -independent Mg^{2+} extrusion has also been proposed. Ebel et al. (134) reported the presence of a choline-dependent Mg^{2+} transporter in erythrocytes. However, the molecular identity of this proposed Mg^{2+} efflux mechanism remains controversial.

4. SLC41A1

Recent reports by Kolisek and co-workers (288, 289) suggested that solute carrier family 41 member 1 (SLC41A1) functions as a Na^+/Mg^{2+} exchanger with a 2:1 stoichiometry. SLC41A1 contains 11 transmembrane domains and was originally described as a Mg^{2+} transporter mediating Mg^{2+} currents in *Xenopus laevis* oocytes (185). Although electrophysiological analysis could not confirm these measurements in mammalian cells, Mg^{2+} efflux studies using mag-fura 2 show Na^+ -dependent Mg^{2+} extrusion (251, 289). Gain-of-function SNPs have been associated with Parkinson's disease, and one mutation in *SLC41A1* was identified in a patient with a nephronophthisis-like phenotype (251, 290, 576). SLC41A1 is part of a larger protein family including two additional members, SLC41A2 and SLC41A3, which are studied less extensively. Although SLC41A2 was initially described as a plasma membrane protein, it has a topology opposite to SLC41A1 (442). This finding suggests that SLC41A2 might be expressed on the membranes of organelles and may be involved in subcellular Mg^{2+} transport.

5. CNNM3

Members of the Cyclin M (CNNM) family have been proposed to function as Mg^{2+} transporters (184, 545).

CNNM1 is mainly expressed in brain, CNNM2 expression is high in kidney, and CNNM4 is primarily expressed in intestine (105). In contrast, CNNM3 has a ubiquitous expression pattern and may play a role in the maintenance of cellular Mg^{2+} homeostasis. A recent study shows that CNNM3 transports Mg^{2+} , and its activity is regulated by oncogene PRL2 (217). The interaction between PRL2 and CNNM3 is essential for Mg^{2+} influx that drives tumor growth. Therefore, CNNM3 should be considered in future studies on cellular Mg^{2+} handling in nonpathological conditions.

6. MRS2

Although most studied in yeast, MRS2 (mitochondrial RNA splicing 2) is considered to be the primary Mg^{2+} channel on the mitochondrial membrane (592). Knockdown of MRS2 results in reduced Mg^{2+} uptake in mitochondria and cell death (399). Using the newly developed mitochondrial Mg^{2+} fluorescent probe KMG-301, Shindo et al. (475) revealed that MRS2 regulates intramitochondrial Mg^{2+} concentrations. This finding is interesting, since it indicates that mitochondria may store intracellular Mg^{2+} . Given that Mg^{2+} is of major importance for ATP binding, intramitochondrial Mg^{2+} concentrations may indirectly influence the progression of the citric acid cycle. Recently, it was shown that MRS2 mutations cause demyelination. The relevance of this observation to Mg^{2+} homeostasis still remains to be determined (296).

7. Others

In addition to the aforementioned Mg^{2+} transporters, several other proteins have been proposed to transport Mg^{2+} . However, these claims are based mainly on overexpression in the *Xenopus oocytes* model, and functional evidence for these proteins is scarce. For example, the nonimprinted in Prader-Willi/Angelman syndrome (NIPA) family of proteins has been proposed to transport Mg^{2+} , based on Mg^{2+} currents in *Xenopus laevis* oocytes (182), but recent studies indicate that NIPA proteins have a role in bone morphogenetic protein (BMP) signaling (525). Likewise, Huntingtin-interacting protein 14 (HIP14) was thought to mediate Mg^{2+} fluxes at the Golgi membrane (183). Now it has become apparent that its main function consists of palmitoyl acyltransferase activity, specifically involved in the palmitoylation of Huntingtin (129, 582). Therefore, the role of NIPA proteins and HIP14 in Mg^{2+} transport should be questioned. Additionally, members of the membrane Mg^{2+} transporter (MMgT) family have been shown to transport divalent cations in *Xenopus oocytes* (186). However, as they have only one transmembrane domain after signal peptide cleavage, it is unlikely that they form functional Mg^{2+} transporters themselves. It is possible that MMgT proteins may form subunits of other Mg^{2+} channels, and as a consequence, future studies should be directed to the identification of its protein partners.

D. Cell Signaling

Mg^{2+} acts as a physiological Ca^{2+} antagonist within cells, and as a result, the Mg^{2+}/Ca^{2+} ratio is of major importance for the activity of Ca^{2+} -ATPases and other Ca^{2+} transporting proteins (257). Small changes in the Mg^{2+} availability within the cell may therefore cause disturbed Ca^{2+} signaling or Ca^{2+} toxicity.

Since 1974, when Mg^{2+} influx was detected upon insulin stimulation, several groups have suggested a second messenger role for Mg^{2+} (311, 322, 503). Most recently in a study on T-cell activation, MagT1 channels were shown to mediate Mg^{2+} influx upon T-cell receptor activation and EGF stimulation (311). In these T cells, Mg^{2+} activates phospholipase C- γ (PLC γ 1), resulting in reduced phosphorylation of protein kinase C (PKC) and inositol trisphosphate (IP_3) generation downstream, eventually leading to reduced Ca^{2+} influx (FIGURE 1). In contrast, other reports suggest that PLC γ activation precedes Mg^{2+} influx (240, 241). The proposition of Mg^{2+} as a dynamic second messenger raises many questions. How do MagT1 or other Mg^{2+} transporters facilitate rapid Mg^{2+} influx when the intracellular and extracellular Mg^{2+} concentrations are almost equal? What mechanism is involved in managing Mg^{2+} after the initial influx, given the absence of Mg^{2+} pumps and major Mg^{2+} binding proteins? Follow-up studies demonstrated that MagT1-deficient cells have severely reduced basal intracellular Mg^{2+} concentrations (79). These results suggest that the effects seen on PLC γ 1 are dependent on general intracellular Mg^{2+} availability and further question the physiological role of variable fluxes that are proposed in the second messenger theory. Studying Mg^{2+} dynamics within the cell using fluorescent probes may help to draw definitive conclusions on this matter.

E. Cell Proliferation

Given its effect on RNA, DNA, and protein synthesis, Mg^{2+} is an important factor in the control of cell proliferation. Over the last 40 years, the role of Mg^{2+} in cell cycle control, protein synthesis, and growth factor response has been extensively studied, pioneered by several groundbreaking studies from the group of Harry Rubin (428, 430). Cell proliferation is largely dependent on protein synthesis, more than DNA or RNA synthesis. Inhibition of protein synthesis directly shuts down DNA synthesis, whereas there is a 2-h delay to achieve the same effect using RNA synthesis inhibitors (279). Protein synthesis is highly dependent on intracellular Mg^{2+} concentrations; increasing the Mg^{2+} content amplifies protein synthesis within 60 min, whereas DNA synthesis is only enhanced after 10 h (429, 514). Activation of proliferation is initiated by growth factors that increase glucose uptake and protein synthesis within minutes (225). Interestingly, Mg^{2+} is tightly regulated during these intracellular processes. Initial studies in cultured

cells showed that applying insulin induced 20% higher intracellular Mg^{2+} concentrations after 16 h (445). Later studies with EGF using the fluorescent probe mag-fura 2 showed an impressive fourfold increase of intracellular Mg^{2+} from 0.3 to 1.4 mM after 20 min of epidermal growth factor (EGF) stimulation (199). The authors state that a rise in Mg^{2+} precedes DNA synthesis, but coincides with and thus may contribute to increases in protein synthesis. Recent studies identifying the molecular mediators of Mg^{2+} -dependent cell proliferation have resulted in the membrane, magnesium, mitosis (MMM) model (432). The MMM model proposes that, upon growth factor binding, Mg^{2+} enters the cell or is released from phospholipids in the cell membranes (FIGURE 1). Increased cytosolic Mg^{2+} levels contribute to ribosomal activity and protein synthesis, eventually leading to DNA replication and mitosis. The mammalian target of rapamycin (mTOR) complex is a critical component of the MMM model, as it is the master regulator of cell cycle progression and proliferation (551). Growth factors binding to their receptors leads to phosphoinositide 3-kinase (PI3K) phosphorylation, which activates the mTOR complex (431, 548). Activation of mTOR is $MgATP^{2-}$ dependent, and ATP has been suggested as the main regulator of mTOR activity. However, ATP levels do not change upon growth factor stimulation, whereas Mg^{2+} levels do (445, 537). Therefore, the MMM model proposes Mg^{2+} as the primary regulator of mTOR dynamics and cell proliferation.

1. Cancer

Tumor cells contain high concentrations of intracellular Mg^{2+} (508). In a mammary tumor cell line, Mg^{2+} can be transported into the cell even when extracellular Mg^{2+} concentrations were below physiological levels (203, 264, 566). Mg^{2+} uptake via divalent cation channel TRPM7 has been suggested to stimulate tumor cell proliferation (203, 264). TRPM7 expression is upregulated in hepatoma, pancreatic adenocarcinoma, gastric cancer, and breast cancer tissue (203, 278, 346, 581). Although TRPM7 has been primarily described as a Mg^{2+} channel, it is also permeable for other divalent cations (349). Given the involvement of Mg^{2+} in cell proliferation, the influx of Mg^{2+} via TRPM7 has been proposed as the main regulator of tumor growth. However, recent studies using prostate cancer cells suggest that TRPM7-mediated Ca^{2+} uptake may also play an important role in tumor growth (501). The expression of Mg^{2+} transporter CNNM3 is increased in human breast cancer tissue (217). CNNM3 binds oncogene PRL2 and facilitates the entry of Mg^{2+} in the tumor cell to drive cell proliferation. Elevated intracellular Mg^{2+} concentrations have been suggested to be beneficial for tumor growth because Mg^{2+} regulates several cancer-associated enzymes including telomerase and protein phosphatase 1D, which are involved in the glycolytic cycle and BER (74). However, the regulatory role of Mg^{2+} on these enzymes during the pathogenic state of tumor cell proliferation has never been inves-

tigated and, therefore, the exact role of Mg^{2+} in enzymatic regulation in cancer remains speculative.

In contrast to the proliferative phase of tumor growth, in which tumor cells have high intracellular Mg^{2+} concentrations, low intracellular Mg^{2+} concentrations are associated with increased rates of carcinogenesis and metastasis (74). Low Mg^{2+} conditions and impaired activity of DNA repair mechanisms reduces DNA protection against oxidative stress. Indeed, low dietary Mg^{2+} intake has been associated with the risk of several types of cancers. Epidemiological studies have established a correlation between low Mg^{2+} intake and colon cancer risk (160, 302, 534). In addition, in a study with 1,200 lung cancer patients and a similar number of controls, low dietary Mg^{2+} intake was associated with reduced lung cancer risk (326). However, these results could not be reproduced in other patient cohorts (325, 502).

III. REGULATION OF MAGNESIUM HOMEOSTASIS

Mg^{2+} serum concentrations range between 0.7 and 1.1 mM in healthy people (323). To maintain constant plasma Mg^{2+} levels, the United States Food and Nutrition Board recommends a daily Mg^{2+} intake of 420 mg for men and

320 mg for women (1). Mg^{2+} homeostasis depends on the collaborative actions of the intestine, responsible for Mg^{2+} uptake from food, the bone, which stores Mg^{2+} in its hydroxy-apatite form, and the kidneys, regulating urinary Mg^{2+} excretion (FIGURE 2).

A. Magnesium in Intestine

Given a daily Mg^{2+} intake of 370 mg, ~30–50% is absorbed in the intestine, resulting in a net uptake of ± 100 mg. However, if Mg^{2+} intake is low, early reports suggest that up to 80% of dietary Mg^{2+} can be absorbed (189). Mg^{2+} absorption in the gut depends on two separate pathways; paracellular transport is responsible for bulk Mg^{2+} absorption and takes place mostly in the small intestine, whereas fine-tuning occurs in the cecum and colon via transcellular transport (FIGURE 3). In spite of this, the intestine seems to have a limited role in regulation of the Mg^{2+} balance. In contrast to other minerals, intestinal Mg^{2+} absorption is poorly regulated and depends mainly on Mg^{2+} intake (216, 461). Thus the kidneys presumably primarily regulate the maintenance of Mg^{2+} homeostasis.

1. Small intestine

Mg^{2+} absorption in the small intestine is hypothesized to be exclusively of a paracellular nature, since Mg^{2+} absorption

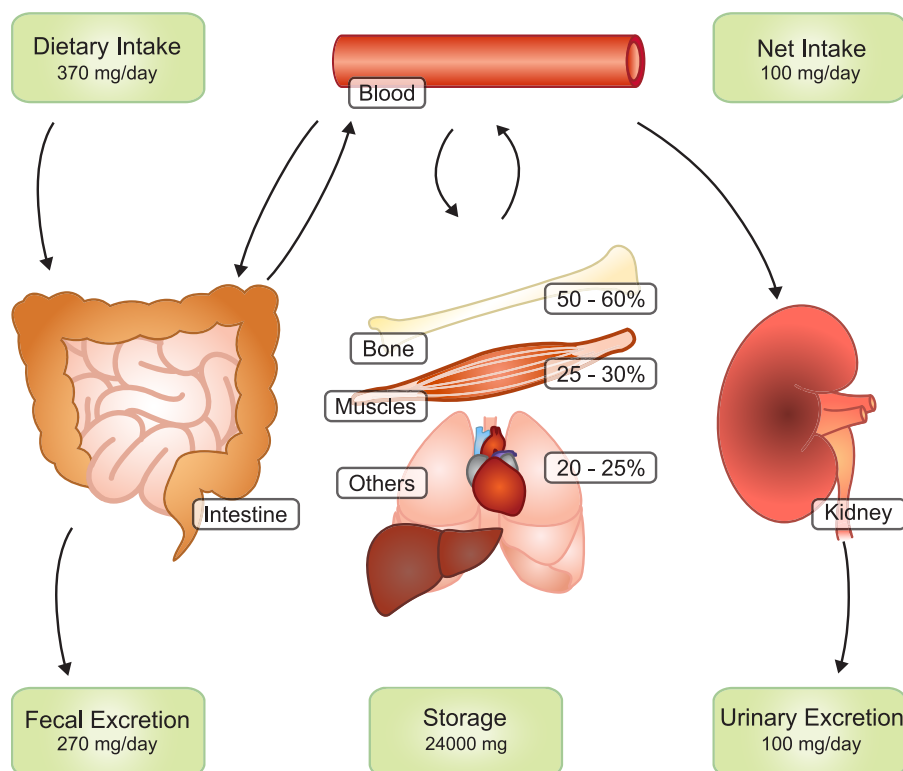


FIGURE 2. Magnesium homeostasis. Panels represent the daily amount of Mg^{2+} intake and excretion. Daily the intestines absorb ~120 mg and secrete 20 mg of Mg^{2+} , resulting in a net absorption of 100 mg. In the kidney daily ~2,400 mg Mg^{2+} is filtered by the glomerulus, of which 2,300 mg is reabsorbed along the kidney tubule. This results in a net excretion of 100 mg, which matches the intestinal absorption. Bone and muscle provide the most important Mg^{2+} stores.

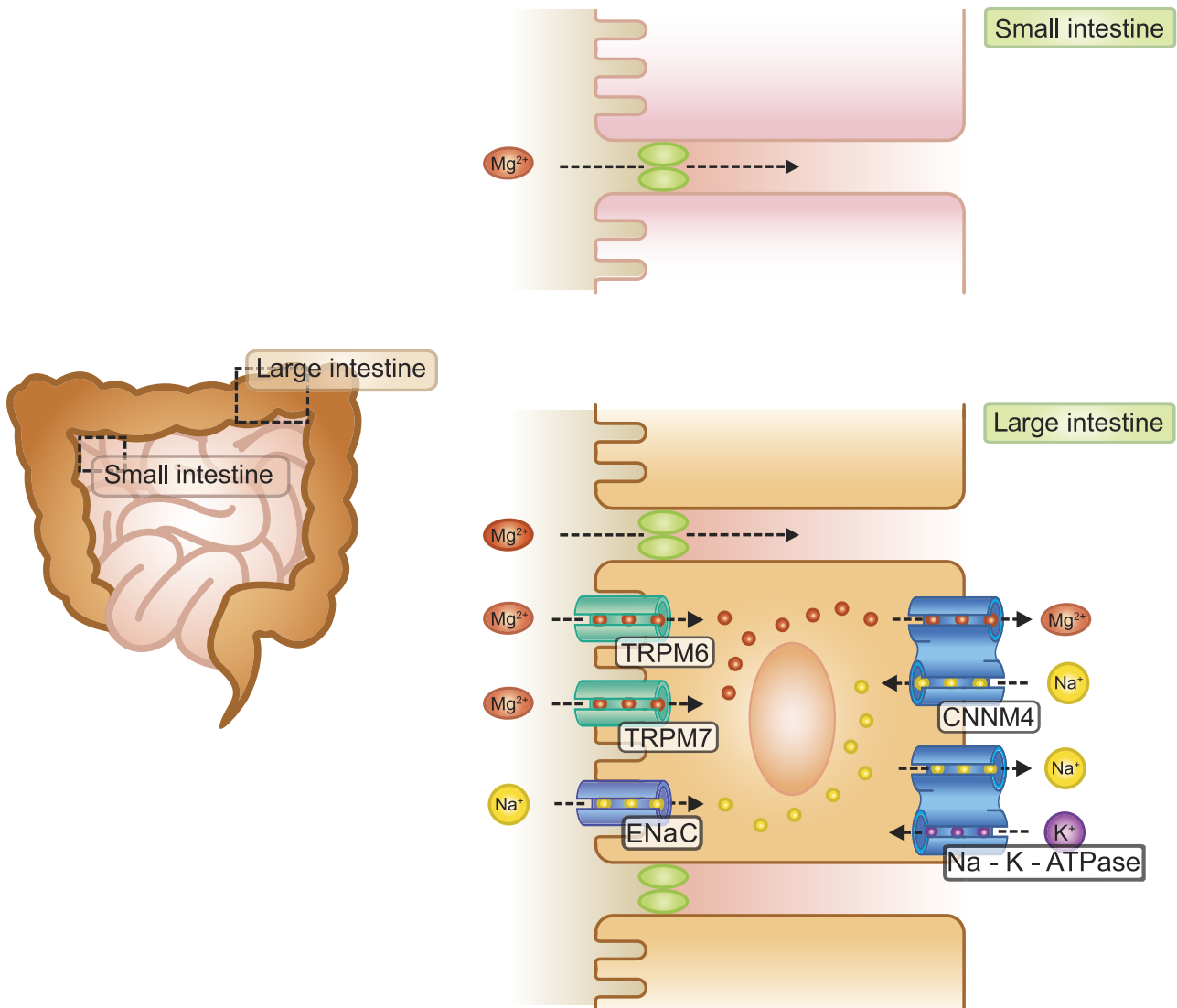


FIGURE 3. Magnesium absorption in the intestine. Bulk Mg^{2+} is absorbed paracellularly by the late part of the small intestine. Fine-tuning of Mg^{2+} absorption takes place transcellularly by the colon, where TRPM6 and TRPM7 Mg^{2+} channels facilitate luminal Mg^{2+} uptake in the enterocyte. CNNM4 provides the basolateral Mg^{2+} extrusion mechanism. TRPM6, transient receptor potential melastatin type 6; TRPM7, transient receptor potential melastatin type 7; ENaC, epithelial sodium channel; CNNM4, cyclin M4.

in this region of the intestine correlates linearly to luminal Mg^{2+} concentrations (273, 409). Moreover, the epithelial Mg^{2+} channel TRPM6 is not expressed in the small intestine (196). Mg^{2+} is poorly absorbed in the duodenum, where unfavorable electrochemical gradients may even result in a limited amount of paracellular Mg^{2+} excretion (389). In more distal parts of the small intestine, such as late jejunum and ileum, the driving force for passive Mg^{2+} transport is established by the high luminal Mg^{2+} concentration and the lumen-positive transepithelial voltage of ~ 15 mV (164). The K_m for Mg^{2+} transport in the distal small intestine has been reported to be in the range of 4–12 mM (343, 461). These results suggest that NaCl and water absorption are prerequisites for Mg^{2+} uptake, since water absorption concentrates luminal Mg^{2+} . Tight junction permeability underlying paracellular Mg^{2+} transport is still

poorly understood. The small intestine is described as the most ion-permeable part of the intestine because of the relatively low expression of “tightening” claudins 1, 3, 4, 5, and 8 (20, 301). Claudins 16 and 19, which are linked to Mg^{2+} transport, are not expressed in the intestine (20, 246). The exact composition of the tight junction complex facilitating intestinal Mg^{2+} absorption remains to be elucidated.

2. Large intestine

Mg^{2+} absorption in cecum and colon is thought to be transcellular of nature and is mediated by TRPM6 and TRPM7 on the luminal side of the enterocyte (FIGURE 3). Intestinal expression of TRPM6 is located in cecum and colon (196, 300). In a study with rat colon epithelium, 37% of Mg^{2+} was transported transcellularly (272). This suggests signif-

icant paracellular transport of Mg^{2+} in the colon, which would be unlikely given the expression of tightening claudins 3, 4, and 8 in this segment (301). In contrast to Ca^{2+} , Mg^{2+} transport in colon is independent of 1,25-dihydroxyvitamin D_3 [$1,25(OH)_2D_3$] signaling, nor is *TRPM6* expression dependent on $1,25(OH)_2D_3$ (196, 272). It has been suggested that the basolateral Mg^{2+} extrusion mechanism of the enterocyte is coupled to the Na^+ gradient (424). Indeed, the results of a recent study using *CNNM4* KO mice suggest that *CNNM4* may act as a Na^+/Mg^{2+} exchanger at the basolateral membrane of enterocytes (575). *CNNM4* KO mice suffer from hypomagnesemia, and functional analysis using Magnesium Green showed that *CNNM4* overexpression increased Mg^{2+} efflux in HEK293 cells. However, patients with *CNNM4* mutations do not suffer from hypomagnesemia (387, 402).

B. Magnesium in Bone

Approximately 50–60% of the total body Mg^{2+} content is stored in bone. Serum Mg^{2+} concentrations are closely related to bone metabolism; bone surface Mg^{2+} is continuously exchanged with blood Mg^{2+} (14). In bone, Mg^{2+} ions bind at the surface of the hydroxyapatite crystals. Mg^{2+} increases the solubility of P_i and Ca^{2+} hydroxyapatite and thereby acts on the crystal size and formation (443). Mg^{2+} induces osteoblast proliferation; therefore, Mg^{2+} deficiency results in decreased bone formation (320) (FIGURE 4). Mg^{2+} -deficient rats have reduced osteoblast numbers and decreased bone mass (434). Additionally, Mg^{2+} deficiency increases the secretion of proinflammatory cytokines such as tumor necrosis factor (TNF)- α , interleukin (IL)-1 β , and substance P (434, 552), all of which have been implicated in increased osteoclastic bone resorption (280). These effects may be further enhanced by reduced parathyroid hormone (PTH) and $1,25(OH)_2D_3$ levels, which are often associated with hypomagnesemia (435). Interestingly, Mg^{2+} -deficient rats have reduced chondrocyte column formation, which is associated with reduced *SRY* (sex determining region Y)-box 9 (*SOX9*) expression (200) (FIGURE 4). *SOX9* is a key transcription factor in chondrogenesis. In a recent gene ex-

pression study in the Mg^{2+} -transporting segment of the kidney, *SOX9* mRNA was the most increased in the low- Mg^{2+} diet group (104). These findings suggest that *SOX9* is an important transcription factor for bone and kidney Mg^{2+} homeostasis.

C. Magnesium in Kidney

Approximately 2,400 mg of Mg^{2+} is filtered by the glomeruli on a daily basis. The nephron recovers 95–99% of this; the remaining 100 mg leaves the body via the urine (FIGURE 2).

1. Proximal tubule

The mechanisms of proximal tubule (PT) Mg^{2+} reabsorption are poorly understood, but early micropuncture studies showed that ~10–25% of Mg^{2+} is reabsorbed by the proximal convoluted tubule segment of the nephron (304, 411). In the glomeruli, 70% of the serum Mg^{2+} is freely filterable, suggesting that the concentration in the glomerular filtrates and thus at the start of the PT ranges between 0.5–0.7 mM. The transepithelial potential difference ranges from slightly lumen negative (–6 mV) in the early parts of the PT to positive (3 mV) in later parts (287). Micropuncture studies have shown that a 1.9 ratio between the concentrations of Mg^{2+} in the tubular fluid and the interstitial fluid is necessary to initiate Mg^{2+} transport (304). This finding could be explained by the poor tight junction permeability for Mg^{2+} in PT. As a result, water uptake via aquaporin 1 (AQP1) precedes Mg^{2+} reabsorption (410) (FIGURE 5). Consequently, Mg^{2+} reabsorption mainly occurs in the late parts of the PT, where the transepithelial chemical Mg^{2+} gradient is sufficient to favor Mg^{2+} transport. PT Mg^{2+} reabsorption is generally considered to be a passive paracellular process, but there might be some transcellular Mg^{2+} transport via a poorly characterized amiloride-sensitive mechanism (254). In both cases, sufficient Na^+ transport is required to drive water transport that is a prerequisite for Mg^{2+} reabsorption. Hormonal effects on Na^+ reabsorption in the PT will therefore also affect Mg^{2+} reabsorption in this segment. However, disturbances of proximal tubular

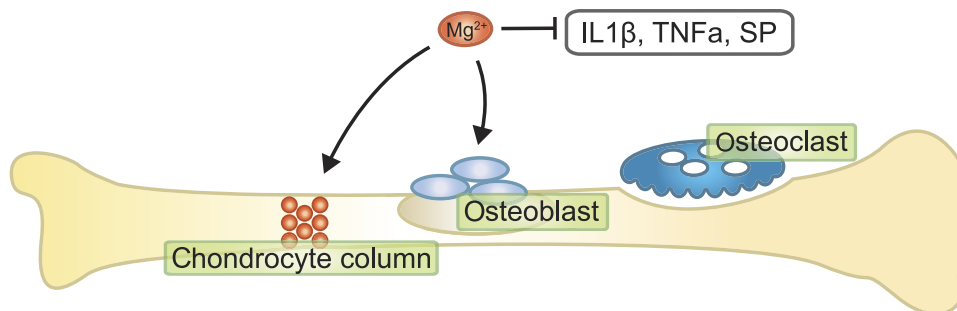


FIGURE 4. Magnesium storage in bone. Mg^{2+} stimulates osteoblast proliferation in bone and inhibits the release of proinflammatory molecules such as IL-1 β , TNF- α , and SP, which stimulate osteoclast activity. In bone development, Mg^{2+} stimulates chondrocyte column formation. IL-1 β , interleukin-1 β ; TNF- α , tumor necrosis factor- α ; SP, substance P.

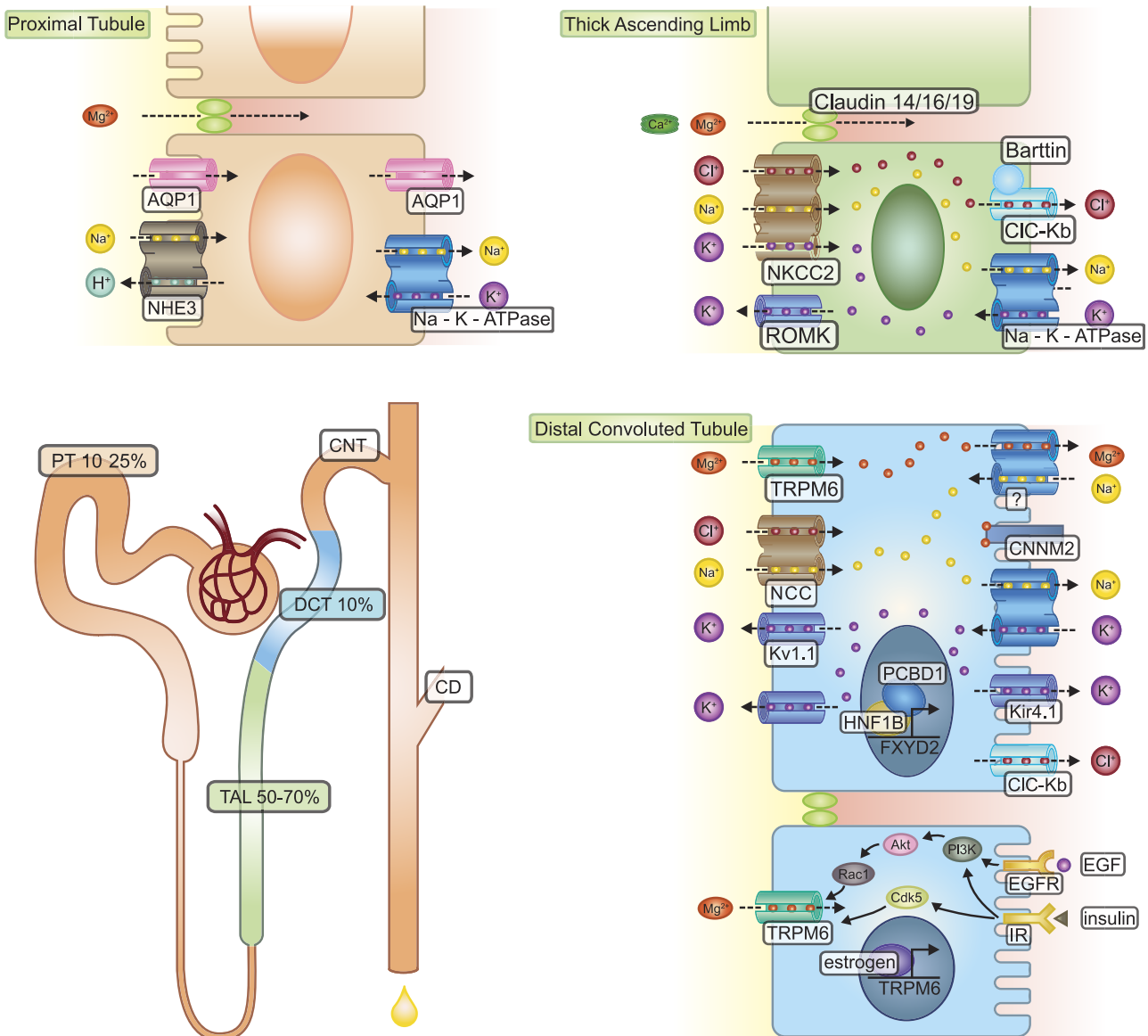


FIGURE 5. Magnesium reabsorption in the kidney. The glomerulus filters the blood, and along the nephron 95% is reabsorbed. In the late proximal tubule (PT), Na^+ and H_2O reabsorption via NHE3 and AQP1 are prerequisites for paracellular Mg^{2+} transport. Approximately 10–25% of Mg^{2+} is reabsorbed in the proximal tubule. Bulk Mg^{2+} reabsorption (50–70%) takes place in the thick ascending limb of Henle’s Loop (TAL). In TAL, Mg^{2+} reabsorption take place paracellular and depends on the uptake of Na^+ and K^+ via NKCC2. Fine-tuning (10%) of Mg^{2+} transport takes place transcellular in the distal convoluted tubule (DCT). In DCT, TRPM6 facilitates Mg^{2+} uptake from the pro-urine, which depends on the voltage gradient set by backleak of K^+ via ROMK and $\text{K}_v1.1$ potassium channels. At the basolateral membrane, Mg^{2+} is extruded via an unknown mechanism, which may be regulated by CNNM2 acting as Mg^{2+} sensor. Mg^{2+} extrusion depends on the Na^+ gradient, set by the $\text{Na}^+\text{-K}^+\text{-ATPase}$ is in turn dependent on K^+ recycling via Kir4.1. FXYP2 transcription encoding the γ -subunit of the $\text{Na}^+\text{-K}^+\text{-ATPase}$ is regulated by HNF1 β and PCBD1. Regulation of Mg^{2+} transport in DCT depends on EGF and insulin. Upon activation of the EGFR and IR, an intracellular signaling cascade including PI3K, Akt, and Rac1 results in an increased TRPM6 membrane expression and increased channel activity. Additionally, estrogens have been shown to increase TRPM6 expression. PT, proximal tubule; TAL, thick ascending limb of Henle’s loop; DCT, distal convoluted tubule; CNT, connecting tubule; CD, collecting duct; NHE3, $\text{Na}^+\text{-H}^+$ exchanger type 3; AQP1, aquaporin 1; NKCC2, $\text{Na}^+\text{-K}^+\text{-2Cl}^-$ cotransporter; ROMK, renal outer medulla K^+ channel; ClC-Kb, chloride channel Kb; $\text{Kv}1.1$, voltage-gated K^+ channel 1.1; TRPM6, transient receptor potential melastatin type 6; NCC, $\text{Na}^+\text{-Cl}^-$ cotransporter; CNNM2, cyclin M2; FXYP2, FXYP-domain containing 2; HNF1 β , hepatocyte nuclear factor 1 β ; PCBD1, pterin-4 alpha-carbinolamine dehydratase 1; EGF, epidermal growth factor; EGFR, epidermal growth factor receptor; IR, insulin receptor; PI3K, phosphoinositide 3-kinase; Rac1, Ras-related C3 botulinum toxin substrate 1; Cdk5, cyclin-dependent kinase 5.

Mg²⁺ reabsorption generally do not result in clinical symptoms, since more distal segments will compensate for reduced Mg²⁺ uptake in PT.

2. Thick ascending limb of Henle's loop

Whereas most electrolytes are majorly transported in the PT, the thick ascending limb of Henle's loop (TAL) is the main location for Mg²⁺ reabsorption (236, 299, 420). Due to the unique properties of this segment, ~50–70% of filtered Mg²⁺ is reabsorbed here. Most of the Mg²⁺ is reabsorbed by the cortical part of the TAL, since medullary Mg²⁺ reabsorption is negligible (468). Paracellular bulk Mg²⁺ transport is dependent on the lumen-positive transepithelial voltage (+10 mV) that is determined by the activity of the Na⁺-K⁺-2Cl⁻ cotransporter (NKCC2) and the subsequent secretion of K⁺ at the apical membrane (192). Inhibition of NKCC2 by furosemide diuretics therefore decreases TAL Mg²⁺ reabsorption (see sect. VB3). Further contributors to the transepithelial membrane voltage are K⁺ secretion via renal outer medullary potassium channel (ROMK) and paracellular backflux of Na⁺ ions as a result of decreasing luminal Na⁺ concentrations (329).

The reabsorption of Mg²⁺ in the TAL follows the paracellular pathway and therefore depends on the tight junction permeability (FIGURE 5). Tight junctions form a physical and chemical barrier between the epithelial cells. Their major components are proteins of the claudin family. Currently, 26 claudins have been described in humans (209). Tight junction permeability is determined by the individual claudins in each tight junction complex. TAL tubuli are known to express claudins 3, 10, 11, 14, 16, and 19. Claudins 16 and 19 are considered to be the main claudins influencing Mg²⁺ permeability, since mutations in these proteins result in renal Mg²⁺ wasting (293) (FIGURE 5).

However, the role of claudin 16 in Mg²⁺ reabsorption is controversial. Claudin 16 was initially considered to act as a paracellular Mg²⁺ channel, but this hypothesis has not been unequivocally confirmed (207, 245). Reports using the claudin 16 knockdown (KD) mouse model and the LLC-PK₁ cell model suggest that claudin 16 increases Na⁺ permeability (227, 245, 466). This would imply that claudin 16 is mainly involved in the regulation of the transepithelial voltage gradient by controlling the paracellular Na⁺ back-leak. In MDCK-C7 cells overexpressing claudin 16, Na⁺ permeability yet remained stable, whereas Mg²⁺ permeability increased significantly (207). Claudin 16 KD mice demonstrate a twofold lower permeability ratio for Na⁺ over Cl⁻ without a change in paracellular conductance. Consequently, the transepithelial voltage collapsed, reducing the driving force for Mg²⁺ reabsorption in TAL (466).

Claudin 19 has been studied less extensively, but has been suggested to increase the tight junction barrier function (24). The claudin 19 KD mouse exhibits highly increased

urinary excretion of K⁺, Mg²⁺, and Ca²⁺, but Mg²⁺ is the only electrolyte altered at the serum level (246). The discrepancy between studies with claudin 16 and claudin 19 isoforms might be explained by their interdependence in forming functional tight junction barriers (246, 247). Both in vitro and in vivo studies demonstrated that claudins 16 and 19 need to interact for proper insertion in the tight junction to become functionally active. Further differences in experimental results may depend on the endogenous expression of other claudin isoforms in the specific cell types used in these experiments.

Claudin 14 reduces the cation specificity of tight junction barriers, when coexpressed with claudin 16, or with claudin 16-claudin 19 complexes (180). Consequently, claudin 14 KO mice exhibit increased serum Mg²⁺ values and decreased urinary Mg²⁺ excretion. This agrees with previous findings in MDCK cells showing that claudin 14 acts as a nonspecific cation blocker (45, 560). Studies on claudin 14 KO mice have mainly focused on Ca²⁺ homeostasis, since claudin 14 expression is highly Ca²⁺ sensitive (121, 180). The CaSR regulates claudin 14 expression and Ca²⁺ reabsorption in the TAL by downregulation of two microRNAs, miR-9 and miR-374. Since CaSR is also activated by Mg²⁺, although to a lesser extent than Ca²⁺, it would be interesting to address the effect of elevated serum Mg²⁺ levels on claudin 14 expression in future studies.

Recently, claudin 10 has been identified as an important factor in cation selectivity in TAL, as demonstrated in a mouse model where claudin 10 was deleted specifically in this segment (57). Claudin 10 TAL-KO mice show hypermagnesemia, nephrocalcinosis, and impaired paracellular Na⁺ permeability. In the absence of claudin 10, TAL tight junctions became more permeable to Ca²⁺ and Mg²⁺ and the transepithelial voltage increased. These results are in line with in vitro studies overexpressing claudin 10b, suggesting that this splice variant is mainly expressed in TAL (57, 208).

3. Distal convoluted tubule

The distal convoluted tubule (DCT) determines the final urinary Mg²⁺ concentration, since no reabsorption of Mg²⁺ takes place beyond this segment. Approximately 10% of the total Mg²⁺ is reabsorbed by tightly regulated transcellular transport mechanisms (64). DCT cells form a high-resistance epithelium with a lumen-negative voltage of approximately -5 mV (192, 571). In DCT, TRPM6 divalent cation channels mediate luminal Mg²⁺ uptake (234, 235) (FIGURE 5). Within the kidney, TRPM6 is specifically expressed in DCT, and its activity is regulated by intracellular Mg²⁺ (540). TRPM6 contains six transmembrane spanning domains with a pore region between the fifth and sixth segment and a large kinase domain fused to the channel's intracellular COOH terminus. TRPM6 may function in homo- and heteromeric tetramers with TRPM7, al-

though there is some controversy about the necessity of TRPM7 for TRPM6 function (314, 588).

TRPM6 is regulated by numerous factors at the level of transcription, plasma membrane availability, and activity (69). EGF and insulin act on TRPM6 by a PI3K-Akt-Rac1 dependent mechanism, increasing the insertion of TRPM6 in the membrane (361, 515) (FIGURE 5). Insulin may directly affect TRPM6 activity through cyclin-dependent kinase 5 (cdk5)-dependent phosphorylation of the channel. Patients with reduced EGFR or insulin receptor (IR) activity are therefore more susceptible to hypomagnesemia (361, 459). Additionally, estrogens increase *TRPM6* mRNA expression (196). Over the last decade, several important interactors of TRPM6, including receptor for activated C-kinase 1 (RACK1) and prohibitin2 (PHB2/REA), have been identified (70, 71). RACK1 interacts with the α -kinase domain of TRPM6 in the autophosphorylated state, thereby reducing TRPM6 activity (70). Other modulators of TRPM6 activity include dietary Mg^{2+} , pH, and ATP (516). Interestingly, acidification-induced current potentiation is dependent on residues p.Glu1024 and p.Glu1029, which also determine the pore selectivity for Mg^{2+} (313, 367). Moreover, recent findings indicate that TRPM6 is inhibited by low concentrations of intracellular ATP (IC_{50} 29 μM), questioning the physiological activity of monomeric TRPM6 channels (588). Extracellular ATP also inhibits TRPM6 activity via the purinergic receptor P2X4 (103).

A chemical gradient for Mg^{2+} entry in DCT cells is almost absent. The luminal Mg^{2+} concentrations vary between 0.2 and 0.7 mM, and the intracellular Mg^{2+} levels are typically in the range of 0.5–1 mM. Therefore, luminal Mg^{2+} entry is purely dependent on the negative membrane potential in the DCT cell. Luminal K^+ channels are indispensable for maintaining the necessary driving force for Mg^{2+} uptake. The voltage-gated K^+ channel Kv1.1 has been suggested to provide efflux K^+ currents resulting in hyperpolarization of the luminal membrane, although expression levels in the DCT are limited (104, 176). To prevent Mg^{2+} overload and hyperpolarization of the luminal membrane, intracellular Mg^{2+} blocks Kv1.1 (179). Interestingly, recent studies suggest that other potassium channels in the luminal membrane of DCT cells may have comparable roles. ROMK is prominently expressed in DCT, and its expression in this segment is regulated by dietary Mg^{2+} (104, 572). Similar to Kv1.1, intracellular Mg^{2+} blocks ROMK currents, suggesting a regulatory function on Mg^{2+} homeostasis (578). Moreover, indirect inhibition of ROMK by aldosterone or epithelial Na^+ channel (ENaC) blockers represent the only effective approach to prevent renal Mg^{2+} wasting in most clinical situations (140).

Several proteins have been proposed to mediate Mg^{2+} extrusion to the bloodstream, but general consensus of the extrusion mechanism has not been reached (424). Due to

the absence of a representative DCT cell model, the properties of Mg^{2+} extrusion have not been elucidated. Nevertheless, over the last decade several groups claimed to have identified Mg^{2+} extrusion proteins. Originally described in 2002, cyclin M2 (CNNM2, previously known as ACDP2) is exclusively expressed at the basolateral membrane of DCT and CNT cells within the kidney (105, 497, 545). Moreover, expression of CNNM2 is sensitive to dietary Mg^{2+} availability (104, 497). CNNM2 was initially depicted as Mg^{2+} transporter, since overexpression in *Xenopus laevis* oocytes allows uptake of a variety of divalent cations, with highest affinity for Mg^{2+} (184). However, these results could not be confirmed in mammalian cell lines (497). Alternatively, a Mg^{2+} -sensing function has been proposed, since CNNM2 harbors a Mg-ATP binding site in its cystathionine- β -synthase (CBS) domains (105). CNNM2 increases Mg^{2+} uptake in HEK293 cells (28). Nevertheless, it remains unclear whether CNNM2 mediates Mg^{2+} uptake directly or activates other Mg^{2+} carriers.

Recently mutations in the *SLC41A1* Mg^{2+} transporter were described to cause a nephronophthisis-like phenotype (251). Immunohistological studies showed expression in DCT, but the stainings were not conclusive about the subcellular localization (apical or basolateral) of SLC41A1 proteins (251). By the use of Mag-Fura, SLC41A1 was demonstrated to increase both Mg^{2+} absorption and Mg^{2+} extrusion (251, 289). These results suggest that SLC41A1 plays a role in DCT Mg^{2+} reabsorption, although further studies are necessary to elucidate the mechanisms by which SLC41A1 mediates Mg^{2+} transport.

Parvalbumin is exclusively expressed in DCT within the kidney, where it may function as a Ca^{2+}/Mg^{2+} buffer (456). Although parvalbumin has much higher affinity for Ca^{2+} than for Mg^{2+} (dissociation constants are ~ 5 – 10 nM for Ca^{2+} and ~ 30 μM for Mg^{2+}), the cation binding sites of parvalbumin will be mainly occupied by Mg^{2+} (377). This can be explained by the fact that the intracellular concentration of Mg^{2+} (0.5–1 mM) vastly exceeds that of Ca^{2+} (50–100 nM). In mouse and human kidney, parvalbumin is exclusively expressed in the early DCT (44). In late DCT and CNT, calbindin- D_{28K} is the main Ca^{2+} -binding protein. The exact role of parvalbumin in DCT remains to be investigated. Parvalbumin KO mice do not have altered serum or urine Mg^{2+} levels under basal conditions; these mice have reduced NCC expression, but display normal tubule morphology (44). DCT parvalbumin expression is highly sensitive to dietary Mg^{2+} availability, suggesting an important role for parvalbumin in Mg^{2+} reabsorption in DCT (104).

IV. MAGNESIUM IN PHYSIOLOGY AND PATHOPHYSIOLOGY

The human body contains $\sim 24g$ Mg^{2+} , of which 99% is stored in bone, muscle, and other soft tissues. Mg^{2+} is crit-

ical to the function of basically every organ in the human body. Moreover, Mg^{2+} deficiency is associated with a wide range of diseases, and as a result Mg^{2+} supplementation is considered as potential treatment in many of them (TABLE 2). This part of the review focuses on the organ-specific functions of Mg^{2+} and provides an overview of all major diseases in which Mg^{2+} may play a role.

A. Magnesium in Brain

Low serum Mg^{2+} values are associated with a wide range of neurological diseases such as migraine, depression, and epilepsy. Neuronal Mg^{2+} concentrations are of major importance in the regulation of *N*-methyl-D-aspartate (NMDA) receptor excitability. NMDA receptors are essential for excitatory synaptic transmission, neuronal plasticity, and excitotoxicity and therefore play an important role in developmental plasticity, learning, and memory (384). NMDA receptors are activated upon glutamate binding and mediate the influx of Ca^{2+} and Na^{+} ions and the efflux of K^{+} ions. Every NMDA receptor consists of four subunits, each with different biochemical properties (97). In Mg^{2+} deficiency, NMDA receptors become hyperexcitable, which can be explained by inhibitory function of extracellular Mg^{2+} on the receptors (335, 372) (FIGURE 6). Glutamate from the presynaptic neuron will bind both the ionotropic 2-amino-3-(3-hydroxy-5-methyl-isoxazol-4-yl)propanoic acid (AMPA)

and NMDA receptors on the postsynaptic neuron. At a normal membrane potential of -70 mV, Mg^{2+} ions block NMDA receptors. Therefore, only AMPA receptors will be activated and consequently facilitate an influx of cations. Only when the membrane potential rises above -60 mV the Mg^{2+} block is relieved and NMDA receptors are opened upon glutamate binding. The unlocking mechanism consists of a slow and a fast component, which depend on the relative expression of subunits that compose the channel (87, 536). Upon reduced extracellular Mg^{2+} concentrations, less NMDA channels will be blocked, and more NMDA channels can be opened at relatively low membrane potentials (351). This increased excitatory postsynaptic potential causes hyperexcitability of the neurons.

In the adult brain, this process is further amplified by the action of inhibitory γ -aminobutyric acid (GABA) receptors, whose function is also regulated by Mg^{2+} . GABA_A receptors are ionotropic anion channels that open upon GABA binding and facilitate Cl^{-} influx (250). Since the equilibrium potential of Cl^{-} is 10–20 mV lower than the membrane potential, this influx contributes to hyperpolarization of the neuronal cells. Extracellular Mg^{2+} stimulates GABA_A receptors resulting in hyperpolarized neuronal cells (353). When Mg^{2+} concentrations in the central nervous system (CNS) are low, GABA_A receptors are less stimulated. Consequently, the membrane potential will be higher,

Table 2. Therapeutic use of Mg^{2+}

Disease	Cochrane Review	Large-Scale Clinical Studies	Guidelines
<i>First drug of choice</i>			
Preeclampsia	RR: 0.41, 95% CI: 0.29–0.58 (130)		(223)
Arrhythmia–Torsades des Pointes			(591)
<i>Alternative drug of choice</i>			
Migraine			(237)
Asthma	RR: 0.53, 95% CI: 0.05–5.31 (404)	Magnetic	(3)
Super-refractory status epilepticus			(476)
Muscle cramps	No. cramps: -3.93% , 95% CI: -21.12 to 13.26% (168)		(274)
<i>Experimental</i>			
Stroke		FAST-MAG Images	
Subarachnoid hemorrhage	RR: 0.75, 95% CI: 0.57–1.00* (125)	IMASH MASH-II	
Myocardial infarction	OR: 0.59, 95% CI: 0.49–0.70 ⁺ (312)	LIMIT-2 ISIS-4	
Hypertension	DBP: -2.2 mmHg, 95% CI: -3.4 to -0.9 (119)		
Traumatic brain injury	GS: 0.02, 95% CI: -0.38 to 0.041 (25)		

This table summarizes all diseases in which Mg^{2+} has been considered as treatment by the official American guidelines. Moreover, it lists all large-scale clinical studies and meta-analyses of small-scale clinical studies by the Cochrane collaboration. RR, risk ratio; OR, odds ratio; CI, confidence interval; DBP, diastolic blood pressure; GS, Glasgow score. Reference numbers are given in parentheses. *In addition to standard nimodipine treatment. ⁺Early mortality in patients treated with <75 mmol of magnesium compared with placebo groups.

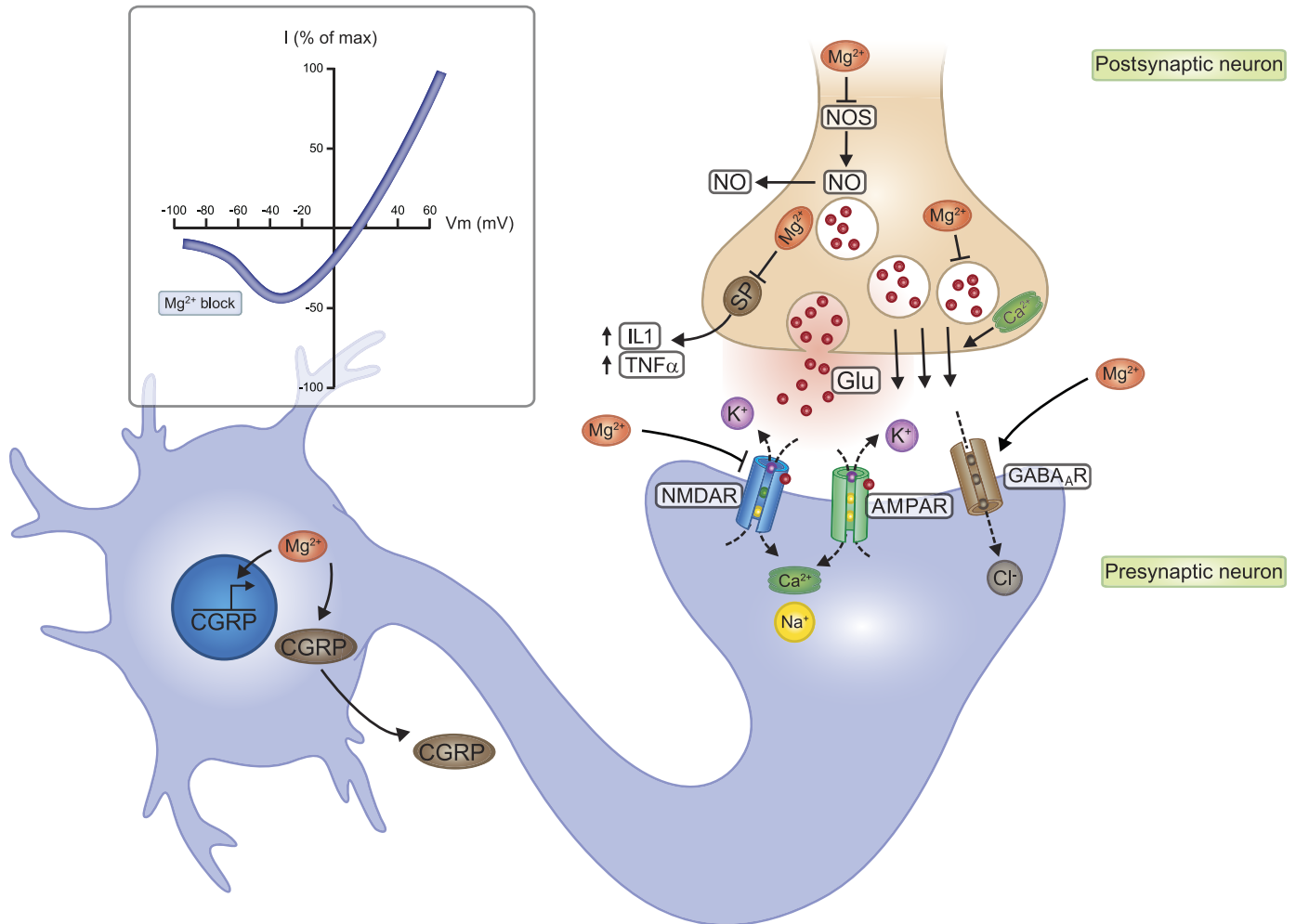


FIGURE 6. Magnesium in brain. Mg^{2+} is an important regulator of glutamate signaling in the brain. Upon glutamate release, glutamate binds NMDAR and AMPAR in the postsynaptic neuron. Mg^{2+} blocks the NMDAR at membrane potentials less than -60 mV. Therefore, AMPAR needs to depolarize the cell membrane before NMDAR will be activated. Moreover, Mg^{2+} stimulates $GABA_A$ R and thereby strongly influences the membrane potential of the postsynaptic neuron. In the presynaptic neuron, Mg^{2+} inhibits glutamate release by antagonizing calcium. Moreover, Mg^{2+} increases the expression and secretion of CGRP and inhibits the production of NO and the release of SP. *Inset:* current-voltage curve of NMDAR current. Mg^{2+} blocks the NMDAR receptor at voltages lower than -60 mV. NMDAR, *N*-methyl-D-aspartate receptor; AMPAR, α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptor; $GABA_A$ R, γ -aminobutyric acid receptor; CGRP, calcitonin gene-related peptide; NO, nitric oxide; SP, substance P; IL-1 β , interleukin-1 β ; TNF- α , tumor necrosis factor- α .

which in turn relieves the Mg^{2+} block of the NMDA receptor and contributes to hyperexcitability of the neurons.

The final mechanism contributing to the hyperexcitability of NMDA-receptor rich neurons is inhibiting glutamate release from the presynaptic neuron. Release of glutamate can be inhibited by high extracellular Mg^{2+} concentrations (257, 318, 486). Although the exact mechanism by which Mg^{2+} reduces glutamate release is still unknown, it could be related to the inhibition of voltage-gated Ca^{2+} channels, as glutamate release is triggered by an influx of Ca^{2+} after an action potential (364).

As a result of the described mechanisms, low extracellular Mg^{2+} levels in the CNS contribute to the hyperexcitability

of NMDA receptor. The excessive intracellular Ca^{2+} in the neurons may lead to the production of toxic reactive oxygen species (ROS) and eventually to neuronal cell death.

In addition to increasing the hyperexcitability of excitatory neuronal pathways, Mg^{2+} has an important role in the regulation of oxidative stress and the release of neuropeptides such as calcitonin gene-related peptide (CGRP) and substance P. CGRP is secreted from sensory neurons and has a vasodilatory effect (48). Mg^{2+} may increase CGRP expression and secretion, as has been shown in women with preeclampsia (26, 151), although an opposite effect was reported in women with Raynaud's phenomenon (359). Mg^{2+} deficiency increases the release of substance P, which is a neuroinflammatory tachykinin (554), stimulating the

secretion of inflammatory mediators such as IL-1, -2, -4, -5, -10, -12, and -13 as well as TNF- α (552, 554). Moreover, Mg²⁺ enhances the activity of nitric oxide synthases (NOS) through a NMDA receptor-dependent mechanism (80, 392). Nitric oxide (NO) has multiple functions in the brain including vasodilation, regulation of gene transcription, channel activity, and neurotransmitter release (493). Altogether, the role of Mg²⁺ in the regulation of neuropeptide release may have serious consequences in neuronal disease.

1. Migraine

Migraine has been linked to low levels of magnesium in serum and cerebrospinal fluid (CSF) (414, 457). Migraine headaches are the consequence of cortical spreading depression (CSD), which consists of an intense membrane depolarization and repolarization in neurons and glial cells (308, 388). CSD can be evoked by NMDA receptor activation (181). Therefore, patients with an increase in neuronal excitability due to low CSF Mg²⁺ levels are more susceptible to migraine attacks. Moreover, patients suffering from migraine are often hypotensive during attacks (462), which can be explained by increased NO levels. NO is an important vasodilator and modulator of brain blood flow (376). As an inhibitor of NO production, reduced Mg²⁺ values may result in decreased NO levels (80, 392).

The first reports of Mg²⁺ treatment for migraine patients appeared in the 1960s and 1970s (542), and since then the pharmacological role of Mg²⁺ has been slowly recognized. Although presently the effectiveness of Mg²⁺ treatment for the majority of patients is still debated (334, 386), Mg²⁺ is a second line drug for migraine patients (148, 237). Over the last decades several double-blind placebo-controlled randomized trials have provided evidence for a beneficial effect of oral Mg²⁺ supplementation on the number of migraine attacks (294, 393, 507) as well as the intensity of the pain during these attacks (149, 294). One clinical trial failed to show any favorable effect (397), but patients in this study suffered from diarrhea. Although Mg²⁺ is generally well tolerated, supplementation with certain Mg²⁺ salts often results in diarrhea and malabsorption of the Mg²⁺. Intravenous Mg²⁺ supplementation in acute migraine and cluster headache treatment provided pain relief in several cases (47, 112, 332, 333). In contrast, other studies failed to demonstrate such an effect (78). A recent meta-analysis combining five studies on the effect of intravenous Mg²⁺ administration on migraine did not show significant improvements on pain relief [Risk ratio (RR): -0.07, 95% confidence interval (CI): -0.23 to 0.09]. However, the relatively low number of patients included ($n = 295$) is one of the major limits of this analysis (86). Efficacy of Mg²⁺ treatment in combination with other drugs is doubtful; Mg²⁺ combined with metoclopramide and riboflavin did not demonstrate any efficacy in treating migraine (94, 328).

2. Depression

In 1921, Weston (557) reported the beneficial role of Mg²⁺ in treatment of patients with depression. Nevertheless, large-scale, placebo-controlled double blind clinical trials assessing the efficacy of Mg²⁺ supplementation on depression are still lacking. Studies examining the association between serum Mg²⁺ concentrations and depression severity are not conclusive; some studies report altered blood Mg²⁺ levels, while others do not find differences (reviewed in Ref. 114). Since serum Mg²⁺ levels do not necessarily reflect neuronal Mg²⁺ availability, determining Mg²⁺ levels in CSF may be more relevant for patients with depression. Only three cross-sectional studies have addressed this issue, and none of them found altered CSF Mg²⁺ concentrations in depressed patients compared with healthy controls (35, 171, 310). Despite this, several investigations have proposed that Mg²⁺ may relieve depression by blocking the NMDA receptor, whose dysfunction is a major causative factor in depression pathology (135, 143, 400, 557). To date, two interventional studies have investigated the role of Mg²⁺ in treating depression and their results are contradictory. In a randomized trial examining depressed elderly patients with diabetes mellitus type 2 and hypomagnesemia, Mg²⁺ supplementation was as effective as standard imipramine treatment (37). However, this study lacked a placebo control group. Moreover, it should be noted that imipramine therapy is nowadays largely replaced by selective serotonin reuptake inhibitors (SSRI). Another limit of the studies is the small sample population (23 patients). Therefore, large-scale studies are necessary to delineate a role of Mg²⁺ in the treatment and prevention of depression.

3. Epilepsy

Seizures are often associated with genetic and acquired forms of hypomagnesemia (see sect. V). Many studies have found that patients suffering from epilepsy display lower blood Mg²⁺ values (212, 375, 482). The link between Mg²⁺ status and the development of seizures may be explained by the role of Mg²⁺ in NMDA receptor blockade. Most studies addressing this issue show a small but significant decrease in CSF Mg²⁺ levels in epilepsy patients (492). In eclampsia patients, Mg²⁺ has proven to be successful in reducing the risk of recurrent convulsions (198). For other types of seizures, the evidence is less conclusive. In 1933 the first report of Mg²⁺ infusions in eight status epilepticus patients, a life-threatening form of epilepsy in which patients suffer from continuous seizures without regaining consciousness, was successful in all cases (496). However, modern reports of Mg²⁺ infusion treatment of status epilepticus patients have more variable outcomes and are not as conclusive (159, 383). Mg²⁺ infusions are therefore considered a second line of treatment, when anti-epileptic drugs and anesthetics have proven to be unsuccessful. Absence of large-scale randomized double blind placebo-controlled tri-

als obstructs the implementation of Mg^{2+} as general anti-epileptic treatment (9).

4. Stroke

Stroke is one of the major causes of death in the Western society and has been associated with a drop in serum Mg^{2+} levels (19). There may be multiple roles for Mg^{2+} in the etiology of stroke. Low serum Mg^{2+} levels increase NMDA receptor activity and thus more glutamate and Ca^{2+} influx. Excessive Ca^{2+} and glutamate influx via the NMDA receptor may be the basis of excitotoxicity during stroke (137). Since clinical trials with NMDA receptor antagonists have proven to be unsuccessful in treatment of stroke (256), it is, however, unlikely that NMDA receptor blockade alone can fully explain the role of Mg^{2+} in the development and onset of stroke. Mg^{2+} also blocks other voltage-gated Ca^{2+} channels that may be involved in Ca^{2+} cytotoxicity. Additionally, Mg^{2+} has a vasodilatory effect, which may be beneficial for patients suffering from ischemic stroke. Although more than 100 neuroprotective agents were tested in animals, not a single agent has been proven successful in a phase 3 clinical trial (111). After several pilot studies showed beneficial effects of Mg^{2+} on clinical outcome parameters, two large randomized controlled trials have been performed to determine the role of Mg^{2+} administration in stroke treatment (355, 356, 449). The Intravenous Magnesium Efficacy in Stroke Trial (IMAGES) enrolled over 2,500 stroke patients and gave a 16 mmol $MgSO_4$ bolus injection within 12 h of a stroke, followed by a maintenance dose of 65 mmol over 24 h. No beneficial effects were reported on the primary outcome, death and disability at 3 mo [odds ratio (OR) 0.95, 95% CI 0.80–1.13]. However, in a subgroup of patients treated within the first 3 h (3.3% of the cohort), a favorable death or disability outcome of 0.66 (95% CI 0.25–1.70) was reported (356). Since animal studies indicate that Mg^{2+} treatment is only successful when applied within 3 h of the onset of the stroke (579), a large-scale follow-up randomized controlled trial is currently running which aims to treat patients with Mg^{2+} within the first hours after stroke in a prehospital and emergency department setting. Although the trial is still running, first reports indicate that they have succeeded in including 72% of ~1,000 patients within the first hour (448). Final results from this FAST-MAG study will help determine the efficiency of Mg^{2+} treatment for stroke patients.

Approximately 5% of all strokes are caused by subarachnoid hemorrhage (SAH) that results from ruptured aneurysms. Delayed cerebral ischemia (DCI) is the major cause of death and disability in patients that survive the first 24 h (126). Interestingly, patients with SAH often present with hypomagnesemia (531, 532). Vasoconstriction is the main cause of DCI, and this may be enhanced when the patient is Mg^{2+} deficient. Over the last decade, several clinical trials have examined the addition of Mg^{2+} administration to the standard nimodipine treatment (358, 533, 555, 568). In

2005, in the Magnesium and Acetylsalicylic acid in Subarachnoid Hemorrhage (MASH-I) study poor outcomes were 23% reduced (RR 0.77; 95% CI 0.54–1.09) and DCI was reduced by 35% (RR 0.65; 95% CI 0.40–1.05) (533). However, more recent studies in larger study-cohorts did not report beneficial effects. In the intravenous magnesium sulfate for aneurysmal subarachnoid hemorrhage (IMASH) double-blind randomized placebo-controlled trial, 6 mo favorable outcome was similar between the patients given $MgSO_4$ and the control, 64 and 63%, respectively (RR 1.0; 95% CI 0.7–1.6) (568). Comparable results were obtained more recently, in the MASH-II multicenter randomized placebo-controlled in a cohort of 1,200 patients; 74% had a favorable outcome in the Mg^{2+} group and 75% in the control group (RR 1.03; 95% CI 0.85–1.25) (124). Overall, these studies suggest that intravenous Mg^{2+} treatment does not improve clinical outcome after aneurysmal subarachnoid hemorrhage.

5. Brain injury

Mg^{2+} deficiency is regularly found in patients with traumatic brain injury (TBI) and spinal cord injury (SCI) (81, 268). Reduced CSF Mg^{2+} levels increase oxidative stress (ROS, NO) and lipid peroxidation, which both contribute to the severity of TBI (77, 416). Additionally, it has been proposed that Mg^{2+} deficiency increases the release of substance P in TBI, resulting in neuronal cell death and edema (539). In animal experiments, Mg^{2+} improved sensorimotor/motor function as well as cognitive function (233). In a small study of 30 TBI patients, Mg^{2+} supplementation improved patient outcomes as measured by the Glasgow outcome scale (OR 4.13; 95% CI 1.39–12.27) (116). A large phase 3 randomized placebo-controlled clinical trial tested the effect of two doses of Mg^{2+} treatment on 6 mo mortality, seizures, functional measures, and neuropsychological tests in 500 TBI patients. Surprisingly, patients receiving Mg^{2+} treatment did significantly perform worse on the primary outcome than control patients (48 vs. 54; 95% CI –10.5 to –2), suggesting an adverse effect of Mg^{2+} (511). A meta-analysis on all clinical trials with TBI patients confirmed that there is no evidence for a neuroprotective role of Mg^{2+} in TBI (25). This demonstrates again the difficulties in translating the results obtained in animal studies to the clinic (111).

6. Parkinson's disease

Parkinson's disease is characterized by a loss of dopaminergic neurons. Parkinson's patients have low Mg^{2+} concentrations in cortex, white matter, basal ganglia, and brain stem (580). Interestingly, rats with chronic low Mg^{2+} intake exhibit a significant loss of dopaminergic neurons (381). In vitro experiments often use differentiated PC12 cells and 1-methyl-4-phenylpyridium ion (MPP^+) to model Parkinson's disease at the cellular level. In this experimental

model, mitochondrial Mg^{2+} concentrations were decreased as was demonstrated using the mitochondrial KMG-301 fluorescent Mg^{2+} probe (475). Moreover, Mg^{2+} transporter SLC41A1 is located on the PARK16 locus that is associated with Parkinson's disease (576). Recent characterization of the SLC41A1-pA350V single nucleotide polymorphism (SNP) linked to Parkinson's disease evidenced a gain-of-function effect (290). These studies suggest that Mg^{2+} supplementation may be beneficial for patients suffering from Parkinson's disease.

7. Other brain pathologies

Low serum Mg^{2+} levels have also been associated with a wide range of neurological pathologies including schizophrenia, bipolar disorder, neuroses, addiction, stress, and Alzheimer's disease (538). Although this suggests that Mg^{2+} deficiency plays a role in the etiology of these pathologies, all reports to date are of an epidemiological nature. There are currently no reports of clinical trials examining the effect of Mg^{2+} supplementation on the disease outcome of these diseases.

B. Magnesium in Lung

Dietary Mg^{2+} intake has been repeatedly associated with lung function, as assessed by forced expiratory volume

(FEV) and forced vital capacity (FVC) (60, 174). Lung Mg^{2+} research suffers from a lack of fundamental studies. As a result, the mechanisms that explain the role of Mg^{2+} in lung function are poorly understood, and hypotheses are mainly based on studies in other cell types and organs. Nevertheless, the role of Mg^{2+} in lung function may be explained at three levels: 1) Mg^{2+} has a strong vasodilator and bronchodilator effect; 2) Mg^{2+} regulates the release of acetylcholine (ACh) and histamine; and 3) Mg^{2+} acts as anti-inflammatory agent (FIGURE 7).

From studies on coronary artery related diseases, it is known that Mg^{2+} has a vasodilatory effect (512, 513). Like many vasodilators, Mg^{2+} also has a bronchodilating effect (229, 374). Although the mechanisms underlying Mg^{2+} -induced bronchodilation remain to be elucidated, Mg^{2+} is known to inhibit the release of ACh and histamine, both known to induce bronchoconstriction (88, 291, 398, 427). Moreover, Mg^{2+} may reduce the airway inflammation that underlies several lung diseases, including chronic obstructive pulmonary disorder (COPD) and cystic fibrosis. In line with this, Mg^{2+} deficiency has been reported in children with bronchitis, and low Mg^{2+} levels can induce an inflammatory response in lung allografts (43, 440). Most of our understanding of the role of Mg^{2+} in inflammation comes from studies in brain, heart, and intestine (450, 509, 553), and only a few studies have examined the anti-inflamma-

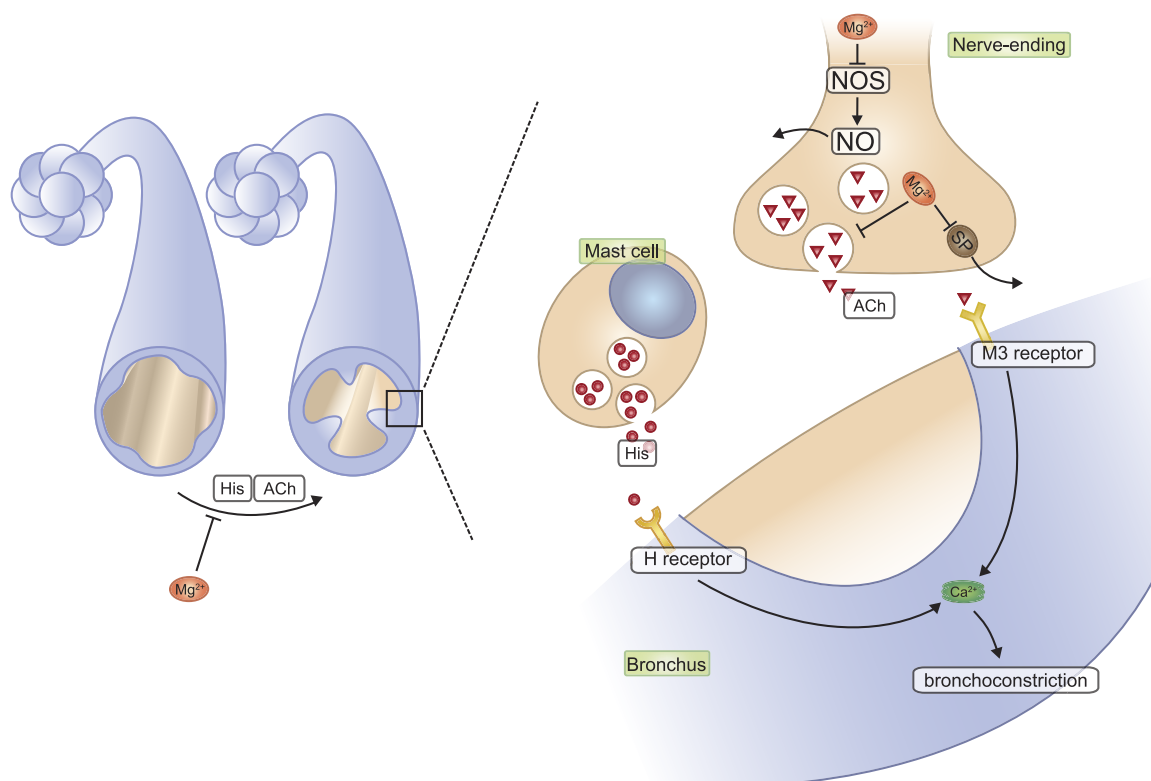


FIGURE 7. Magnesium in lung. Mg^{2+} stimulates bronchodilation by inhibiting the release of bronchoconstrictors histamine and acetylcholine from mast cells and neurons, respectively. Moreover, it inhibits the vasoconstriction by reducing the release of NO and SP. His, histamine; ACh, acetylcholine; NO, nitric oxide; SP, substance P.

tory function of Mg^{2+} in lung. Nevertheless, it is generally accepted that Mg^{2+} protects against inflammation by reducing oxidative stress, inhibiting substance P release, and preventing Ca^{2+} toxicity by inhibiting voltage-gated Ca^{2+} channels (450, 553). Mg^{2+} also modulates NF κ B activation by influencing lipid peroxidation (17, 153). All together, these characteristics make Mg^{2+} a potential therapeutic agent for lung diseases such as asthma and COPD.

1. Asthma

Several studies have reported low serum Mg^{2+} levels or low erythrocyte Mg^{2+} levels in asthmatic patients (21, 141, 219). However, others could not detect a Mg^{2+} deficiency in patients with asthma, suggesting that Mg^{2+} levels may depend on the severity of the disease (109, 276). Since Mg^{2+} relaxes smooth muscle cells, low Mg^{2+} levels cause bronchoconstriction and vasoconstriction, resulting in more asthmatic exacerbations (374). Moreover, Mg^{2+} regulates the release of ACh and histamine, which have both been implicated in asthma (170, 371). Interestingly, asthmatic C57/Bl6 mice have lower serum and intracellular Mg^{2+} concentrations than controls, which could be explained by a decreased renal TRPM6 expression (265). In 1940, Victor Haury (221) was the first to treat bronchial asthma patients with Mg^{2+} injections to relieve asthmatic paroxysms. Since then, ~25 randomized controlled studies have been published examining the effects of nebulized and intravenous Mg^{2+} administration in asthma patients. A recent systematic review failed to demonstrate significant improvement of respiratory function [standardized mean difference (SMD) 0.17, 95% CI -0.02 to 0.36] or hospital admissions (RR 0.68, 95% CI 0.46 to 1.02) for patients with acute asthma who had been given nebulized Mg^{2+} , although both parameters almost reached statistical significance (348). In studies using intravenous Mg^{2+} injection, respiratory function increased slightly in adults, but the most significant improvements were found in children (SMD 1.94, 95% CI 0.80–3.08), and children's hospital admissions were reduced (RR 0.70, 95% CI 0.54–0.90) (348). A recent systematic review within the Cochrane collaboration addressing nebulized Mg^{2+} for treatment of acute asthma concluded that respiratory function is not significantly improved in Mg^{2+} -treated patients compared with patients receiving β 2-agonists (404). However, the patients covered by this systematic review were mainly adult patients. Recently, the outcomes of the MAGNESium Trial In Children (MAGNETIC) randomized controlled study were published, showing improvement of the asthma severity score at 60 min (0.25, 95% CI 0.02–0.48) and 240 min (0.20, 95% CI 0.01–0.40) after inhalation of $MgSO_4$ (405). The MAGNETIC study was not published at the time of the systematic reviews, and future analysis including this trial may further substantiate the beneficial effects of Mg^{2+} in treatment of children with asthma.

2. Cystic fibrosis

Mg^{2+} deficiency has been repeatedly reported in patients with cystic fibrosis (210, 380). Patients with cystic fibrosis are often treated with recombinant human DNase-I to degrade the viscous mucus. However, the recombinant DNase requires Mg^{2+} to function, and efficiency of the treatment depends, therefore, on the Mg^{2+} status of the patient (444). It was suggested that Mg^{2+} treatment in itself would be sufficient to trigger endogenous DNase activity in the sputum (426). To further examine the potential role of Mg^{2+} as treatment for cystic fibrosis patients, a small double-blind, randomized, placebo-controlled crossover study tested the effect of oral Mg^{2+} supplementation on respiratory muscle strength and disease severity (444). Patients supplemented with Mg^{2+} increased their maximal inspiratory pressure (MIP) and maximal expiratory pressure (MEP). Although these first results are promising, more large-scale follow-up studies are necessary to assess the effects of Mg^{2+} in cystic fibrosis patients.

3. COPD

Serum Mg^{2+} levels have been associated with disease progression in patients with COPD (31, 46). Given the bronchodilating effects of Mg^{2+} , several studies have examined whether intravenous or nebulized Mg^{2+} may benefit COPD patients. In 1995, a small study of 27 COPD patients examined the effect of 1.2 g $MgSO_4$ infusion after β -agonist administration. Although the sample size was small, Mg^{2+} -treated patients demonstrated higher peak expiratory flow values compared with placebo-treated patients (22 ± 29 vs. $6 \pm 24\%$) (485). In another small randomized double-blind controlled trial, Mg^{2+} infusion resulted in improved functional respiratory capacity (-0.48 l, 95% CI: -0.96, -0.01), inspiratory capacity (0.21 l, 95% CI: 0.04–0.37), MIP (10 cmH $_2$ O, 95% CI: 1.6–18.4), and MEP (10.7 cmH $_2$ O, 95% CI: 0.20–21.2) (123). In contrast, a recent study using combined intravenous and nebulized $MgSO_4$ administration in 62 COPD patients did not detect significant effects on the primary outcome, as measured by hospital admission, intubation, and hospital death rates, nor did they find improved lung function (370). A larger recent study testing nebulized $MgSO_4$ did not detect improved lung function after 90 min, as determined by FEV (136). Patients that received 151 mg $MgSO_4$ in addition to standard salbutamol treatment demonstrated similar FEV (-0.026 l, 95% CI -0.15 to 0.095). Altogether, the efficacy of Mg^{2+} treatment in COPD remains unclear and may depend on the route of administration and the combination with the use of additional drugs.

C. Magnesium in Heart and Vasculature

Mg^{2+} plays an important role in heart function by influencing myocardial metabolism, Ca^{2+} homeostasis, vascular

tone, peripheral vascular resistance, and cardiac output. Mg^{2+} exerts its effects in three ways: 1) Mg^{2+} regulates the activity of ion channels in the cardiac cells, thereby affecting the electrical properties of the myocardium (354); 2) Mg^{2+} regulates myocardial contractility by influencing the intracellular Ca^{2+} mobility; and 3) Mg^{2+} has an anti-inflammatory and vasodilatory effect (FIGURE 8).

The cardiac action potential consists of five phases: phase 0 is the rapid depolarization by the influx of Na^+ . Phase 1 consists of rapid repolarization by efflux of K^+ . Phase 2, named the plateau phase, is the longest phase and marks Ca^{2+} entry. Phase 3 allows final repolarization of the cell by restoration of the membrane potential. Phase 4 is the stable phase with a resting potential of ± 90 mV (190). Mg^{2+} is

mainly important in phases 2 and 3 of the myocardial action potential, exerting its effect on K^+ and Ca^{2+} channels. In phase 2, Mg^{2+} inhibits L-type Ca^{2+} channels ($Ca_v1.2$) to prevent Ca^{2+} overload and cell toxicity (558). Mg^{2+} can bind a COOH-terminal EF hand motif of the channel and thereby influences the Ca^{2+} current (63). The effects of Mg^{2+} on the current through the L-type Ca^{2+} channels (I_{CaL}) may depend on the channel's phosphorylation state, since phosphatase treatment decreases the inhibitory effects of Mg^{2+} (547). In phase 3, delayed rectifier K^+ channels repolarize the cell by rapid-activating (I_{Kr}) and slow-activating (I_{Ks}) currents. High $[Mg^{2+}_i]$ inhibits I_K currents in frog and guinea pig cardiomyocytes (128, 563). This effect probably depends on the slow-activating component of the current, since rapid-activating currents seem insensitive to

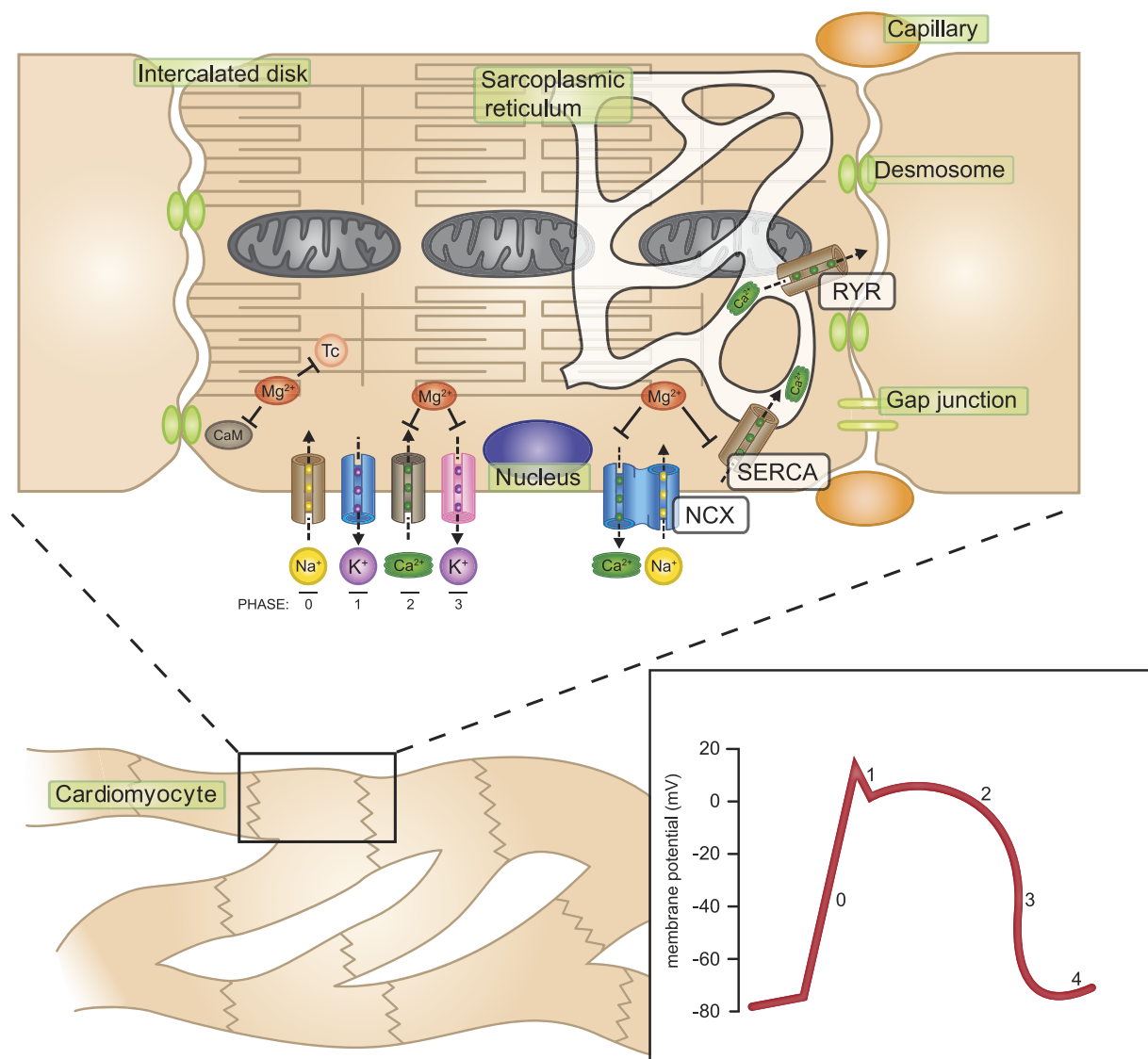


FIGURE 8. Magnesium in heart. Mg^{2+} influences phase 2 and phase 3 of the cardiac action potential by inhibiting L-type Ca^{2+} channels (phase 2) and delayed rectifier K^+ currents (phase 3). Moreover, Mg^{2+} directly influences the cardiac muscle contraction by antagonizing Ca^{2+} binding of troponin C and calmodulin. It further modifies Ca^{2+} availability by affecting NCX and SERCA activity. *Inset:* the cardiac action potential. The numbers indicate the phases of the action potential. Tc, troponin C; CaM, calmodulin; NCX, Na^+ - Ca^{2+} -exchanger; SERCA, sarcoplasmic/endoplasmic reticulum Ca^{2+} -ATPase; RyR, ryanodine receptor.

Mg²⁺ inhibition (499). The intracellular block of inward rectifier K⁺ channels Kir2.1 and Kir2.2 by Mg²⁺ substantially influences phase 3 and phase 4 of the action potential (556). This block of the I_{K1} current is relieved by high extracellular K⁺ concentrations (56).

In recent years an increasing amount of attention has been directed to the role of [Mg²⁺_i] in cardiac excitation-contraction coupling (342). Mg²⁺ has often been considered as a natural Ca²⁺ antagonist, since it can compete with Ca²⁺ for binding sites in proteins and Ca²⁺ transporters (257). The effect of Mg²⁺ on cardiomyocytes is mainly explained by its role of Ca²⁺ mobilization. Mg²⁺ binds calmodulin, troponin C, and parvalbumin, and therefore a reduced [Mg²⁺_i] may result in alterations in the unbound Ca²⁺ fraction (15). Mg²⁺ may also affect the main Ca²⁺-transporting proteins in the cardiomyocytes. Mg²⁺ acts as substrate in a complex with ATP for cardiac Ca²⁺-ATPases and alters the affinity of Na⁺-Ca²⁺ exchanger type 1 for Ca²⁺ (NCX1) (55, 58, 342). There is a dearth of physiological studies on the effect of Mg²⁺ on NCX1 and SERCA activity, and available studies mainly rely on modeling and in vitro experiments. Nevertheless, tight regulation of [Mg²⁺_i] in cardiac cells is necessary for optimal cardiac function. This is substantiated by the fact that high [Mg²⁺_i] can cause cardiac arrest, and by the impressive capacity of cardiomyocytes to maintain constant [Mg²⁺_i] (506, 521).

An important role of Mg²⁺ in heart and vasculature function is a substantial vasodilatory effect that has been reported in animal and human studies (18, 161, 512, 513). Although the results from animal studies suggest that Mg²⁺-induced vasodilation is due to the regulation of NO synthesis, human studies show that it is independent of NO activity (512, 513). Moreover, Mg²⁺ deficiency promotes oxidative stress notably in endothelial cells (118, 561), resulting in increased reactive oxygen species (ROS) and cytotoxicity (590). In contrast, high Mg²⁺ in the cell increases eNOS activity and suppresses the synthesis of vasoconstrictor endothelin-1 (277, 327). In conditions of low Mg²⁺-induced oxidative stress, the endothelium develops a state of permanent inflammation, which is marked by increased NFκB activity (153). NFκB is the master regulator of transcription of cytokines and pro-inflammatory genes, including IL-1α. As a result of this local inflammation, the vessel wall will recruit monocytes and trigger the proliferation and migration of vascular smooth muscle cells. These processes are facilitated by the increased expression of matrix metalloproteases 2 and 9 in low Mg²⁺ conditions (153, 382). Eventually low Mg²⁺ concentrations may, therefore, result in atherosclerosis, vascular calcifications, or thrombosis.

1. Coronary artery disease

Over the last 20 years, an increasing number of studies have demonstrated that low serum Mg²⁺ levels and low Mg²⁺ intake are associated with an increased risk of coronary

artery disease (CAD), atherosclerosis, and metabolic syndrome (8, 222, 316, 587). Low serum Mg²⁺ levels have been associated with a higher mortality risk in CAD patients (162). There may be several ways in which Mg²⁺ supplementation benefits patients with CAD. Since Mg²⁺ has a strong anti-inflammatory role, Mg²⁺ results in an improved lipid profile, reduced free oxygen radicals, and improved endothelial function (317, 490). Mg²⁺ prevents blood clotting by reducing platelet aggregation (437), and it has a strong vasodilator effect (512, 513). These properties make Mg²⁺ an important factor in the development and management of CAD. Mg²⁺ improves several aspects of vascular function in CAD. Reduced serum Mg²⁺ concentrations are associated with an increase in carotid intima-media thickness and risk for sudden cardiac death (218, 324, 391). In a randomized, double-blind, placebo-controlled study of 50 CAD patients, oral Mg²⁺ supplementation ameliorated endothelial function (472). Moreover, Mg²⁺ reduced platelet-induced thrombosis in a randomized prospective, double-blind, crossover, and placebo-controlled study in 42 CAD patients (471). Six months of Mg²⁺ supplementation increased maximal oxygen uptake ($\dot{V}O_{2\max}$) and left ventricular ejection fraction (LVEF) in 53 CAD patients (401). Taken together, these studies suggest that Mg²⁺ levels should be closely monitored in CAD patients and propose Mg²⁺ as potential drug to improve quality of life in CAD patients.

2. Myocardial infarction

In the 1970s, a pioneering paper by Abraham et al. (9) associated myocardial infarction with a significant drop in serum Mg²⁺. Indeed, low serum Mg²⁺ levels have been associated with an increased risk of acute myocardial infarction (AMI) (484). The role of Mg²⁺ in preventing myocardial infarction may be caused by relaxing endothelial and smooth muscle cells in the heart and vasculature (18, 161, 512). Moreover, heart rate variability is a risk factor for AMI and Mg²⁺ may prevent arrhythmia (474, 484). Several studies have addressed the effect of Mg²⁺ on myocardial infarction. In the 1980s, studies reported a 20% reduction of infarct size in Mg²⁺-treated patients, and decreased mortality after Mg²⁺ infusion (352, 415). Several follow-up studies suggested that decreased rates of arrhythmias after infarction explain the lower mortality (76, 487). After these initial promising results, several clinical trials have addressed this subject. The Leicester Intravenous Magnesium Intervention Trial 2 (LIMIT-2) included 2,316 AMI patients and found 24% reduced mortality after 28 days in the Mg²⁺ group (95% CI: 1–43%) (569). Moreover, two smaller studies by Shechter and colleagues reported reduced mortality rates of 50 and 40% (469, 470). Contrary to these findings, in the fourth Infarct Survival and Magnesium in Coronaries (ISIS-4) study, a randomized factorial trial in 58,050 patients showed no beneficial effects of intravenous Mg²⁺ administration on survival (4). The results from the ISIS-4 study were called into question because

the statistical analysis did not reflect the heterogeneous studies used in the analysis, the low mortality in the placebo-group was indicative of a low-risk comparison group, and the late time-point of Mg^{2+} administration, namely, after and not during reperfusion, was at odds with animal data demonstrating the effectiveness of Mg^{2+} (463). However, also a second large-scale randomized double-blind study with 6,213 patients and an earlier time point of Mg^{2+} infusion (3.8 h compared with 8 h in ISIS-4) also failed to show a decrease on 30-day mortality rates (2). Indeed, a recent meta-analysis within the Cochrane collaboration concludes that there is no beneficial effect of Mg^{2+} on mortality in AMI patients (OR 0.99, 95% CI 0.94 to 1.04) (312). However, it should be noted that the ISIS-4 study provides 72% of the power in this analysis and that the Mg^{2+} doses used within the analyzed studies differ significantly. Nevertheless, current guidelines do not recommend Mg^{2+} administration in AMI patients.

3. Arrhythmia

In 1935, Dr. Zwillinger was the first to report an anti-arrhythmic effect of Mg^{2+} , and since then sporadic reports of patients treated with Mg^{2+} have appeared in the literature (595). However, the field suffers from a lack of large-scale randomized controlled trials, and therefore the exact clinical benefit of Mg^{2+} in treatment of arrhythmia remains to be determined. Mg^{2+} is known to have a function in regulating cardiac K^+ and Ca^{2+} channels, so it affects the cardiac action potential. As a result, hypomagnesemia in itself has been proposed as a cause of arrhythmia, specifically in combination with stress or alcoholism (362, 524). Clinical studies have demonstrated that treatment success strongly depends on the type of arrhythmia (232). Atrial fibrillation is one of the most common and dangerous complications after cardiac surgery. In the last few decades, several small-scale studies have examined the effect of Mg^{2+} in preventing these fibrillations (93). Meta-analysis of these studies concluded that Mg^{2+} infusion may prevent atrial fibrillations (201, 474). Therefore, the European Association for Cardiothoracic Surgery and the Canadian Cardiovascular Society recommend prophylaxis with intravenous $MgSO_4$. However, recent reports point to problems associated with many studies at the level of double-blinding, primary outcome, and intention-to-treat analysis. When only high quality studies are included, Mg^{2+} does not have a preventive effect on atrial fibrillations (OR: 0.94, 95% CI: 0.61–1.44) (93). A similar discrepancy between low- and high-quality studies has been noted for studies addressing the effects of Mg^{2+} on supraventricular arrhythmias (108). Interestingly, for treatment of torsades des pointes, Mg^{2+} has been implemented as a first line of treatment after several studies in the 1980s showed beneficial effects (395, 527, 528). However, due to the absence of large-scale clinical trials, optimal doses for treatment are still under debate (211, 243, 244). Other arrhythmias, including refractory ventricular fibrillation and monomor-

phic ventricular tachycardia, are insensitive to Mg^{2+} treatment (150, 220).

4. Preeclampsia

Since the 1950s, intravenous $MgSO_4$ administration has gradually become the standard treatment for preeclampsia and eclampsia, and nowadays the treatment is widely advocated by the World Health Organization (223, 407, 594). The mechanisms of action underlying the effect of Mg^{2+} in the treatment of these patients are largely unknown. Mg^{2+} may reduce preeclampsia by its effect as vasodilator in the vasculature, but it cannot be excluded that Mg^{2+} also functions as an anticonvulsant through blockade of the NMDA receptor and reduces cerebral edema (147). Recent Cochrane systematic reviews showed that $MgSO_4$ treatment in preeclamptic women reduced the risk for eclampsia by >50% (RR 0.41, 95% CI 0.29–0.58), and there was a trend towards lower maternal mortality (RR 0.54, 95% CI 0.26–1.10) (130). $MgSO_4$ demonstrated similar ratios for reduced risk for eclamptic convulsions comparable to anti-convulsant medication, with 59% compared with diazepam (RR 0.41, 95% CI 0.29–0.58) and 66% compared with phenytoin (RR 0.34, 95% CI 0.24–0.49) (131, 132). Moreover, a recent meta-analysis of “real-world” use of $MgSO_4$ for the treatment of preeclampsia confirmed the results from the clinical trials and showed ~50% reduction of eclampsia risk in most studies (337). Interestingly, although Mg^{2+} was successful in preventing eclampsia, it did not change the risk of death or disability for children at 18 mo (RR 1.06, 95% CI 0.90–1.25) or the risk of death or disability for women at 2 yr (RR 0.84, 95% CI 0.60–1.18) (5, 6).

5. Hypertension

Low serum Mg^{2+} levels are frequently linked to high blood pressure (30, 267, 270). Intracellular Mg^{2+} may reduce the intracellular Ca^{2+} concentration within vascular smooth muscle cells. This Mg^{2+} -induced vasodilation is thought to be the mechanism by which Mg^{2+} alters the blood pressure. Moreover, high extracellular Mg^{2+} reduces the endothelin-1 expression and causes an increase in prostacyclin (PGI_2) levels, contributing to vasodilation (27, 59, 303, 447). Additionally, Mg^{2+} inhibits the production of NO (392). Mg^{2+} was first used to lower blood pressure in 1925, when Mg^{2+} infusion was found to lower blood pressure by reducing the peripheral vascular resistance (53). Several studies have shown that oral Mg^{2+} intake reduces both systolic and diastolic blood pressure (260, 275, 559); however, other studies fail to see such an effect (72, 152). A systematic review of the Cochrane Hypertension Group reported a small reduction of diastolic blood pressure (DBP; -2.2 mmHg, 95% CI -3.4 to -0.9), but not of the systolic blood pressure (SBP; -1.3 mmHg, 95% CI -4.0 to 1.5) (119). Another meta-analysis detected a small but dose-

dependent effect of Mg^{2+} on blood pressure, for each 10 mmol/day -4.3 mmHg SBP (95% CI -6.3 to -2.2) and -2.3 mmHg DBP (95% CI -4.9 to 0.0) (262). However, the outcomes may be biased by the inclusion of poor quality studies that tend to overestimate the effects. In contrast, a meta-analysis of a subset of studies, including patients on antihypertensive drugs with high blood pressure (SBP >155 mmHg) reports much stronger effects of oral Mg^{2+} treatment on SBP (-18.7 mmHg, 95% CI -14.95 to -22.45) and DBP (-10.9 mmHg, 95% CI -8.73 to -13.1) (425). These results suggest that Mg^{2+} may be beneficial for certain subgroups of hypertensive patients and that comparing the highly heterogeneous studies may underestimate the effects of Mg^{2+} in these groups.

B. Vascular calcification

Vascular calcification is frequently observed in patients suffering from chronic kidney disease (CKD) (452). Calcifications are a major contributor to cardiovascular death that accounts for 50% of all deaths in CKD (522). Vascular calcification is the consequence of a disturbed mineral metabolism including increased serum P_i levels. Increased serum levels of FGF23 and PTH enhance the formation of calcification. Low serum Mg^{2+} levels are associated with vascular calcification, and hemodialysis patients with higher serum Mg^{2+} levels show higher survival (258, 259). Although the mechanisms of action are not completely understood, there are two contributing factors: 1) Mg^{2+} prevents the formation and deposition of Ca/P nanocrystals and the development of apatite structures (83); and 2) Mg^{2+} inhibits the transdifferentiation of the smooth muscle cells in the vessel wall into osteoblast-like cells (282). In both processes Mg^{2+} prevents vascular calcification, and thus hypomagnesemic patients are at risk. Therefore, Mg^{2+} supplementation has been proposed as P_i -binder to reduce vascular calcification in CKD patients. Several studies have shown that combined administration of Ca^{2+} and Mg^{2+} is as effective as standard treatment options (252). In the recent CALMAG study comparing $MgCO_3/Ca(OAc)_2$ with Sevelamer-HCl in 200 hemodialysis patients, both treatments were effective in reducing serum P_i levels without increasing Ca^{2+} levels (106).

D. Magnesium in Muscle

Mg^{2+} mainly exerts its effects on skeletal muscle function as a Ca^{2+} antagonist on Ca^{2+} -permeable channels and Ca^{2+} -binding proteins. Muscle contraction is a highly Ca^{2+} -dependent process, initiated Ca^{2+} release from the sarcoplasmic reticulum. Ca^{2+} binds to troponin C and myosin to induce the conformational changes of these proteins that will result in contraction (193). Mg^{2+} competes for the Ca^{2+} -binding sites on these proteins. Although the affinity of troponin and myosin for Mg^{2+} is much lower than for Ca^{2+} (292), the effects of Mg^{2+} are not negligible (239,

403). In the resting state, Mg^{2+} is present in concentrations 10,000 times higher than Ca^{2+} in muscle cells (292); therefore, Mg^{2+} will occupy all Ca^{2+} binding sites, and it is only after Ca^{2+} is released from the sarcoplasmic reticulum that Mg^{2+} is replaced. However, in Mg^{2+} -deficient conditions, not as much Ca^{2+} is required to displace Mg^{2+} , resulting in hypercontractibility, which presents as muscle cramps and spasms in the clinic.

Moreover, as a cofactor of ATP, Mg^{2+} availability is essential for the function of the ryanodine receptor (RyR), which allows rapid Ca^{2+} release from the ER, and Ca^{2+} -ATPase of sarcoplasmic reticulum (SERCA), which mediates the return of Ca^{2+} to the SR after contraction. Intracellular Mg^{2+} and Ca^{2+} concentrations determine how many ATP molecules bind to the RyR, which determines the opening of these channel receptors and the release of Ca^{2+} (117). SERCA also depends on Mg^{2+} as cofactor of ATP. Additionally, Mg^{2+} can bind to the Ca^{2+} binding pocket of SERCA (584). The role of Mg^{2+} in the regulation of the Na^+ - Ca^{2+} exchanger (NCX) has been poorly studied so far.

1. Muscle cramps

Muscle cramps are a recurrent and prominent symptom in patients with severe/chronic hypomagnesemia (49, 215). Although the role of Mg^{2+} in the pathogenesis of muscle cramps is not completely understood, it is hypothesized that Mg^{2+} directly influences muscular contractions by antagonizing Ca^{2+} -binding proteins. Moreover, Mg^{2+} -deficient patients may suffer from neuronal hyperexcitability that can contribute to muscular contraction. Mg^{2+} has consequently been considered as treatment for muscle cramps in several studies. However, there is little evidence that Mg^{2+} may relieve muscle cramps in the general population. A recent Cochrane systematic review and meta-analysis of all published studies shows no significant reduction in the number of cramps after Mg^{2+} treatment (-3.93% , 95% CI: -21.12 to 13.26%) (168). In contrast, a Cochrane review from 2002 demonstrated that Mg^{2+} might be beneficial for muscle cramps in pregnancy (OR 0.18, 95% CI 0.05–0.60) (583). Both meta-analyses are limited by a relatively small patient population. Large-scale studies are necessary to ascertain the utility of Mg^{2+} for specific subpopulations or disease-related muscle cramps.

E. Magnesium in Pancreas

Mg^{2+} has been implicated in both endocrine and exocrine functioning of the pancreas. In the pancreatic acini, intracellular Mg^{2+} antagonizes Ca^{2+} -activated signaling events and enzyme secretion (573). It is known that ACh and cholecystokinin 8 (CCK8) evoke an increase in intracellular Ca^{2+} , which in turn activates calmodulin, resulting in the phosphorylation of proteins on the enzyme-containing ves-

icles. These vesicles will migrate towards the plasma membrane for exocytosis and secretion. Interestingly, ACh and CCK8 activation cause a marked Mg^{2+} efflux, allowing Ca^{2+} signaling to occur (483, 564). Its antagonizing role may be explained by inhibition of Ca^{2+} transporting proteins such as SERCA, PMCA, and Ca^{+} -ATPases (120, 584).

In the islets of Langerhans, Mg^{2+} may influence the secretion of insulin, although experimental results are conflicting as to whether insulin secretion is increased or decreased. Milner and Hales (344) reported that Mg^{2+} reduced insulin secretion in an ex vivo model using rabbit pancreas. These results were confirmed in rat pancreas and rat insulinoma cells (99, 357). However, in rats, one study found increased insulin secretion in Mg^{2+} -deficient rats, while other studies in Mg^{2+} -deficient rats did not report alterations in plasma insulin concentrations (202, 338, 418). In addition, patients with low serum Mg^{2+} levels show decreased insulin secretion (422). The discrepancies among different experiments may be explained by the Ca^{2+} availability in each of the models. Since the effect of Mg^{2+} on insulin can be explained by its Ca^{2+} antagonizing role, it may not depend on the Mg^{2+} concentration itself, but on the cytosolic Ca^{2+}/Mg^{2+} ratio.

1. Diabetes

Patients with diabetes mellitus type 2 often have low serum Mg^{2+} levels (36, 491, 541). These low serum levels are associated with poor disease outcome and may even increase mortality (98). Hypomagnesemia may contribute to the development of diabetes mellitus type 2 by increasing insulin resistance. Insulin receptors (IR) are part of the family of tyrosine kinase receptors, and the kinase function is dependent on the binding of two Mg^{2+} ions (249). Upon activation of the IR, a complex intracellular signaling cascade is activated and mediated via insulin receptor substrate proteins (505). In low Mg^{2+} conditions, activation of the IR may result in diminished signal transduction, contributing to insulin resistance. Studies with hypomagnesemic rats bear this out, as lower IR phosphorylation was detected, although differences between individual organs were reported (419, 498). It has also been proposed that increased expression of other effectors such as IL-1, IL-6, IL-8, TNF- α , norepinephrine, epinephrine, and ROS may contribute to insulin resistance in Mg^{2+} deficiency (206). Interestingly, common SNPs in the *TRPM6* gene are associated with an increased risk of developing diabetes mellitus type 2 (489). *TRPM6* cannot be activated by insulin when these SNPs are present (361). These results suggest that Mg^{2+} levels may influence the onset and development of diabetes mellitus type 2. Several studies have examined the clinical effects of oral Mg^{2+} supplementation on glycemic control in diabetes mellitus type 2 patients. Some of these studies demonstrate impressive effects in reducing glucated hemoglobin (HbA_{1c}) levels and fasting glucose concentrations (385, 423), but other studies show no improvement of gly-

cemic control (107, 204). A meta-analysis of 8 studies, including a total of 370 patients, evidenced a reduction of fasting glucose levels (-0.56 mM; 95% CI, -1.10 to -0.01), reflected in a nonsignificant ($P = 0.1$) reduction of HbA_{1c} levels (-0.31% , 95% CI, -0.81 to 0.19) (488). All together, these results indicate that Mg^{2+} supplementation may be a promising avenue for achieving glycemic control in diabetes patients.

F. Magnesium in Liver

The role of Mg^{2+} in the liver is poorly studied. However, given that many of the enzymatic reactions that take place in the hepatocytes are dependent on Mg^{2+} , particularly in fat metabolism, the importance of Mg^{2+} should not be underestimated. Mg^{2+} supplementation has been reported to reduce alanine aminotransferase (ALT) levels in obese women with hypomagnesemia (421). However, this result could not be reproduced in normomagnesemic patients, although a lower dose of Mg^{2+} was used in this study (271). The first reports of hypomagnesemia in liver diseases such as cirrhosis and nonalcoholic fatty liver disease suggest that liver function contributes to proper intestinal Mg^{2+} absorption (286, 526). Currently, the first clinical trials are being initiated to test the effects of Mg^{2+} supplementation in patients with liver cirrhosis.

G. Magnesium in Immunity

Mg^{2+} is considered as an anti-inflammatory agent that reduces the expression and release of substance P and other proinflammatory molecules. Mg^{2+} also influences acquired immunity by regulating the proliferation and development of lymphocytes (156). Deletion of the *TRPM7* Mg^{2+} channel caused cell death in the chicken B cell line DT40, which could be partially rescued by culturing the cells in high Mg^{2+} containing medium (455). In a mouse model with a specific T-cell deletion of *TRPM7*, T lymphocyte development was blocked at the $CD4^{-}CD8^{-}$ stage, resulting in decreased $CD4^{+}$ and $CD4^{+}CD8^{+}$ cells in the thymus (266). Moreover, mutations in the *MagT1* Mg^{2+} channel are causative for immunodeficiency and have been associated with decreased $CD4^{+}$ T lymphocyte levels (311). These results suggest that Mg^{2+} is essential for T lymphocyte development and proliferation.

1. X-linked T-cell immunodeficiency

Patients with X-linked immunodeficiency with Mg^{2+} defect, Epstein-Barr virus infection and neoplasia (XMEN) have mutations in *MagT1* (79, 311). They present with chronic Epstein-Barr virus infections, low $CD4^{+}$ T-cell counts, and defective T-lymphocyte activation. These effects are hypothesized to result from a loss of PLC γ 1 activation due to reduced Mg^{2+} influx via *MagT1* (311). Recent studies in asthma pa-

tients confirm the importance of Mg^{2+} availability for $CD4^+$ function (315). An increased risk in T-cell lymphoblastic leukemia has been associated with Mg^{2+} deficiency (441). However, the role of Mg^{2+} in T-cell signaling needs to be investigated before further conclusions can be drawn. Special attention should be given to the involvement of other Mg^{2+} carriers, since TRPM7-deficient T cells seem protected from Fas-receptor-induced apoptosis (115).

H. Magnesium in Bone

Bone hydroxyapatite structures mainly consist of P_i and Ca^{2+} and are bound by Mg^{2+} ions at the surface of the hydroxyapatite crystals. Mg^{2+} increases the solubility of the minerals and thereby acts on the crystal size and formation (443). Crystals in Mg^{2+} -deficient bone are larger, and the bone may therefore be brittle and more susceptible to fractures (89). Moreover, Mg^{2+} stimulates osteoblast proliferation, suggesting that Mg^{2+} deficiency results in decreased bone formation (320) (FIGURE 4). The role of bone in Mg^{2+} homeostasis is described in more detail in section IIIB.

1. Osteoporosis

Several studies have associated low serum Mg^{2+} values with osteoporosis (61, 213). Although most studies have been performed in postmenopausal women, there is some evidence of low bone Mg^{2+} content in elderly subjects. In this study the patients had normal serum Mg^{2+} levels, but displayed significantly increased retention in a loading/tolerance test (91). A few small-scale studies have examined the effects of oral Mg^{2+} supplementation (200–750 mg/day) on bone mineral density (BMD) in patients with osteoporosis. In a pioneering study in 1991, daily administration of 600 mg Mg resulted in an 11% increased BMD after 12 mo (10). However, many other supplements including 500 mg/day Ca were simultaneously used, making it difficult to distinguish the effects of Mg^{2+} . Subsequently, multiple studies have examined the effect of Mg^{2+} supplementation in different populations (138, 436, 494). Mg^{2+} seems to increase BMD in all of the studies, although the effects are relatively small (1–3%) and the small study sizes limit the conclusions that can be made from them. Testing the effect of Mg^{2+} supplementation in large cohorts of osteoporosis patients may further establish Mg^{2+} supplements to treat osteoporosis.

V. DISTURBANCES OF MAGNESIUM HOMEOSTASIS

Over the last decade, clinical interest in Mg^{2+} has been growing. Mg^{2+} deficiency has been associated with a wide range of diseases including diabetes mellitus type 2, hypertension, migraine, and depression. Therefore, the Mg^{2+} balance in patients is clinically significant; however, Mg^{2+} status in patients is not routinely determined. When Mg^{2+} is assessed in pa-

tients, total serum Mg^{2+} is the most commonly measured parameter. However, total serum Mg^{2+} levels do not necessarily represent body Mg^{2+} availability, since ionized Mg^{2+} determines the bioactive fraction. Only 2% of the clinical laboratories in the United States offer ionized Mg^{2+} tests, thus leaving total serum Mg^{2+} as the most used clinical representative of the patient's Mg^{2+} status (139).

However, serum Mg^{2+} levels represent <1% of the body Mg^{2+} content, and therefore, it is a poor predictor of the body Mg^{2+} status. Additionally, the Mg^{2+} concentration in red blood cells is higher than in the serum (1.65–2.65 mM), and extra care should be taken to prevent hemolysis, which can result in a misrepresentation of total serum Mg^{2+} (520). In a recent systematic review, red blood cell (RBC) Mg^{2+} content has been proposed as an alternative measure to determine Mg^{2+} status, since its value is strongly affected by alterations in dietary Mg^{2+} intake in six studies ($n = 130$) (565). Based on the limited changes of ionized serum concentration after dietary Mg^{2+} supplementation or depletion, the authors reject ionized Mg^{2+} as marker for patient Mg^{2+} status. However, the rationale that alterations in dietary Mg^{2+} intake necessarily result in a physiologically relevant effect in Mg^{2+} availability is questionable, since the kidney and bone have a large capacity to compensate for reduced Mg^{2+} absorption. Only after a long-term depletion, patients may develop a clinically relevant hypomagnesemia.

However, the opposite also holds true; patients may be severely Mg^{2+} deficient, although serum or RBC Mg^{2+} levels are normal. In this case, the Mg^{2+} concentrations in bones and soft tissues are severely decreased, after long-term compensatory Mg^{2+} release to keep serum Mg^{2+} levels within normal range. To identify such patients, a Mg^{2+} loading test has been proposed in which acute oral Mg^{2+} administration is used to assess Mg^{2+} retention (363). In this test, the serum Mg^{2+} concentration and fecal Mg^{2+} excretion are calculated to determine intestinal Mg^{2+} absorption. Mg^{2+} -deficient patients will have lower bone Mg^{2+} content and therefore show high Mg^{2+} retention (90). Thus the loading test allows quantification of the exchangeable pool of Mg^{2+} , which is more sensitive than serum Mg^{2+} concentrations. However, this test is rarely used in clinic, and as a result, standardization is lacking. Within a research setting, a urine excretion of <60% of the Mg^{2+} load is generally considered normal.

If a Mg^{2+} disturbance is suspected, urinary Mg^{2+} concentrations are regularly determined. It should be noted that reliable urine Mg^{2+} determinations require at least a complete 24-h sampling, since the circadian rhythm influences renal Mg^{2+} excretion (158). The results of urine Mg^{2+} tests may provide information about the cause of Mg^{2+} deficiencies; normal or high urinary excretion indicates renal Mg^{2+} wasting, whereas low Mg^{2+} excretion suggests reduced intestinal absorption. This information can then be used to guide treatment plans.

Importantly, physicians must be aware that urinary Mg^{2+} concentrations are poor indicators of a patient's Mg^{2+} status when kidney function is reduced. In several pathophysiological circumstances such as diabetes and chronic kidney disease, filtration may be altered. Additionally, the use of certain drugs including diuretics may cause bias in determining a patient's Mg^{2+} status.

A. Hypermagnesemia

Serum Mg^{2+} levels above 1.1 mM are generally considered hypermagnesemic. Hypermagnesemia may be clinically observed in patients suffering from nausea, vomiting, lethargy, headaches, and/or flushing. When Mg^{2+} levels rise above 3.0 mM it may even cause severe cardiac defects that are characterized by bradycardia, hypotension, and prolongation of the QRS, PR, and QT intervals (TABLE 3). Extreme hypermagnesemia can therefore result in coma, asystole, and death by cardiac arrest. However, hypermagnesemia is rare and until now no genetic causes for it have been identified. Hypermagnesemic patients are often treated by infusion of Ca^{2+} salts to antagonize the cardiac effects of Mg^{2+} (433). Also the efficacy of hemodialysis in the treatment of acute hypermagnesemia has been illustrated (228, 379).

Table 3. Symptoms of Mg^{2+} disturbances

Concentration	Clinical Manifestation
<i>Hypomagnesemia</i>	
<0.7 mM	Neuromuscular irritability
	Hypocalcemia
	Hypokalemia
<0.4 mM	Tetany
	Seizures
	Arrhythmias
<i>Hypermagnesemia</i>	
>1.2 mM	
>2 mM	Lethargy
	Drowsiness
	Flushing
	Nausea and vomiting
	Diminished deep tendon reflex
>3 mM	Somnolence
	Loss of deep tendon reflexes
	Hypotension
	ECG changes
	Complete heart block
>5 M	Cardiac arrest
	Apnea
	Paralysis
	Coma
	Death

1. Drug-induced hypermagnesemia

Only a few drug-induced cases of hypermagnesemia have been reported, and most of them are the direct consequence of the administration of Mg^{2+} itself or Mg^{2+} -containing drugs (TABLE 4).

A) EPSOM SALTS. Epsom salts consist of $MgSO_4$ and are generally used as bath salts or a home remedy against abdominal pain, constipation, and muscle strains. Excessive ingestion of Epsom salts may result in Mg^{2+} overdose and hypermagnesemia. As a result, several fatal cases have been reported in literature (52, 231).

B) CATHARTICS, LAXATIVES, AND ENEMA. Several Mg^{2+} derivatives have been used as cathartics, laxatives, and enemas with the similar goals of softening the stool and easing defecation by increasing their water content. For a long time magnesium citrate [$Mg_3(C_6H_5O_7)_2$] was the most commonly used cathartic. Due to the risk of developing hypermagnesemia and other electrolyte disturbances, polyethylene glycol and electrolyte lavage solutions are currently the first drugs of choice (378, 451). Several Mg^{2+} -containing substances [$Mg_3(C_6H_5O_7)_2$, $Mg(OH)_2$, $MgSO_4$] have been used as laxatives. Mg^{2+} substances elevate the intestinal osmotic pressure, but also act on aquaporin-3 expression and thus increase water permeability (373). Mg^{2+} has been used as a component of enemas to treat constipation. However, both use of Mg^{2+} as a laxative and as an enema may result in fatal hypermagnesemia (413, 521, 586). Therefore, Mg^{2+} administration should be avoided in patients with reduced kidney function, and serum Mg^{2+} should be closely monitored during treatment.

B. Hypomagnesemia

Hypomagnesemia is generally defined as serum Mg^{2+} levels below 0.7 mM. Patients suffer from nonspecific symptoms such as depression, tiredness, muscle spasms, and muscle weakness, and diagnosis therefore may take years (TABLE 3) (177, 523). Only severe Mg^{2+} depletion (<0.4 mM) may lead to cardiac arrhythmias, tetany, and seizures. Secondary to hypomagnesemia, disturbances in K^+ and Ca^{2+} handling are often detected. Hypokalemia can be attributed to increased renal K^+ secretion via ROMK in the connecting tubule (CNT) and collecting duct (CD) (248). Low intracellular Mg^{2+} levels release the Mg^{2+} -dependent inhibition of ROMK channels, resulting in increased renal K^+ secretion. Hypocalcemia can be explained by low PTH levels due to altered activation of the CaSR (574).

Hypomagnesemia is generally treated by oral Mg^{2+} supplementation (± 360 mg/day), although oral Mg^{2+} intake may cause diarrhea at high doses. Intravenous Mg^{2+} supplementation may be more effective, but this treatment has the disadvantage that it requires regular hospital visits. The

treatment regimen of intravenous Mg^{2+} supplementation normally consists of 8–12 g of magnesium sulfate in the first 24 h followed by 4–6 g/day for 3 or 4 days (523). When serum Mg^{2+} levels are extremely low or are accompanied by hypokalemia, Mg^{2+} supplementation may not be sufficient to restore normal Mg^{2+} levels. In that case, patients are often cosupplemented with K^+ or receive amiloride to prevent K^+ secretion.

The following section of this review will focus on the causes of hypomagnesemia. Drug-induced and genetic hypomagnesemia will be distinguished from more general origins of Mg^{2+} deficiency (TABLES 4 AND 5).

1. General causes of hypomagnesemia

A) DIETARY Mg^{2+} INTAKE. Estimations state that up to 60% of American do not meet daily Mg^{2+} requirements (163, 281). Chronic inadequate intake of Mg^{2+} leads to hypomagnesemia. Consumption of Mg^{2+} -rich foods such as kelp,

nuts, green vegetables, and whole grains may prevent this. However, insufficient Mg^{2+} intake may also be caused by pathological conditions such as anorexia nervosa (51).

B) VOMITING AND DIARRHEA. Vomiting and diarrhea may further exacerbate the effects of inadequate Mg^{2+} uptake. Specifically, diarrhea is the consequence of inadequate water reabsorption along the intestine. Since water reabsorption is a prerequisite for Mg^{2+} reabsorption to set the concentration gradient, diarrhea may result in Mg^{2+} deficiency.

C) ALCOHOLISM. Since the early 1960s it has been recognized that alcoholism may cause severe hypomagnesemia (224, 336). Patients suffer from unexplained renal Mg^{2+} wasting, but may also have reduced intestinal Mg^{2+} absorption due to vomiting or diarrhea. In hepatocytes, ethanol completely blocked Mg^{2+} uptake (75). A similar mechanism may take place in the kidney, explaining reduced Mg^{2+} reabsorption. Alcoholics often have reduced PTH levels, which may further contribute to low serum Mg^{2+} levels (7).

Table 4. Drug-induced Mg^{2+} disturbances

Class	Drug	Mechanism	Prevention/Treatment	Reference Nos.
<i>Drug-induced hypomagnesemia</i>				
Diuretics	Furosemide	TAL: reduced paracellular Mg^{2+} reabsorption	Combined K^+ and Mg^{2+} supplementation, Switch to K^+ and Mg^{2+} -sparing diuretics such as amiloride	92, 101, 127, 368
	Thiazide	DCT: reduced TRPM6 expression		
EGFR inhibitors	Cetuximab	DCT: reduced TRPM6 activity	Mg^{2+} supplementation Cetuximab users may switch to erlotinib	341, 459, 510
Proton pump inhibitors	Omeprazole, lansoprazole, pantoprazole, rabeprazole, etc.	Intestine: reduced Mg^{2+} absorption	Mg^{2+} supplementation Switch to histamine2 receptor antagonists	144, 226
Calcineurin inhibitors	Cyclosporin A, tacrolimus	DCT: reduced TRPM6 expression	Mg^{2+} supplementation	39, 517
Platinum derivates	Cisplatin, carboplatinum	DCT: cell death? Reduced TRPM6 expression?	Mg^{2+} supplementation	142, 453, 495, 593
Antimicrobials	AGAs	TAL: reduced paracellular Mg^{2+} reabsorption	Mg^{2+} supplementation	23, 38, 169, 188, 535, 562, 585
	Pentamidine	DCT: cell death?		
	Rapamycin	TAL: reduced paracellular Mg^{2+} reabsorption		
	Amphotericin B	?		
	Foscarnet	Mg^{2+} chelating		
<i>Drug-induced hypermagnesemia</i>				
Epsom salt poisoning	$MgSO_4$	Intestinal Mg^{2+} overload	Hemodialysis	52, 231
Cathartics	$Mg_3(C_6H_5O_7)_2$	Intestinal Mg^{2+} overload	Switch to polyethyleneglycol or electrolyte lavage solutions	378, 451
Laxatives	$MgSO_4$, $Mg(OH)_2$, $Mg_3(C_6H_5O_7)_2$	Intestinal Mg^{2+} overload	Switch to bulk (fiber-based) laxatives	413, 521, 586
Enema	$MgSO_4$	Intestinal Mg^{2+} overload	Switch to fleet (sodium phosphate) enema	

EGFR, epidermal growth factor receptor; TAL, thick ascending limb of Henle's loop; DCT, distal convoluted tubule; TRPM6, transient receptor potential melastatin type 6; AGAs, aminoglycoside antibiotics.

Table 5. Genetic causes of hypomagnesemia

Gene	Protein	Disease	OMIM	Inh.	Segment	Blood Mg ²⁺	Urine Mg ²⁺	Blood Ca ²⁺	Urine Ca ²⁺	Other Symptoms	Reference Nos.
<i>Human genetic Mg²⁺ disorders in TAL</i>											
CLDN16	Claudin 16	FHHNC type 1	248250	R	TAL	↓	–	–	–	Nephrocalcinosis, renal failure	480
CLDN19	Claudin 19	FHHNC type 2	248190	R	TAL	↓	–	–	–	Nephrocalcinosis, renal failure, visual impairment	293
SLC12A1	NKCC2	Bartter type 1	601678	R	TAL	↓	–	–	–	Na ⁺ wasting, hypokalemic alkalosis, high renin/aldosterone	478
KCNJ1	ROMK	Bartter type 2	241200	R	TAL	↓	–	–	–	Na ⁺ wasting, hypokalemic alkalosis, high renin/aldosterone	479
CLCNKB	CIC-Kb	Bartter type 3	607364	R	TAL	↓	–	–	–	Na ⁺ wasting, hypokalemic alkalosis, high renin/aldosterone	477
BSND	Barttin	Bartter type 4	602522	R	TAL	↓	–	–	–	Na ⁺ wasting, hypokalemic alkalosis, high renin/aldosterone	50
<i>Human genetic Mg²⁺ disorders in DCT</i>											
TRPM6	TRPM6	HSH	602014	R	DCT	↓	–	↓	–	Seizures, muscle spasms, mental retardation	454, 543
EGF	EGF	IRH	611718	R	DCT	↓	–	–	–	Seizures, mental retardation	197
CNNM2	CNNM2	HSMR	613882	D/R	DCT	↓	–	–	–	Seizures, mental retardation	497
KCNA1	Kv1.1	ADH	176260	D	DCT	↓	–	–	–	Muscle cramps, tetany, myokymia	176
KCNJ10	Kir4.1	SeSAME/EAST	612780	R	DCT	↓	–	–	↓	Hypokalemia, metabolic alkalosis, sensorineural deafness, seizures, ataxia, mental retardation	54, 458
FXYD2	FXYD2	IDH	154020	D	DCT	↓	–	–	↓	Convulsions	339
HNF1B	HNF1β	RCAD	137920	D	DCT	↓	–	–	↓	Renal cysts, MODY5, renal malformations	11
PCBD1	PCBD1	RCAD-like	264070	R	DCT	↓	–	–	–	Transient hyperphenylalaninemia, MODY5-like	154
SLC12A3	NCC	Gitelman syndrome	263800	R	DCT	↓	–	–	↓	Hypokalemia, metabolic alkalosis, tetany, chondrocalcinosis	481

SLC, solute carrier; NKCC2, Na⁺-K⁺-2Cl⁻ cotransporter; TRPM6, transient receptor potential melastatin type 6; EGF, epidermal growth factor; CNNM2, cyclin M2; FXYD2, FXYD domain containing ion transport regulator 2; HNF1B, hepatocyte nuclear factor 1B; PCBD1, pterin-4-α-carbinolamine dehydratase; NCC, Na⁺-Cl⁻ cotransporter; FHHNC, familial hypomagnesemia with hypercalciuria and nephrocalcinosis; HSH, hypomagnesemia with secondary hypocalcemia; IRH, isolated recessive hypomagnesemia; HSMR, hypomagnesemia with seizures and mental retardation; ADH, autosomal dominant hypomagnesemia; SeSAME, sensorineural deafness, seizures, ataxia, mental retardation, and electrolyte imbalance; EAST, epilepsy, ataxia, sensorineural deafness, and tubulopathy; IDH, isolated dominant hypomagnesemia; RCAD, renal cysts and diabetes; OMIM: online Mendelian inheritance in man; D, dominant; R, recessive; TAL, thick ascending limb of Henle's loop; DCT, distal convoluted tubule; MODY, maturity-onset diabetes of the young.

2. Genetic hypomagnesemia

A) CLDN16. Familial hypomagnesemia with hypercalciuria and nephrocalcinosis type I (FHHNC type I; OMIM 248250) is caused by mutations in claudin 16, previously known as paracellin-1 (480). Patients suffer from renal Mg²⁺ wasting, hypomagnesemia, renal Ca²⁺ wasting, renal parenchymal calcification (nephrocalcinosis), and renal

failure. Serum and urinary Na⁺, K⁺, Cl⁻, and HCO₃⁻ values are initially normal, but may be indirectly affected after progression of renal failure. Sometimes these symptoms are extended to urinary tract infections, kidney stones, and hyperuricemia; Mg²⁺ supplementation is not capable of restoring normal serum Mg²⁺ levels or slowing disease progression (567). A few dozen different mutations have been reported, all characterized by a recessive mode of inheri-

tance (177). All symptoms can be traced to the TAL, the main site for paracellular Ca^{2+} and Mg^{2+} reabsorption. Claudin 16 is part of the tight junction between the cells, and disruption of these tight junctions results in a lack of Ca^{2+} and Mg^{2+} reabsorption in TAL, which can only partially be compensated in the downstream DCT and CNT segments.

B) *CLDN19*. Similar to FHHNC type I, patients with FHHNC type 2 (OMIM 248190) suffer from severe hypomagnesemia accompanied by hypercalciuria and nephrocalcinosis. Additionally, patients have ocular defects consisting of macular colobomata, significant myopia, and horizontal nystagmus. FHHNC type 2 is caused by mutations in claudin 19, which is expressed in the TAL segment of the kidney, in parallel with claudin 16. In the initial publication from Konrad et al. (293), 12 patients from 10 families were genotyped and characterized. Remarkably, 9 of 12 patients developed chronic kidney disease or underwent kidney transplantation. Indeed, other studies confirmed that FHHNC type 2 patients are more prone to developing of CKD and develop the disease at an earlier age compared with type I patients (177). Over the years, several treatment regimens have been proposed, including oral magnesium supplementation, thiazide diuretics, and indomethacin. However, none of these treatments significantly increased serum Mg^{2+} values (177).

C) *TRPM6*. Hypomagnesemia with secondary hypocalcemia (HSH; OMIM 602014) is characterized by extremely low serum Mg^{2+} levels (0.1–0.3 mM) accompanied by low serum Ca^{2+} levels, which result in severe muscular and neurological complications including seizures and mental retardation (454, 543). The disorder was first characterized by Paunier and colleagues in 1968 and later mapped to a region at chromosome 9q in 1997 (390, 544). In 2002, two independent groups identified mutations in *TRPM6* to be causative for HSH (454, 543). *TRPM6* forms the epithelial Mg^{2+} channel responsible for transcellular Mg^{2+} transport in the colon and DCT segment of the kidney (540). Therefore, mutations result in reduced intestinal absorption and renal Mg^{2+} wasting. HSH has an autosomal-recessive mode of inheritance, and currently a few dozen mutations have been found. Patients are generally treated with Mg^{2+} supplements and antiepileptic drugs against seizures. Serum Mg^{2+} levels improve after supplementation, but did not recover to normal levels (297).

D) *EGF*. Isolated autosomal recessive hypomagnesemia (IRH; OMIM 611718) is caused by mutations in the *EGF* gene (197). In a consanguine family from Dutch origin, two sisters presented with serum Mg^{2+} levels of 0.53 and 0.56 mM and urinary Mg^{2+} values of 3.9 and 3.7 mmol/24 h, respectively (173). Serum Ca^{2+} , Na^+ , K^+ , Cl^- , HCO_3^- , and blood pH values were normal. The patients presented with epileptic seizures during their first year of life, which could be

controlled by antiepileptic drugs. Moreover, psychomotor retardation was observed in these patients. Plasma renin activity, plasma aldosterone, and parathyroid hormone concentrations were in the normal range. Homozygosity mapping and subsequent Sanger sequencing of gene candidates led to the identification of a homozygous c.C3209T mutation in exon 22 resulting a p.P1070L missense mutation at protein level (197). This residue is particularly important for plasma membrane targeting of the EGF molecule and the mutation results in impaired basolateral sorting of pro-EGF. Therefore, *TRPM6* activity is not stimulated, resulting in renal Mg^{2+} wasting (197, 515). Until now, only a single family has been described with EGF mutations, but studies with EGFR antagonists further underline the clinical importance of EGF for renal Mg^{2+} handling (discussed in detail in sect. VB3).

E) *KCNA1*. In a Brazilian family, mutations in *KCNA1* encoding voltage-gated K^+ channel Kv1.1. cause autosomal dominant hypomagnesemia (OMIM 176260) (176). The patient presented in the clinic with muscle cramps, muscle weakness, tetanic episodes, and tremor. Serum Mg^{2+} values were low (0.37 mM), whereas other electrolytes and metabolites including Na^+ , K^+ , Ca^{2+} , P_i , uric acid, bicarbonate, urea, creatinine, glucose, bilirubin, aminotransferases, alkaline phosphate, and lactate dehydrogenase were all normal. Urinary creatinine clearance as well as Mg^{2+} and Ca^{2+} excretion were within normal range. *KCNA1* mutations were previously linked to ataxia and myokymia (62, 146). Therefore, a cerebral MRI was performed in these patients, showing slight atrophy of the cerebral vermis. Family members suffer from myokymic discharge in electromyograph analysis, which is in line with the previously observed mixed phenotype. Intravenous Mg^{2+} infusion improved the clinical symptoms. Kv1.1 has been proposed to cause apical hyperpolarization, which allows the uptake of Mg^{2+} via *TRPM6*. The p.N255D (c.A763G) mutation identified in the Brazilian family disrupts Kv1.1 activity and therefore may reduce the driving force for Mg^{2+} transport. Although many *KCNA1* mutations have been reported, even in residues very close to the p.N255, none of these has yet been associated with hypomagnesemia, even though Kv1.1 function is impaired (12, 263). Identification of additional hypomagnesemic families with Kv1.1 mutations may aid our understanding of Kv1.1 function in DCT. It has been suggested that other factors contribute to the apical membrane potential and may compensate for a loss of Kv1.1 function; ROMK may be one of the compensatory factors (104, 140).

F) *CNNM2*. Mutations in *CNNM2* are causative for hypomagnesemia with seizures and mental retardation (HSMR; OMIM 613882). Two unrelated families with seizures and dominant hypomagnesemia were reported to carry *CNNM2* mutations (497). In these patients, serum Mg^{2+} levels range between 0.3 and 0.5 mM, but no other electrolyte disturbances were detected. The patients' symptoms

include seizures, loss of consciousness, loss of muscle tone, headaches, and staring (340). Recently, five additional families were reported (28), making CNNM2 the most common genetic cause of isolated hypomagnesemia after TRPM6 and CLDN16–19. Interestingly, in this new cohort CNNM2 mutations were linked to a phenotype of impaired brain development and mental retardation. This intellectual disability was most prominent in a family with a recessive pattern of inheritance, emphasizing the heterogeneous inheritance of CNNM2 depending on the location and severity of the mutations. HSMR patients are treated with anti-epileptic drugs and Mg^{2+} supplements. Serum Mg^{2+} levels improved after supplementation, but did not reach normal levels. Although the exact function of CNNM2 remains to be elucidated, mutations can disrupt the MgATP binding domain and reduce CNNM2 membrane expression (28, 105). Recent data from patch-clamp and Mg^{2+} uptake experiments favor the hypothesis that CNNM2 does not transport Mg^{2+} itself, but rather regulates other Mg^{2+} -transporting proteins (28, 105, 497).

G) *KCNJ10*. Mutations in *KCNJ10* encoding the Kir4.1 K^+ channel can cause seizures, sensorineural deafness, ataxia, mental retardation, and electrolyte imbalance/epilepsy, ataxia, sensorineural deafness, and renal tubulopathy syndrome (SeSAME/EAST; OMIM 612780) (54, 417, 458). Patients were reported to have marked electrolyte abnormalities, including hypokalemic metabolic alkalosis without hypertension, severe hypomagnesemia, and renal Na^+ , K^+ , and Mg^{2+} wasting. In some patients, high renin and aldosterone levels, salt craving, and polyuria were observed. Kir4.1 is hypothesized to be involved in K^+ recycling at the basolateral membrane of DCT cells. When Kir4.1 is mutated, K^+ availability becomes rate limiting for Na^+ - K^+ -ATPase activity. Thus the Na^+ - K^+ -ATPase will be inhibited, resulting in a reduced potential across the basolateral membrane. Consequently, Na^+ and Mg^{2+} transport will be reduced in DCT. To compensate for this, ENaC activity in CNT will be increased at the expense of K^+ excretion via ROMK. Therefore, SeSAME/EAST patients suffer from severe hypomagnesemia and hypokalemia. To treat the hypomagnesemia, patients are often given Mg^{2+} and K^+ supplements, in combination with aldosterone antagonists or ENaC inhibitors (34). SeSAME/EAST patients suffer from a severe neurological phenotype consisting of tonic-clonic seizures in infancy, cerebellar ataxia, and hearing loss. Moreover, magnetic resonance imaging evidenced subtle symmetrical signal changes in the cerebellar dentate nuclei (96).

H) *FXD2*. Gene linkage studies identified the FXD domain containing ion transport regulator 2 (*FXD2*) mutations in a family with dominant isolated renal Mg^{2+} wasting (IDH; OMIM 154020) (339). The patients in this family presented with low serum Mg^{2+} values (± 0.4 mM), while other plasma electrolytes, including Na^+ , K^+ , Ca^{2+} , Cl^- , and

HCO_3^- , were normal. Urinary Mg^{2+} excretion was increased, whereas Ca^{2+} excretion was slightly lowered (172). The c.G121A mutation results in a p.G41R missense mutation at the protein level, which causes misrouting of *FXD2* to the membrane (66). *FXD2* encodes for the γ -subunit of the Na^+ - K^+ -ATPase. Although the exact role of *FXD2* in the DCT is unknown, it has been hypothesized to stabilize the Na^+ - K^+ -ATPase, influencing the membrane potential necessary for Mg^{2+} transport (345). However, other reports suggest that it may function independently as an inward rectifier channel (464). Functional analysis of the patient's proximal tubular cells showed no differences in Na^+ , K^+ , or ATP affinity of the Na^+ - K^+ -ATPase, but demonstrated a lower *FXD2* protein expression (67).

I) *HNF1B*. Renal cysts and diabetes syndrome (RCAD; OMIM 137920) is caused by mutations in hepatocyte nuclear factor 1 β (*HNF1B*) and consists of a heterogeneous group of symptoms including renal cysts ($\pm 70\%$ of patients), maturity onset diabetes of the young subtype 5 (MODY5; $\pm 50\%$), and hypomagnesemia ($\pm 45\%$) (11, 82). *HNF1B* is a transcription factor regulating gene expression in kidney development (331). *FXD2b* expression is one of several genes that is regulated by *HNF1B* (155), which may explain its role in renal Mg^{2+} handling. However, the possibility that *HNF1B* regulates other DCT genes involved in renal Mg^{2+} transport cannot be excluded.

J) *PCBD1*. In a small cohort of three patients, mutations in pterin-4 α -carbinolamine dehydratase 1 (*PCBD1*) have been linked to hypomagnesemia, renal Mg^{2+} wasting, and MODY5-like diabetes (154). *PCBD1* mutations are known to cause transient neonatal hyperphenylalaninemia and high urinary levels of primapterin (HPABH4D; OMIM 264070) (518, 519). HPABH4D patients are diagnosed at birth by Guthrie testing and suffer from a transient, benign defect in impaired BH4 regeneration. A follow-up study of three patients at ± 18 yr of age showed that the patients display a mild hypomagnesemia (± 0.6 mM) and MODY5-like diabetes, but no renal cysts (154). Interestingly, serum and urinary Na^+ , K^+ , Ca^{2+} , and Cl^- levels were within the normal range. The phenotype of the HPABH4D patients resembles that of RCAD patients, and the treatment regime consists of sulfonyleureas and Mg^{2+} supplements. The origin of renal cysts in RCAD patients may be traced to the CD where *HNF1B* regulates *PKHD1* (194). However, since *PCBD1* is not expressed in CD, HPABH4D patients are protected from cyst formation (154).

K) *SLC12A3*. Hypomagnesemia and hypokalemia are the cardinal symptoms of a hereditary electrolyte disorder characterized by Dr. Gitelman in 1969 that has been known since as Gitelman's syndrome (175). Patients present with tetany, paresthesias, and chondrocalcinosis (284). The severity of the symptoms depends on the degree of hypokalemia. Except hypokalemia and hypomagnesemia, laboratory inves-

tigations often show metabolic alkalosis and hypocalciuria, sometimes associated with a mild hypotension and prolonged QT interval. *SLC12A3* encodes the thiazide-sensitive $\text{Na}^+\text{-Cl}^-$ cotransporter (NCC), and mutations here cause Gitelman's syndrome (165, 481). Patients with Gitelman's syndrome are often treated with oral Mg^{2+} supplements (284). Interestingly, in some patients Mg^{2+} supplementation restores normal K^+ levels, suggesting that hypokalemia is secondary to hypomagnesemia (205). This hypothesis is further substantiated by the NCC KO mouse, which is hypomagnesemic but does not display K^+ disturbances under basal conditions (460). NCC KO mice have markedly reduced *TRPM6* expression levels, possibly explaining the renal Mg^{2+} wasting observed in Gitelman's syndrome (368). However, the mechanism by which a loss of NCC function results in reduced *TRPM6* expression remains unresolved. It has been suggested that the atrophy of the DCT segment observed in KO mice may partially explain this phenomenon (321).

L) *SLC12A1*, *BSND*, *CLCNKB*, AND *KCNJ1*. Bartter's syndrome was originally described by Dr. Bartter in 1962 and is characterized by salt wasting, hypokalemic alkalosis, elevated plasma renin and aldosterone levels, and low blood pressure (40). Mutations in *SLC12A1*, encoding NKCC2, *Barttin*, *ClC-Kb*, *KCNJ1*, encoding ROMK, or *CaSR* form the genetic basis of Bartter's syndrome (50, 477–479). Mild hypomagnesemia is sometimes observed in Bartter's patients, which may be explained by a reduced driving force for paracellular Mg^{2+} reabsorption in the TAL. Compensation for reduced TAL Mg^{2+} reabsorption may take place in the DCT, which explains why Bartter's patients often have normal Mg^{2+} levels. *ClC-Kb* and *Barttin* are also expressed in DCT, which justifies why patients with mutations in these genes more often show hypomagnesemia (261).

3. Drug-induced hypomagnesemia

A) DIURETICS. Hypomagnesemia has been associated with diuretics targeting the TAL and DCT segments of the kidney. In 1968, Duarte (127) reported renal Ca^{2+} and Mg^{2+} wasting as a consequence of furosemide treatment. Furosemide inhibits the activity of NKCC2, reducing the positive transepithelial membrane potential that drives paracellular Mg^{2+} transport in TAL (408). Although the incidence of furosemide-induced hypomagnesemia is unclear, a significant number of patients who use it may suffer from Mg^{2+} wasting (92, 330). In a recent animal study, furosemide treatment did not result in hypomagnesemia since increased *TRPM6* expression in the DCT was able to compensate for reduced Mg^{2+} reabsorption in TAL (530). The clinical effects of furosemide treatment on Mg^{2+} levels may, therefore, depend on the individual's ability to compensate at DCT level.

The use of thiazide diuretics, which target NCC in DCT, frequently induces renal Mg^{2+} wasting (366). Although most studies do not report hypomagnesemia in patients being treated with thiazide diuretics, some patient groups may be at risk (101). Patients with low initial Mg^{2+} values, such as elderly patients or patients with chronic heart failure, may develop hypomagnesemia after chronic thiazide treatment (238, 285). In mice, thiazide treatment reduces the renal expression of *TRPM6*, explaining the high urinary Mg^{2+} excretion (368). However, in most patients these effects may be small and the clinical consequences may depend on the patients' basal serum Mg^{2+} values.

V) EGFR INHIBITORS. In 2005, it was first reported that the use of the EGFR inhibitor cetuximab can result in severe hypomagnesemia (459). Cetuximab (erbitux) is a monoclonal antibody against the EGFR and is generally prescribed for the treatment of colorectal or head and neck cancer. Up to 50–60% of patients using cetuximab may develop, and 10–20% display serum levels below 0.4 mM (341, 510). A recent meta-analysis showed a RR of 3.87 (396). Immediately after the first reports of cetuximab-induced hypomagnesemia, questions were raised whether the EGFR inhibitor erlotinib (tarceva) may have similar effects (16). However, as of this writing, no clinical reports on the Mg^{2+} status after the use of erlotinib are available. Animal studies demonstrated a small reduction in serum Mg^{2+} values after erlotinib administration (122). Erlotinib is generally administered as a tablet that also contains Mg^{2+} stearate. The copresence of Mg^{2+} in these tablets may explain the absence of clinical consequences of erlotinib administration on serum Mg^{2+} levels.

C) PROTON PUMP INHIBITORS. In 2006, the use of protein pump inhibitors (PPI) was associated with hypomagnesemia for the first time in two separate patients receiving long-term omeprazole treatment (144). Since then, many new cases of PPI-induced hypomagnesemia have been reported (295). A recent systematic review of 36 cases demonstrated that discontinuation of PPIs resulted in recovery from hypomagnesemia within 4 days, and rechallenge led to reoccurrence within 4 days (226). Urinary Mg^{2+} excretion is low in these patients, suggesting normal kidney function and thus an effect of PPI use on intestinal Mg^{2+} absorption. In mice, administration of omeprazole increased the expression of *TRPM6* in colon (300). Therefore, it was hypothesized that omeprazole may inhibit the activity of the colonic $\text{H}^+\text{-K}^+\text{-ATPase}$, resulting in reduced extrusion of protons into the colon. Since *TRPM6* activity increases at lower external pH, decreased proton secretion may reduce *TRPM6* activity, for which increased *TRPM6* expression may compensate (300, 313). However, increased *TRPM6* expression may not be sufficient to prevent malabsorption of Mg^{2+} in colon in all patients. Individual variability of this compensatory mechanism may explain why only a subset of PPI users develop hypomagnesemia.

D) CALCIINEURIN INHIBITORS. The calcineurin inhibitors (CNI) cyclosporin A (CsA) and tacrolimus (FK506) are currently the first immunosuppressant drugs of choice after transplantation. The use of CNIs has been associated with hypertension and renal Mg^{2+} wasting (39, 517). Whereas the hypertension may be explained by an increased activity of NCC, renal Mg^{2+} wasting is not fully understood (242). Up to 90% of all patients suffer from significantly reduced serum Mg^{2+} levels after initiation of CsA treatment, and in a recent cohort even 35% of patients remained hypomagnesemic despite Mg^{2+} supplementation (39, 439, 517). In rats, CsA and FK506 treatment increased renal Mg^{2+} wasting and hypomagnesemia (29, 365). In FK506-treated rats *TRPM6* mRNA is downregulated, resulting in dramatically increased Mg^{2+} excretion (365). In a recent study with CsA treatment in rats, *TRPM6*, *TRPM7*, and *EGF* mRNA expression were reduced, although the fractional excretion of Mg^{2+} did not significantly change (306). Interestingly, EGF treatment did not change Mg^{2+} excretion and *TRPM6* expression in CsA-treated rats, whereas it reduced Mg^{2+} wasting and increased *TRPM6* expression in control rats. These results suggest that CsA may interfere with the EGF signaling pathway in DCT cells. Patients receiving CNI treatment are generally supplemented with Mg^{2+} to prevent hypomagnesemia.

E) CISPLATIN/CARBOPLATIN. Already from the introduction of cisplatin (*cis*-diamminedichloridoplatinum) as anti-cancer therapeutic, hypomagnesemia has been reported in ~40%-80% of treated patients (453, 593). Nephrotoxicity is a common side effect of cisplatin treatment, mainly as a consequence of proximal tubule cisplatin accumulation, which results in necrosis of the tubular cells (298). However, the effect of cisplatin on electrolyte wasting is highly specific for Mg^{2+} ; concomitant Ca^{2+} and K^{+} wasting is only observed in severely hypomagnesemic patients. This suggests that hypomagnesemia cannot be explained by the nephrotoxicity and that Ca^{2+} and K^{+} disturbances are secondary to Mg^{2+} wasting. Treatment with carboplatin [paraplatin, *cis*-diammine(1,1-cyclobutanedicarboxylato)platinum], another platinum derivate, results in similar side effects including hypomagnesemia (142, 495). Recently, two animal studies have examined the effects of cisplatin treatment in detail (305, 529). Both observe significant downregulation of *TRPM6* mRNA levels, although the causative mechanisms of reduced *TRPM6* expression may differ. In the mice study of Van Angelen et al. (529), all DCT markers including parvalbumin and NCC are reduced, suggesting that cisplatin treatment induced atrophy of the DCT segment. In rats, Ledeganck et al. (305) demonstrated no effects on NCC expression, which implies that the DCT cells are still intact. Nevertheless, *TRPM6* and *EGF* expression were reduced. Both studies show compensation of Mg^{2+} uptake in the TAL by increased expression of claudins. Generally, patients that develop hypomagnesemia during cisplatin

treatment are supplemented by adding Mg^{2+} to pre- and posthydration fluids to prevent hypomagnesemia.

F) ANTIMICROBIALS. Although several classes of antimicrobials may cause hypomagnesemia, the underlying mechanisms leading to Mg^{2+} wasting differ greatly. Aminoglycoside antibiotics (AGA) including gentamycin, neomycin, tobramycin, and amikacin may induce renal Mg^{2+} wasting (562, 585). Estimates of the incidence of hypomagnesemia as a consequence of AGA use range from 20 to 80% (157, 562). AGAs may activate the CaSR, resulting in reduced paracellular Mg^{2+} transport in the TAL and inhibition of Mg^{2+} transport in the DCT. Studies in MDCT cells show reduced PTH-activated Mg^{2+} transport (269). Moreover, animal studies have evidenced that use of AGAs cause hypomagnesemia, due to reduced expression of NKCC2 that provides the driving force for TAL Mg^{2+} transport (167, 446). In a recent study with gentamycin-treated rats, *TRPM6* expression was upregulated, suggesting that the DCT compensates for reduced TAL Mg^{2+} reabsorption (307).

Pentamidine is an antimicrobial against *Pneumocystis jirovecii* infections that are often diagnosed in AIDS patients. The use of pentamidine has been associated with severe hypomagnesemia due to renal Mg^{2+} wasting at the start of the 1990s (65, 188, 465). The exact mechanism of reduced Mg^{2+} reabsorption remains unresolved. However, pentamidine reduces ENaC activity, resulting in hyperkalemia (283). Moreover, there have been reports of tubular necrosis after pentamidine treatment (546), which may cause atrophy of the DCT segment.

Rapamycin (sirolimus) is an antibiotic that is frequently used to prevent organ rejection after transplantation. Rapamycin inhibits mTOR activity, and its use has been associated with hypomagnesemia in 10–25% of patients (23, 535). Rapamycin-treated rats exhibit reduced expression of NKCC2 (100). Interestingly, *TRPM6* expression was increased in the same study. This could be compensation for the reduced Mg^{2+} uptake in TAL, but a direct effect of rapamycin on *TRPM6* expression cannot be excluded. A recent in vitro study showed the opposite effect; rapamycin decreased *TRPM6* expression in an EGF-dependent manner (255).

Amphotericin B is an antifungal agent that has been associated with hypomagnesemia and hypokalemia (38). The mechanism underlying urinary Mg^{2+} wasting in these patients is unknown. Oral Mg^{2+} supplementation together with amiloride treatment is generally used to restore Mg^{2+} levels (178).

Foscarnet inhibits viral DNA polymerases by chelating divalent cations and, therefore, its use may cause hypomagnesemia (169). Patients also suffer from hypocalcemia and

hypokalemia, which may be secondary to the Mg^{2+} disturbances (253, 369). Until now, no studies have examined the effect of foscarnet on the expression of renal ion transporters. It would be interesting to examine whether foscarnet exhibits effects beyond its chelating function.

VI. CONCLUDING REMARKS

Over the last decade, Mg^{2+} has been considered as a treatment for several major diseases including preeclampsia, stroke, myocardial infarction, and asthma in several large-scale clinical trials. These findings have raised interest in Mg^{2+} among neurologists, cardiologists, and pneumologists. Nevertheless, Mg^{2+} levels are still not determined routinely in daily clinical practice, even though up to 60% of all critically ill patients are Mg^{2+} deficient (84, 145). Serum Mg^{2+} should be determined standardly, alongside Na^+ , K^+ , and Ca^{2+} measurements in patients. Mg^{2+} disturbances can cause muscle cramps, arrhythmias, and seizures, and therefore, Mg^{2+} should be considered when patients present in clinic with these symptoms. However, the molecular mechanisms underlying the effects of Mg^{2+} in brain, heart, and lung are still largely unknown. Finding the molecular basis of the role of Mg^{2+} in these diseases may further extend the significance of Mg^{2+} therapies in clinic.

Genetic and drug-induced disorders of Mg^{2+} homeostasis have enhanced the knowledge on Mg^{2+} (re)absorption in the kidney and intestine. These studies form a perfect example of the powerful interaction of clinical and fundamental studies. For instance, the increased knowledge on the role of EGF in renal Mg^{2+} handling has resulted in the standardization of Mg^{2+} measurements in patients using EGFR blockers. This resulted in early detection of Mg^{2+} disturbances and changes in treatment strategies. Through synergistic clinical and fundamental efforts in the fields of brain, heart, and lung Mg^{2+} research, the unexplained role of Mg^{2+} in, among others, migraine, depression, epilepsy, COPD, and hypertension, may be elucidated. Over the last decades Mg^{2+} research has been centered around the kidney and the intestine. By extending the field to the heart, brain, and lungs and by involving both fundamental and clinical researchers, Mg^{2+} will never be thought of as “a forgotten cation” anymore.

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REFERENCES

1. Institute of Medicine (US) Standing Committee on the Scientific Evaluation of Dietary Reference Intakes. *Dietary Reference Intakes for Calcium, Phosphorus, Magnesium, Vitamin D, and Fluoride*. Washington, DC: National Academies Press, 1997.
2. Early administration of intravenous magnesium to high-risk patients with acute myocardial infarction in the Magnesium in Coronaries (MAGIC) Trial: a randomised controlled trial. *Lancet* 360: 1189–1196, 2002.
3. Expert Panel Report 3 (E.P.R.-3): Guidelines for the Diagnosis, and Management of Asthma-Summary Report 2007. *J Allergy Clin Immunol* 120: S94–138, 2007.
4. ISIS4: a randomised factorial trial assessing early oral captopril, oral mononitrate, and intravenous magnesium sulphate in 58,050 patients with suspected acute myocardial infarction. ISIS-4 (Fourth International Study of Infarct Survival). *Collaborative Group Lancet* 345: 669–685, 1995.
5. The Magpie Trial: a randomised trial comparing magnesium sulphate with placebo for pre-eclampsia. Outcome for children at 18 months. *Br J Obstet Gynaecol* 114: 289–299, 2007.
6. The Magpie Trial: a randomised trial comparing magnesium sulphate with placebo for pre-eclampsia. Outcome for women at 2 years. *Br J Obstet Gynaecol* 114: 300–309, 2007.
7. Abbott L, Nadler J, Rude RK. Magnesium deficiency in alcoholism: possible contribution to osteoporosis and cardiovascular disease in alcoholics. *Alcoholism Clin Exp Res* 18: 1076–1082, 1994.
8. Abbott RD, Ando F, Masaki KH, Tung KH, Rodriguez BL, Petrovitch H, Yano K, Curb JD. Dietary magnesium intake and the future risk of coronary heart disease (the Honolulu Heart Program). *Am J Cardiol* 92: 665–669, 2003.
9. Abraham AS, Eylath U, Weinstein M, Czaczkes E. Serum magnesium levels in patients with acute myocardial infarction. *N Engl J Med* 296: 862–863, 1977.
10. Abraham GE. The importance of magnesium in the management of primary postmenopausal osteoporosis. *J Nutr Environ Med* 2: 165–178, 1991.
11. Adalat S, Woolf AS, Johnstone KA, Wirsing A, Harries LW, Long DA, Hennekam RC, Ledermann SE, Rees L, van't Hoff W, Marks SD, Trompeter RS, Tullus K, Winyard PJ, Cansick J, Mushtaq I, Dhillon HK, Bingham C, Edghill EL, Shroff R, Stanescu H, Ryffel GU, Ellard S, Bockenhauer D. HNF1B mutations associate with hypomagnesemia and renal magnesium wasting. *J Am Soc Nephrol* 20: 1123–1131, 2009.
12. Adelman JP, Bond CT, Pessia M, Maylie J. Episodic ataxia results from voltage-dependent potassium channels with altered functions. *Neuron* 15: 1449–1454, 1995.
13. Alderton A, Davies P, Illman K, Brown DR. Ancient conserved domain protein-1 binds copper and modifies its retention in cells. *J Neurochem* 103: 312–321, 2007.
14. Alfrey AC, Miller NL, Trow R. Effect of age and magnesium depletion on bone magnesium pools in rats. *J Clin Invest* 54: 1074–1081, 1974.
15. Allouche D, Parello J, Sanejouand YH. Ca^{2+}/Mg^{2+} exchange in parvalbumin and other EF-hand proteins. A theoretical study. *J Mol Biol* 285: 857–873, 1999.

16. Altundag K, Altundag O, Baptista MZ, Turen S, Atik MA. Re: Cetuximab therapy and symptomatic hypomagnesemia. *J Natl Cancer Inst* 97: 1791–1792, 2005.
17. Altura BM, Gebrewold A, Zhang A, Altura BT. Low extracellular magnesium ions induce lipid peroxidation and activation of nuclear factor-kappa B in canine cerebral vascular smooth muscle: possible relation to traumatic brain injury and strokes. *Neurosci Lett* 341: 189–192, 2003.
18. Altura BT, Altura BM. Endothelium-dependent relaxation in coronary arteries requires magnesium ions. *Br J Pharmacol* 91: 449–451, 1987.
19. Altura BT, Memon ZI, Zhang A, Cheng TP, Silverman R, Cracco RQ, Altura BM. Low levels of serum ionized magnesium are found in patients early after stroke which result in rapid elevation in cytosolic free calcium and spasm in cerebral vascular muscle cells. *Neurosci Lett* 230: 37–40, 1997.
20. Amasheh S, Fromm M, Gunzel D. Claudins of intestine and nephron: a correlation of molecular tight junction structure and barrier function. *Acta Physiol* 201: 133–140, 2011.
21. Amin M, Abdel-Fattah M, Zaghoul SS. Magnesium concentration in acute asthmatic children. *Iranian J Pediatr* 22: 463–467, 2012.
22. Anastassopoulou J, Theophanides T. Magnesium-DNA interactions and the possible relation of magnesium to carcinogenesis. Irradiation and free radicals. *Crit Rev Oncol/Hematol* 42: 79–91, 2002.
23. Andoh TF, Burdman EA, Fransechini N, Houghton DC, Bennett WM. Comparison of acute rapamycin nephrotoxicity with cyclosporine and FK506. *Kidney Int* 50: 1110–1117, 1996.
24. Angelow S, El-Husseini R, Kanzawa SA, Yu AS. Renal localization and function of the tight junction protein, claudin-19. *Am J Physiol Renal Physiol* 293: F166–F177, 2007.
25. Arango MF, Bainbridge D. Magnesium for acute traumatic brain injury. *Cochrane Database Systematic Rev* CD005400, 2008.
26. Ariza AC, Bobadilla N, Diaz L, Avila E, Larrea F, Halhali A. Placental gene expression of calcitonin gene-related peptide and nitric oxide synthases in preeclampsia: effects of magnesium sulfate. *Magnesium Res* 22: 44–49, 2009.
27. Ariza AC, Ponce X, Gonzalez-Gonzalez ME, Larrea F, Halhali A. Effects of magnesium sulphate on placental expression of endothelin 1 and its receptors in preeclampsia. *Clin Biochem* 40: 976–980, 2007.
28. Arjona FJ, De Baaij JHF, Schlingmann KP, Lameris ALL, Van Wijk E, Flik G, Regele S, Korenke GC, Nephytou B, Rust S, Reintjes N, Konrad M, Bindels RJM, Hoenderop JGJ. CNNM2 Mutations cause Impaired Brain Development and Seizures in Patients with Hypomagnesemia. *PLoS Genet* 10: e1004267, 2014.
29. Asai T, Nakatani T, Yamanaka S, Tamada S, Kishimoto T, Tashiro K, Nakao T, Okamura M, Kim S, Iwao H, Miura K. Magnesium supplementation prevents experimental chronic cyclosporine a nephrotoxicity via renin-angiotensin system independent mechanism. *Transplantation* 74: 784–791, 2002.
30. Ascherio A, Hennekens C, Willett WC, Sacks F, Rosner B, Manson J, Witteman J, Stampfer MJ. Prospective study of nutritional factors, blood pressure, and hypertension among US women. *Hypertension* 27: 1065–1072, 1996.
31. Aziz HS, Blamoun AI, Shubair MK, Ismail MM, DeBari VA, Khan MA. Serum magnesium levels and acute exacerbation of chronic obstructive pulmonary disease: a retrospective study. *Ann Clin Lab Sci* 35: 423–427, 2005.
32. Bairoch A. The ENZYME database in 2000. *Nucleic Acids Res* 28: 304–305, 2000.
33. Ban C, Junop M, Yang W. Transformation of MutL by ATP binding and hydrolysis: a switch in DNA mismatch repair. *Cell* 97: 85–97, 1999.
34. Bandulik S, Schmidt K, Bockenhauer D, Zdebik AA, Humberg E, Kleta R, Warth R, Reichold M. The salt-wasting phenotype of EAST syndrome, a disease with multifaceted symptoms linked to the KCNJ10 K⁺ channel. *Pflügers Arch* 461: 423–435, 2011.
35. Banki CM, Vojnik M, Papp Z, Balla KZ, Arato M. Cerebrospinal fluid magnesium and calcium related to amine metabolites, diagnosis, and suicide attempts. *Biol Psychiatry* 20: 163–171, 1985.
36. Barbagallo M, Dominguez LJ. Magnesium metabolism in type 2 diabetes mellitus, metabolic syndrome and insulin resistance. *Arch Biochem Biophys* 458: 40–47, 2007.
37. Barragan-Rodriguez L, Rodriguez-Moran M, Guerrero-Romero F. Efficacy and safety of oral magnesium supplementation in the treatment of depression in the elderly with type 2 diabetes: a randomized, equivalent trial. *Magnesium Res* 21: 218–223, 2008.
38. Barton CH, Pahl M, Vaziri ND, Cesario T. Renal magnesium wasting associated with amphotericin B therapy. *Am J Med* 77: 471–474, 1984.
39. Barton CH, Vaziri ND, Martin DC, Choi S, Alikhani S. Hypomagnesemia and renal magnesium wasting in renal transplant recipients receiving cyclosporine. *Am J Med* 83: 693–699, 1987.
40. Bartter FC, Pronove P, Gill JR Jr, Maccardle RC. Hyperplasia of the juxtaglomerular complex with hyperaldosteronism and hypokalemic alkalosis. A new syndrome. *Am J Med* 33: 811–828, 1962.
41. Barzilay G, Mol CD, Robson CN, Walker LJ, Cunningham RP, Tainer JA, Hickson ID. Identification of critical active-site residues in the multifunctional human DNA repair enzyme HAP1. *Nature Struct Biol* 2: 561–568, 1995.
42. Bates-Withers C, Sah R, Clapham DE. TRPM7, the Mg²⁺ inhibited channel and kinase. *Adv Exp Med Biol* 704: 173–183, 2011.
43. Bednarek A, Pasternak K, Karska M. Evaluation of blood serum, erythrocyte and urine magnesium concentrations in babies with pneumonia or bronchial obstructive bronchitis. *Magnesium Res* 16: 271–280, 2003.
44. Belge H, Gailly P, Schwaller B, Loffing J, Debaix H, Riveira-Munoz E, Beauwens R, Devogelaer JP, Hoenderop JG, Bindels RJ, Devuyst O. Renal expression of parvalbumin is critical for NaCl handling and response to diuretics. *Proc Natl Acad Sci USA* 104: 14849–14854, 2007.
45. Ben-Yosef T, Belyantseva IA, Saunders TL, Hughes ED, Kawamoto K, Van Itallie CM, Beyer LA, Halsey K, Gardner DJ, Wilcox ER, Rasmussen J, Anderson JM, Dolan DF, Forge A, Raphael Y, Camper SA, Friedman TB. Claudin 14 knockout mice, a model for autosomal recessive deafness DFNB29, are deaf due to cochlear hair cell degeneration. *Hum Mol Genet* 12: 2049–2061, 2003.
46. Bhatt SP, Khandelwal P, Nanda S, Stoltzfus JC, Fioravanti GT. Serum magnesium is an independent predictor of frequent readmissions due to acute exacerbation of chronic obstructive pulmonary disease. *Respir Med* 102: 999–1003, 2008.
47. Bigal ME, Bordini CA, Tepper SJ, Speciali JG. Intravenous magnesium sulphate in the acute treatment of migraine without aura and migraine with aura. A randomized, double-blind, placebo-controlled study. *Cephalgia* 22: 345–353, 2002.
48. Bigal ME, Walter S, Rapoport AM. Calcitonin gene-related peptide (CGRP) and migraine current understanding and state of development. *Headache* 53: 1230–1244, 2013.
49. Bilbey DL, Prabhakaran VM. Muscle cramps and magnesium deficiency: case reports. *Canadian Family Physician* 42: 1348–1351, 1996.
50. Birkenhager R, Otto E, Schurmann MJ, Vollmer M, Ruf EM, Maier-Lutz I, Beekmann F, Fekete A, Omran H, Feldmann D, Milford DV, Jeck N, Konrad M, Landau D, Knoers NV, Antignac C, Sudbrak R, Kispert A, Hildebrandt F. Mutation of BSND causes Bartter syndrome with sensorineural deafness and kidney failure. *Nature Genet* 29: 310–314, 2001.
51. Birmingham CL, Gritzner S. Heart failure in anorexia nervosa: case report and review of the literature. *Eating Weight Disorders* 12: e7–10, 2007.
52. Birrer RB, Shallash AJ, Totten V. Hypermagnesemia-induced fatality following epsom salt gargles(1). *J Emergency Med* 22: 185–188, 2002.
53. Blackfan KD, Hamilton B. Uremia in acute glomerular nephritis: the cause and treatment in children. *Boston Med Surg J* 193: 617–621, 1925.
54. Bockenhauer D, Feather S, Stanescu HC, Bandulik S, Zdebik AA, Reichold M, Tobin J, Lieberer E, Sterner C, Landouere G, Arora R, Sirimanna T, Thompson D, Cross JH, van't Hoff W, Al Masri O, Tullus K, Yeung S, Anikster Y, Klootwijk E, Hubank M, Dillon MJ, Heitzmann D, Arcos-Burgos M, Knepper MA, Dobbie A, Gahl WA, Warth R, Sheridan E, Kleta R. Epilepsy, ataxia, sensorineural deafness, tubulopathy, and KCNJ10 mutations. *N Engl J Med* 360: 1960–1970, 2009.
55. Boyman L, Mikhasenko H, Hiller R, Khananavili D. Kinetic and equilibrium properties of regulatory calcium sensors of NCX1 protein. *J Biol Chem* 284: 6185–6193, 2009.
56. Brautigam CA, Steitz TA. Structural and functional insights provided by crystal structures of DNA polymerases and their substrate complexes. *Curr Opin Struct Biol* 8: 54–63, 1998.

57. Breiderhoff T, Himmerkus N, Stuiver M, Mutig K, Will C, Meij IC, Bachmann S, Bleich M, Willnow TE, Muller D. Deletion of claudin-10 (Cldn10) in the thick ascending limb impairs paracellular sodium permeability and leads to hypermagnesemia and nephrocalcinosis. *Proc Natl Acad Sci USA* 109: 14241–14246, 2012.
58. Breukels V, Konijnenberg A, Nabuurs SM, Touw WG, Vuister GW. The second Ca^{2+} -binding domain of NCX1 binds Mg^{2+} with high affinity. *Biochemistry* 50: 8804–8812, 2011.
59. Briel RC, Lippert TH, Zahradnik HP. Action of magnesium sulfate on platelet prostacyclin interaction and prostacyclin of blood vessels. *Am J Obstet Gynecol* 153: 232, 1985.
60. Britton J, Pavord I, Richards K, Wisniewski A, Knox A, Lewis S, Tattersfield A, Weiss S. Dietary magnesium, lung function, wheezing, and airway hyperreactivity in a random adult population sample. *Lancet* 344: 357–362, 1994.
61. Brodowski J. [Levels of ionized magnesium in women with various stages of postmenopausal osteoporosis progression evaluated on the basis of densitometric examinations]. *Przegląd Lekarski* 57: 714–716, 2000.
62. Browne DL, Gancher ST, Nutt JG, Brunt ER, Smith EA, Kramer P, Litt M. Episodic ataxia/myokymia syndrome is associated with point mutations in the human potassium channel gene, KCNA1. *Nature Genet* 8: 136–140, 1994.
63. Brunet S, Scheuer T, Kleivit R, Catterall WA. Modulation of $\text{CaV}1.2$ channels by Mg^{2+} acting at an EF-hand motif in the COOH-terminal domain. *J Gen Physiol* 126: 311–323, 2005.
64. Brunette MG, Vigneault N, Carriere S. Micropuncture study of magnesium transport along the nephron in the young rat. *Am J Physiol* 227: 891–896, 1974.
65. Burnett RJ, Reents SB. Severe hypomagnesemia induced by pentamidine. *Ann Pharmacotherapy* 24: 239–240, 1990.
66. Cairo ER, Friedrich T, Swarts HG, Knoers NV, Bindels RJ, Monnens LA, Willems PH, De Pont JJ, Koenderink JB. Impaired routing of wild type $\text{FX}1\text{D}2$ after oligomerisation with $\text{FX}1\text{D}2$ -G41R might explain the dominant nature of renal hypomagnesemia. *Biochim Biophys Acta* 1778: 398–404, 2008.
67. Cairo ER, Swarts HG, Wilmer MJ, Willems PH, Levchenko EN, De Pont JJ, Koenderink JB. $\text{FX}1\text{D}2$ and Na,K-ATPase expression in isolated human proximal tubular cells: disturbed upregulation on renal hypomagnesemia? *J Membr Biol* 231: 117–124, 2009.
68. Calsou P, Salles B. Properties of damage-dependent DNA incision by nucleotide excision repair in human cell-free extracts. *Nucleic Acids Res* 22: 4937–4942, 1994.
69. Cao G, Hoenderop JG, Bindels RJ. Insight into the molecular regulation of the epithelial magnesium channel TRPM6. *Curr Opin Nephrol Hypertens* 17: 373–378, 2008.
70. Cao G, Thebaud S, van der Wijst J, van der Kemp A, Lasonder E, Bindels RJ, Hoenderop JG. RACK1 inhibits TRPM6 activity via phosphorylation of the fused alpha-kinase domain. *Curr Biol* 18: 168–176, 2008.
71. Cao G, van der Wijst J, van der Kemp A, van Zeeland F, Bindels RJ, Hoenderop JG. Regulation of the epithelial Mg^{2+} channel TRPM6 by estrogen and the associated repressor protein of estrogen receptor activity (REA). *J Biol Chem* 284: 14788–14795, 2009.
72. Cappuccio FP, Markandu ND, Beynon GW, Shore AC, Sampson B, MacGregor GA. Lack of effect of oral magnesium on high blood pressure: a double blind study. *Br Med J* 291: 235–238, 1985.
73. Caspi R, Altman T, Dreher K, Fulcher CA, Subhraveti P, Keseler IM, Kothari A, Krummenacker M, Latendresse M, Mueller LA, Ong Q, Paley S, Pujar A, Shearer AG, Travers M, Weerasinghe D, Zhang P, Karp PD. The MetaCyc database of metabolic pathways and enzymes and the BioCyc collection of pathway/genome databases. *Nucleic Acids Res* 40: D742–753, 2012.
74. Castiglioni S, Maier JA. Magnesium and cancer: a dangerous liaison. *Magnesium Res* 24: S92–100, 2011.
75. Cefaratti C, Young A, Romani A. Effect of ethanol administration on Mg^{2+} transport across liver plasma membrane. *Alcohol* 36: 5–18, 2005.
76. Ceremuzynski L, Jurgiel R, Kulakowski P, Gebalska J. Threatening arrhythmias in acute myocardial infarction are prevented by intravenous magnesium sulfate. *Am Heart J* 118: 1333–1334, 1989.
77. Cernak I, Savic VJ, Kotur J, Prokic V, Veljovic M, Grbovic D. Characterization of plasma magnesium concentration and oxidative stress following graded traumatic brain injury in humans. *J Neurotrauma* 17: 53–68, 2000.
78. Cete Y, Dora B, Ertan C, Ozdemir C, Oktay C. A randomized prospective placebo-controlled study of intravenous magnesium sulphate vs. metoclopramide in the management of acute migraine attacks in the Emergency Department. *Cephalgia* 25: 199–204, 2005.
79. Chaigne-Delalande B, Li FY, O'Connor GM, Lukacs MJ, Jiang P, Zheng L, Shatzer A, Biancalana M, Pittaluga S, Matthews HF, Jancel TJ, Blessing JJ, Marsh RA, Kuijpers TW, Nichols KE, Lucas CL, Nagpal S, Mehmet H, Su HC, Cohen JL, Uzel G, Lenardo MJ. Mg^{2+} regulates cytotoxic functions of NK and CD8 T cells in chronic EBV infection through NKG2D. *Science* 341: 186–191, 2013.
80. Chandler LJ, Guzman NJ, Summers C, Crews FT. Magnesium and zinc potentiate ethanol inhibition of *N*-methyl-D-aspartate-stimulated nitric oxide synthase in cortical neurons. *J Pharmacol Exp Ther* 271: 67–75, 1994.
81. Chanimov M, Berman S, Gofman V, Weissgarten Y, Averbukh Z, Cohen ML, Vitin A, Bahar M. Total cell associated electrolyte homeostasis in rat spinal cord cells following apparently irreversible injury. *Medical Science Monitor* 12: BR63–67, 2006.
82. Chen YZ, Gao Q, Zhao XZ, Chen YZ, Bennett CL, Xiong XS, Mei CL, Shi YQ, Chen XM. Systematic review of TCF2 anomalies in renal cysts and diabetes syndrome/maturity onset diabetes of the young type 5. *Chinese Med J* 123: 3326–3333, 2010.
83. Cheng PT, Grabber JJ, LeGeros RZ. Effects of magnesium on calcium phosphate formation. *Magnesium* 7: 123–132, 1988.
84. Chernow B, Bamberger S, Stoiko M, Vadnais M, Mills S, Hoellerich V, Warsaw AL. Hypomagnesemia in patients in postoperative intensive care. *Chest* 95: 391–397, 1989.
85. Chiu TK, Dickerson RE. I A crystal structures of B-DNA reveal sequence-specific binding and groove-specific bending of DNA by magnesium and calcium. *J Mol Biol* 301: 915–945, 2000.
86. Choi H, Parmar N. The use of intravenous magnesium sulphate for acute migraine: meta-analysis of randomized controlled trials. *Eur J Emergency Medicine* 21: 2–9, 2014.
87. Clarke RJ, Johnson JW. NMDA receptor NR2 subunit dependence of the slow component of magnesium unblock. *J Neurosci* 26: 5825–5834, 2006.
88. Cochrane DE, Douglas WW. Histamine release by exocytosis from rat mast cells on reduction of extracellular sodium: a secretory response inhibited by calcium, strontium, barium or magnesium. *J Physiol* 257: 433–448, 1976.
89. Cohen L, Kitzes R. Infrared spectroscopy and magnesium content of bone mineral in osteoporotic women. *Israel J Med Sci* 17: 1123–1125, 1981.
90. Cohen L, Laor A. Correlation between bone magnesium concentration and magnesium retention in the intravenous magnesium load test. *Magnesium Res* 3: 271–274, 1990.
91. Cohen L, Laor A, Kitzes R. Bone magnesium, crystallinity index and state of body magnesium in subjects with senile osteoporosis, maturity-onset diabetes and women treated with contraceptive preparations. *Magnesium* 2: 70–75, 1983.
92. Cohen N, Almozino-Sarafian D, Zaidenstein R, Alon I, Gorelik O, Shteinshneider M, Chachashvily S, Averbukh Z, Golik A, Chen-Levy Z, Modai D. Serum magnesium aberrations in furosemide (frusemide) treated patients with congestive heart failure: pathophysiological correlates and prognostic evaluation. *Heart* 89: 411–416, 2003.
93. Cook RC, Yamashita MH, Kearns M, Ramanathan K, Gin K, Humphries KH. Prophylactic magnesium does not prevent atrial fibrillation after cardiac surgery: a meta-analysis. *Ann Thoracic Surg* 95: 533–541, 2013.
94. Corbo J, Esses D, Bijur PE, Iannaccone R, Gallagher EJ. Randomized clinical trial of intravenous magnesium sulfate as an adjunctive medication for emergency department treatment of migraine headache. *Ann Emergency Med* 38: 621–627, 2001.
95. Cowan JA. *The Biological Chemistry of Magnesium*. New York: VCH Publishers, 1995.
96. Cross JH, Arora R, Heckemann RA, Gunny R, Chong K, Carr L, Baldeweg T, Differ AM, Lench N, Varadkar S, Sirimanna T, Wassmer E, Hulton SA, Ognjanovic M, Ramesh V, Feather S, Kleta R, Hammers A, Bockenbauer D. Neurological features of epilepsy, ataxia, sensorineural deafness, tubulopathy syndrome. *Dev Med Child Neurol* 55: 846–856, 2013.

97. Cull-Candy S, Brickley S, Farrant M. NMDA receptor subunits: diversity, development and disease. *Curr Opin Neurobiol* 11: 327–335, 2001.
98. Curiel-Garcia JA, Rodriguez-Moran M, Guerrero-Romero F. Hypomagnesemia and mortality in patients with type 2 diabetes. *Magnesium Res* 21: 163–166, 2008.
99. Curry DL, Joy RM, Holley DC, Bennett LL. Magnesium modulation of glucose-induced insulin secretion by the perfused rat pancreas. *Endocrinology* 101: 203–208, 1977.
100. Da Silva CA, de Braganca AC, Shimizu MH, Sanches TR, Fortes MA, Giorgi RR, Andrade L, Seguro AC. Rosiglitazone prevents sirolimus-induced hypomagnesemia, hypokalemia, and downregulation of NKCC2 protein expression. *Am J Physiol Renal Physiol* 297: F916–F922, 2009.
101. Davies DL, Fraser R. Do diuretics cause magnesium deficiency? *Br J Clin Pharmacol* 36: 1–10, 1993.
102. Davy H. Electro-chemical researches, on the decomposition of the earths; with observations on the metals obtained from the alkaline earths, and on the amalgam procured from ammonia. *Philos Trans R Soc Lond* 98: 333–370, 1808.
103. De Baaij JH, Blanchard MG, Lavrijsen M, Leipziger J, Bindels RJ, Hoenderop JG. P2X4 receptor regulation of transient receptor potential melastatin type 6 (TRPM6) Mg channels. *Pflügers Arch* 466: 1941–1952, 2014.
104. De Baaij JH, Groot Koerkamp MJ, Lavrijsen M, van Zeeland F, Meijer H, Holstege FC, Bindels RJ, Hoenderop JG. Elucidation of the distal convoluted tubule transcriptome identifies new candidate genes involved in renal magnesium handling. *Am J Physiol Renal Physiol* 305: F1563–F1573, 2013.
105. De Baaij JH, Stuiver M, Meij IC, Lainez S, Kopplin K, Venselaar H, Muller D, Bindels RJ, Hoenderop JG. Membrane topology and intracellular processing of cyclin M2 (CNNM2). *J Biol Chem* 287: 13644–13655, 2012.
106. De Francisco AL, Leidig M, Covic AC, Ketteler M, Benedyk-Lorens E, Mircescu GM, Scholz C, Ponce P, Passlick-Deetjen J. Evaluation of calcium acetate/magnesium carbonate as a phosphate binder compared with sevelamer hydrochloride in haemodialysis patients: a controlled randomized study (CALMAG study) assessing efficacy and tolerability. *Nephrol Dialysis Transplant* 25: 3707–3717, 2010.
107. De Lorges Lima M, Cruz T, Pousada JC, Rodrigues LE, Barbosa K, Cangucu V. The effect of magnesium supplementation in increasing doses on the control of type 2 diabetes. *Diabetes Care* 21: 682–686, 1998.
108. De Oliveira GS Jr, Knautz JS, Sherwani S, McCarthy RJ. Systemic magnesium to reduce postoperative arrhythmias after coronary artery bypass graft surgery: a meta-analysis of randomized controlled trials. *J Cardiothoracic Vasc Anesthesia* 26: 643–650, 2012.
109. De Valk HW, Kok PT, Struyvenberg A, van Rijn HJ, Haalboom JR, Kreukniet J, Lammers JW. Extracellular and intracellular magnesium concentrations in asthmatic patients. *Eur Respir J* 6: 1122–1125, 1993.
110. Deason-Towne F, Perraud AL, Schmitz C. The Mg^{2+} transporter MagT1 partially rescues cell growth and Mg^{2+} uptake in cells lacking the channel-kinase TRPM7. *FEBS Lett* 585: 2275–2278, 2011.
111. DeGraba TJ, Pettigrew LC. Why do neuroprotective drugs work in animals but not humans? *Neurol Clinics* 18: 475–493, 2000.
112. Demirkaya S, Vural O, Dora B, Topcuoglu MA. Efficacy of intravenous magnesium sulfate in the treatment of acute migraine attacks. *Headache* 41: 171–177, 2001.
113. Denis W. Determination of magnesium in blood. *J Biol Chem* 41: 363–365, 1920.
114. Derom ML, Sayon-Orea C, Martinez-Ortega JM, Martinez-Gonzalez MA. Magnesium and depression: a systematic review. *Nutr Neurosci* 16: 191–206, 2013.
115. Desai BN, Krapivinsky G, Navarro B, Krapivinsky L, Carter BC, Febvay S, Delling M, Penumaka A, Ramsey IS, Manasian Y, Clapham DE. Cleavage of TRPM7 releases the kinase domain from the ion channel and regulates its participation in Fas-induced apoptosis. *Dev Cell* 22: 1149–1162, 2012.
116. Dhandapani SS, Gupta A, Vivekanandhan S, Sharma BS, Mahapatra AK. Randomized controlled trial of magnesium sulphate in severe closed traumatic brain injury. *Indian J Neurotrauma* 5: 27–33, 2008.
117. Dias JM, Szegedi C, Jona I, Vogel PD. Insights into the regulation of the ryanodine receptor: differential effects of Mg^{2+} and Ca^{2+} on ATP binding. *Biochemistry* 45: 9408–9415, 2006.
118. Dickens BF, Weglicki WB, Li YS, Mak IT. Magnesium deficiency in vitro enhances free radical-induced intracellular oxidation and cytotoxicity in endothelial cells. *FEBS Lett* 311: 187–191, 1992.
119. Dickinson HO, Nicolson DJ, Campbell F, Cook JV, Beyer FR, Ford GA, Mason J. Magnesium supplementation for the management of essential hypertension in adults. *Cochrane Database of Systematic Rev* CD004640, 2006.
120. Diederichs F. Ion homeostasis and the functional roles of SERCA reactions in stimulus-secretion coupling of the pancreatic beta-cell: a mathematical simulation. *Biophys Chem* 134: 119–143, 2008.
121. Dimke H, Desai P, Borovac J, Lau A, Pan W, Alexander RT. Activation of the Ca^{2+} -sensing receptor increases renal claudin-14 expression and urinary Ca^{2+} excretion. *Am J Physiol Renal Physiol* 304: F761–F769, 2013.
122. Dimke H, van der Wijst J, Alexander TR, Meijer IM, Mulder GM, van Goor H, Tejpar S, Hoenderop JG, Bindels RJ. Effects of the EGFR Inhibitor Erlotinib on Magnesium Handling. *J Am Soc Nephrol* 21: 1309–1316, 2010.
123. Do Amaral AF, Rodrigues-Junior AL, Terra Filho J, Vannucchi H, Martinez JA. Effects of acute magnesium loading on pulmonary function of stable COPD patients. *Med Sci Monitor* 14: CR524–529, 2008.
124. Dorhout Mees SM, Algra A, Vandertop WP, van Kooten F, Kuijsten HA, Boiten J, van Oostenbrugge RJ, Al-Shahi Salman R, Lavados PM, Rinkel GJ, van den Bergh WM. Magnesium for aneurysmal subarachnoid haemorrhage (MASH-2): a randomised placebo-controlled trial. *Lancet* 380: 44–49, 2012.
125. Dorhout Mees SM, Rinkel GJ, Feigin VL, Algra A, van den Bergh WM, Vermeulen M, van Gijn J. Calcium antagonists for aneurysmal subarachnoid haemorrhage. *Cochrane Database Systematic Rev* CD000277, 2007.
126. Dorsch NW. Cerebral arterial spasm—a clinical review. *Br J Neurosurg* 9: 403–412, 1995.
127. Duarte CG. Effects of ethacrynic acid and furosemide on urinary calcium, phosphate and magnesium. *Metab Clin Exp* 17: 867–876, 1968.
128. Duchatelle-Gourdon I, Hartzell HC, Lagrutta AA. Modulation of the delayed rectifier potassium current in frog cardiomyocytes by beta-adrenergic agonists and magnesium. *J Physiol* 415: 251–274, 1989.
129. Ducker CE, Stettler EM, French KJ, Upson JJ, Smith CD. Huntingtin interacting protein 14 is an oncogenic human protein: palmitoyl acyltransferase. *Oncogene* 23: 9230–9237, 2004.
130. Duley L, Gulmezoglu AM, Henderson-Smart DJ, Chou D. Magnesium sulphate and other anticonvulsants for women with pre-eclampsia. *Cochrane Database Systematic Rev* CD000025, 2010.
131. Duley L, Henderson-Smart DJ, Chou D. Magnesium sulphate versus phenytoin for eclampsia. *Cochrane Database Systematic Rev* CD000128, 2010.
132. Duley L, Henderson-Smart DJ, Walker GJ, Chou D. Magnesium sulphate versus diazepam for eclampsia. *Cochrane Database Systematic Rev* CD000127, 2010.
133. Ebel H, Gunther T. Magnesium metabolism: a review. *J Clin Chem Clin Biochem* 18: 257–270, 1980.
134. Ebel H, Hollstein M, Gunther T. Role of the choline exchanger in Na^{+} -independent Mg^{2+} efflux from rat erythrocytes. *Biochim Biophys Acta* 1559: 135–144, 2002.
135. Eby GA, Eby KL. Rapid recovery from major depression using magnesium treatment. *Medical Hypotheses* 67: 362–370, 2006.
136. Edwards L, Shirtcliffe P, Wadsworth K, Healy B, Jefferies S, Weatherall M, Beasley R, Team obotMCS. Use of nebulised magnesium sulphate as an adjuvant in the treatment of acute exacerbations of COPD in adults: a randomised double-blind placebo-controlled trial. *Thorax* 68: 338–343, 2013.
137. Eimerl S, Schramm M. The quantity of calcium that appears to induce neuronal death. *J Neurochem* 62: 1223–1226, 1994.
138. Eisinger J, Clairet D. Effects of silicon, fluoride, etidronate and magnesium on bone mineral density: a retrospective study. *Magnesium Res* 6: 247–249, 1993.
139. Elin RJ. Assessment of magnesium status for diagnosis and therapy. *Magnesium Res* 23: S194–198, 2010.

140. Ellison DH. The voltage-gated K⁺ channel subunit Kv1.1 links kidney and brain. *J Clin Invest* 119: 763–766, 2009.
141. Emelyanov A, Fedoseev G, Barnes PJ. Reduced intracellular magnesium concentrations in asthmatic patients. *Eur Respir J* 13: 38–40, 1999.
142. English MW, Skinner R, Pearson AD, Price L, Wyllie R, Craft AW. Dose-related nephrotoxicity of carboplatin in children. *Br J Cancer* 81: 336–341, 1999.
143. Enya M, Kanoh Y, Mune T, Ishizawa M, Sarui H, Yamamoto M, Takeda N, Yasuda K, Yasujima M, Tsutaya S, Takeda J. Depressive state and paresthesia dramatically improved by intravenous MgSO₄ in Gitelman's syndrome. *Intern Med* 43: 410–414, 2004.
144. Epstein M, McGrath S, Law F. Proton-pump inhibitors and hypomagnesemic hypoparathyroidism. *N Engl J Med* 355: 1834–1836, 2006.
145. Escuela MP, Guerra M, Anon JM, Martinez-Vizcaino V, Zapatero MD, Garcia-Jalon A, Celaya S. Total and ionized serum magnesium in critically ill patients. *Intensive Care Med* 31: 151–156, 2005.
146. Eunson LH, Rea R, Zuberi SM, Youroukos S, Panayiotopoulos CP, Liguori R, Avoni P, McWilliam RC, Stephenson JB, Hanna MG, Kullmann DM, Spauschus A. Clinical, genetic, and expression studies of mutations in the potassium channel gene KCNA1 reveal new phenotypic variability. *Ann Neurol* 48: 647–656, 2000.
147. Euser AG, Cipolla MJ. Magnesium sulfate for the treatment of eclampsia: a brief review. *Stroke* 40: 1169–1175, 2009.
148. Evers S, Afra J, Frese A, Goadsby PJ, Linde M, May A, Sandor PS. EFNS guideline on the drug treatment of migraine—revised report of an EFNS task force. *Eur J Neurol* 16: 968–981, 2009.
149. Facchinetti F, Sances G, Borella P, Genazzani AR, Nappi G. Magnesium prophylaxis of menstrual migraine: effects on intracellular magnesium. *Headache* 31: 298–301, 1991.
150. Farouque HM, Sanders P, Young GD. Intravenous magnesium sulfate for acute termination of sustained monomorphic ventricular tachycardia associated with coronary artery disease. *Am J Cardiol* 86: 1270–1272, 2000.
151. Fei X, Hongxiang Z, Qi C, Daozhen C. Maternal plasma levels of endothelial dysfunction mediators including AM, CGRP, sICAM-1 and tHcy in pre-eclampsia. *Adv Clin Exp Med* 21: 573–579, 2012.
152. Ferrara LA, Iannuzzi R, Castaldo A, Iannuzzi A, Dello Russo A, Mancini M. Long-term magnesium supplementation in essential hypertension. *Cardiology* 81: 25–33, 1992.
153. Ferre S, Baldoli E, Leidi M, Maier JA. Magnesium deficiency promotes a pro-atherogenic phenotype in cultured human endothelial cells via activation of NFκB. *Biochim Biophys Acta* 1802: 952–958, 2010.
154. Ferre S, de Baaij JH, Ferreira P, Germann R, de Klerk JB, Lavrijsen M, van Zeeland F, Venselaar H, Kluijtmans LA, Hoenderop JG, Bindels RJ. Mutations in PCBD1 Cause Hypomagnesemia and Renal Magnesium Wasting. *J Am Soc Nephrol* 25: 574–586, 2014.
155. Ferre S, Veenstra GJ, Bouwmeester R, Hoenderop JG, Bindels RJ. HNF-1B specifically regulates the transcription of the gamma-a subunit of the Na⁺/K⁺-ATPase. *Biochem Biophys Res Commun* 404: 284–290, 2011.
156. Feske S, Skolnik EY, Prakriya M. Ion channels and transporters in lymphocyte function and immunity. *Nature Rev Immunol* 12: 532–547, 2012.
157. Finton CK, Bjorkland S, Zaloga GP, Uddin DE, Chernow B. Gentamicin-induced hypomagnesemia. *Ann Surgeon* 49: 576–578, 1983.
158. Fiorica V. *Contribution of Activity to the Circadian Rhythm in Excretion of Magnesium and Calcium*. Washington, DC: Federal Aviation Administration, Department of Transportation, Office of Aviation Medicine, 1968.
159. Fisher RS, Kaplan PW, Krumholz A, Lesser RP, Rosen SA, Wolff MR. Failure of high-dose intravenous magnesium sulfate to control myoclonic status epilepticus. *Clin Neuropharmacol* 11: 537–544, 1988.
160. Folsom AR, Hong CP. Magnesium intake and reduced risk of colon cancer in a prospective study of women. *Am J Epidemiol* 163: 232–235, 2006.
161. Fonseca FA, Paiva TB, Silva EG, Ihara SS, Kasinski N, Martinez TL, Filho EE. Dietary magnesium improves endothelial dependent relaxation of balloon injured arteries in rats. *Atherosclerosis* 139: 237–242, 1998.
162. Ford ES. Serum magnesium and ischaemic heart disease: findings from a national sample of US adults. *Int J Epidemiol* 28: 645–651, 1999.
163. Ford ES, Mokdad AH. Dietary magnesium intake in a national sample of US adults. *J Nutr* 133: 2879–2882, 2003.
164. Fordtran JS, Rector FC Jr, Carter NW. The mechanisms of sodium absorption in the human small intestine. *J Clin Invest* 47: 884–900, 1968.
165. Gamba G, Saltzberg SN, Lombardi M, Miyashita A, Lytton J, Hediger MA, Brenner BM, Hebert SC. Primary structure and functional expression of a cDNA encoding the thiazide-sensitive, electroneutral sodium-chloride cotransporter. *Proc Natl Acad Sci USA* 90: 2749–2753, 1993.
166. Garfinkel L, Garfinkel D. Magnesium regulation of the glycolytic pathway and the enzymes involved. *Magnesium* 4: 60–72, 1985.
167. Garland HO, Birdsey TJ, Davidge CG, McLaughlin JT, Oakes LM, Smith AJ, Harpur ES. Effects of gentamicin, neomycin and tobramycin on renal calcium and magnesium handling in two rat strains. *Clin Exp Pharmacol Physiol* 21: 109–115, 1994.
168. Garrison SR, Allan GM, Sekhon RK, Musini VM, Khan KM. Magnesium for skeletal muscle cramps. *Cochrane Database Systematic Rev* 9: CD009402, 2012.
169. Gearhart MO, Sorg TB. Foscarnet-induced severe hypomagnesemia and other electrolyte disorders. *Ann Pharmacother* 27: 285–289, 1993.
170. Gelfand EW. Role of histamine in the pathophysiology of asthma: immunomodulatory and anti-inflammatory activities of H1-receptor antagonists. *Am J Med* 113 Suppl 9A: 2S–7S, 2002.
171. George MS, Rosenstein D, Rubinow DR, Kling MA, Post RM. CSF magnesium in affective disorder: lack of correlation with clinical course of treatment. *Psychiatry Res* 51: 139–146, 1994.
172. Geven WB, Monnens LA, Willems HL, Buijs WC, ter Haar BG. Renal magnesium wasting in two families with autosomal dominant inheritance. *Kidney Int* 31: 1140–1144, 1987.
173. Geven WB, Monnens LA, Willems JL, Buijs W, Hamel CJ. Isolated autosomal recessive renal magnesium loss in two sisters. *Clin Genet* 32: 398–402, 1987.
174. Gilliland FD, Berhane KT, Li YF, Kim DH, Margolis HG. Dietary magnesium, potassium, sodium, and children's lung function. *American journal of epidemiol* 155: 125–131, 2002.
175. Gitelman HJ, Graham JB, Welt LG. A new familial disorder characterized by hypokalemia and hypomagnesemia. *Trans Assoc Am Physicians* 79: 221–235, 1966.
176. Glaudemans B, van der Wijst J, Scola RH, Lorenzoni PJ, Heister A, van der Kemp AW, Knoers NV, Hoenderop JG, Bindels RJ. A missense mutation in the Kv1.1 voltage-gated potassium channel-encoding gene KCNA1 is linked to human autosomal dominant hypomagnesemia. *J Clin Invest* 119: 936–942, 2009.
177. Godron A, Harambat J, Boccio V, Mensire A, May A, Rigotherier C, Couzi L, Barrou B, Godin M, Chauveau D, Faguer S, Vallet M, Cochat P, Eckart P, Guest G, Guignon V, Houillier P, Blanchard A, Jeunemaitre X, Vargas-Poussou R. Familial hypomagnesemia with hypercalciuria and nephrocalcinosis: phenotype-genotype correlation and outcome in 32 patients with CLDN16 or CLDN19 mutations. *Clin J Am Soc Nephrol* 7: 801–809, 2012.
178. Goldman RD, Koren G. Amphotericin B nephrotoxicity in children. *J Pediatr Hematol Oncol* 26: 421–426, 2004.
179. Gomez-Hernandez JM, Lorra C, Pardo LA, Stuhmer W, Pongs O, Heinemann SH, Elliott AA. Molecular basis for different pore properties of potassium channels from the rat brain Kv1 gene family. *Pflügers Arch* 434: 661–668, 1997.
180. Gong Y, Renigunta V, Himmerkus N, Zhang J, Renigunta A, Bleich M, Hou J. Claudin-14 regulates renal Ca²⁺ transport in response to CaSR signalling via a novel microRNA pathway. *EMBO J* 31: 1999–2012, 2012.
181. Gorji A, Scheller D, Straub H, Tegtmeyer F, Köhling R, Höhling JM, Tuxhorn I, Ebner A, Wolf P, Werner Panneck H, Oppel F, Speckmann EJ. Spreading depression in human neocortical slices. *Brain Res* 906: 74–83, 2001.
182. Goytain A, Hines RM, El-Husseini A, Quamme GA. NIPA1 (SPG6), the basis for autosomal dominant form of hereditary spastic paraplegia, encodes a functional Mg²⁺ transporter. *J Biol Chem* 282: 8060–8068, 2007.

183. Goytain A, Hines RM, Quamme GA. Huntingtin-interacting proteins, HIP14 and HIP14L, mediate dual functions, palmitoyl acyltransferase and Mg^{2+} transport. *J Biol Chem* 283: 33365–33374, 2008.
184. Goytain A, Quamme GA. Functional characterization of ACDP2 (ancient conserved domain protein), a divalent metal transporter. *Physiol Genomics* 22: 382–389, 2005.
185. Goytain A, Quamme GA. Functional characterization of human SLC41A1, a Mg^{2+} transporter with similarity to prokaryotic MgtE Mg^{2+} transporters. *Physiol Genomics* 21: 337–342, 2005.
186. Goytain A, Quamme GA. Identification and characterization of a novel family of membrane magnesium transporters, MMgT1 and MMgT2. *Am J Physiol Cell Physiol* 294: C495–C502, 2008.
187. Goytain A, Quamme GA. Identification and characterization of a novel mammalian Mg^{2+} transporter with channel-like properties. *BMC Genomics* 6: 48, 2005.
188. Gradon JD, Fricchione L, Sepkowitz D. Severe hypomagnesemia associated with pentamidine therapy. *Rev Infect Dis* 13: 511–512, 1991.
189. Graham LA, Caesar JJ, Burgen AS. Gastrointestinal absorption and excretion of Mg 28 in man. *Metabolism Clin Exp* 9: 646–659, 1960.
190. Grant AO. Cardiac ion channels. *Circ Arrhythmia Electrophysiol* 2: 185–194, 2009.
191. Green L, Kim CH, Bustamante C, Tinoco J Jr. Characterization of the mechanical unfolding of RNA pseudoknots. *J Mol Biol* 375: 511–528, 2008.
192. Greger R, Velazquez H. The cortical thick ascending limb and early distal convoluted tubule in the urinary concentrating mechanism. *Kidney Int* 31: 590–596, 1987.
193. Greising SM, Gransee HM, Mantilla CB, Sieck GC. Systems biology of skeletal muscle: fiber type as an organizing principle. *Wiley Interdisciplinary Rev Systems Biol Med* 4: 457–473, 2012.
194. Gresh L, Fischer E, Reimann A, Tanguy M, Garbay S, Shao X, Hiesberger T, Fiette L, Igarashi P, Yaniv M, Pontoglio M. A transcriptional network in polycystic kidney disease. *EMBO J* 23: 1657–1668, 2004.
195. Grew N. *A Treatise of the Nature and Use of the Bitter Purging Salt Contain'd in Epsom, and Such Other Waters*. London: [s.n.], 1697.
196. Groenestege WM, Hoenderop JG, van den Heuvel L, Knoers N, Bindels RJ. The epithelial Mg^{2+} channel transient receptor potential melastatin 6 is regulated by dietary Mg^{2+} content and estrogens. *J Am Soc Nephrol* 17: 1035–1043, 2006.
197. Groenestege WM, Thebault S, van der Wijst J, van den Berg D, Janssen R, Tejpar S, van den Heuvel LP, van Cutsem E, Hoenderop JG, Knoers NV, Bindels RJ. Impaired basolateral sorting of pro-EGF causes isolated recessive renal hypomagnesemia. *J Clin Invest* 117: 2260–2267, 2007.
198. Group TETC. Which anticonvulsant for women with eclampsia? Evidence from the Collaborative Eclampsia Trial. *Lancet* 345: 1455–1463, 1995.
199. Grubbs RD. Effect of epidermal growth factor on magnesium homeostasis in BC3H1 myocytes. *Am J Physiol Cell Physiol* 260: C1158–C1164, 1991.
200. Gruber H, Ingram J, Norton H, Wei L, Frausto A, Mills B, Rude R. Alterations in growth plate and articular cartilage morphology are associated with reduced SOX9 localization in the magnesium-deficient rat. *Biotechnol Histochem* 79: 45–52, 2004.
201. Gu WJ, Wu ZJ, Wang PF, Aung LH, Yin RX. Intravenous magnesium prevents atrial fibrillation after coronary artery bypass grafting: a meta-analysis of 7 double-blind, placebo-controlled, randomized clinical trials. *Trials* 13: 41, 2012.
202. Gueux E, Rayssiguier Y. The effect of magnesium deficiency on glucose stimulated insulin secretion in rats. *Horm Metab Res* 15: 594–597, 1983.
203. Guilbert A, Gautier M, Dhennin-Duthille I, Haren N, Sevestre H, Ouadid-Ahidouch H. Evidence that TRPM7 is required for breast cancer cell proliferation. *Am J Physiol Cell Physiol* 297: C493–C502, 2009.
204. Gullestad L, Jacobsen T, Dolva LO. Effect of magnesium treatment on glycemic control and metabolic parameters in NIDDM patients. *Diabetes Care* 17: 460–461, 1994.
205. Gullner HG, Gill JR Jr, Bartter FC. Correction of hypokalemia by magnesium repletion in familial hypokalemic alkalosis with tubulopathy. *Am J Med* 71: 578–582, 1981.
206. Gunther T. The biochemical function of Mg^{2+} in insulin secretion, insulin signal transduction and insulin resistance. *Magnesium Res* 23: 5–18, 2010.
207. Gunzel D, Amasheh S, Pfaffenbach S, Richter JF, Kausalya PJ, Hunziker W, Fromm M. Claudin-16 affects transcellular Cl^{-} secretion in MDCK cells. *J Physiol* 587: 3777–3793, 2009.
208. Gunzel D, Stuver M, Kausalya PJ, Haisch L, Krug SM, Rosenthal R, Meij IC, Hunziker W, Fromm M, Muller D. Claudin-10 exists in six alternatively spliced isoforms that exhibit distinct localization and function. *J Cell Sci* 122: 1507–1517, 2009.
209. Gunzel D, Yu AS. Claudins and the modulation of tight junction permeability. *Physiol Rev* 93: 525–569, 2013.
210. Gupta A, Eastham KM, Wrightson N, Spencer DA. Hypomagnesaemia in cystic fibrosis patients referred for lung transplant assessment. *J Cystic Fibrosis* 6: 360–362, 2007.
211. Gupta A, Lawrence AT, Krishnan K, Kavinsky CJ, Trohman RG. Current concepts in the mechanisms and management of drug-induced QT prolongation and torsade de pointes. *Am Heart J* 153: 891–899, 2007.
212. Gupta SK, Manhas AS, Gupta VK, Bhatt R. Serum magnesium levels in idiopathic epilepsy. *The J Assoc Physicians India* 42: 456–457, 1994.
213. Gur A, Colpan L, Nas K, Cevik R, Sarac J, Erdogan F, Duz MZ. The role of trace minerals in the pathogenesis of postmenopausal osteoporosis and a new effect of calcitonin. *J Bone Miner Metab* 20: 39–43, 2002.
214. Guther T, Vormann J, Forster R. Regulation of intracellular magnesium by Mg^{2+} efflux. *Biochem Biophys Res Commun* 119: 124–131, 1984.
215. Hall RC, Joffe JR. Hypomagnesemia. Physical and psychiatric symptoms. *JAMA* 224: 1749–1751, 1973.
216. Hardwick LL, Jones MR, Buddington RK, Clemens RA, Lee DB. Comparison of calcium and magnesium absorption: in vivo and in vitro studies. *Am J Physiol Gastrointest Liver Physiol* 259: G720–G726, 1990.
217. Hardy S, Uetani N, Wong N, Kostantin E, Labbe DP, Begin LR, Mes-Masson A, Miranda-Saavedra D, Tremblay ML. The protein tyrosine phosphatase PRL-2 interacts with the magnesium transporter CNNM3 to promote oncogenesis. *Oncogene* 2014; doi:10.1038/onc.2014.33.
218. Hashimoto T, Hara A, Ohkubo T, Kikuya M, Shintani Y, Metoki H, Inoue R, Asayama K, Kanno A, Nakashita M, Terata S, Obara T, Hirose T, Hoshi H, Totsune K, Satoh H, Imai Y. Serum magnesium, ambulatory blood pressure, and carotid artery alteration: the Ohasama study. *Am J Hypertens* 23: 1292–1298, 2010.
219. Hashimoto Y, Nishimura Y, Maeda H, Yokoyama M. Assessment of magnesium status in patients with bronchial asthma. *J Asthma* 37: 489–496, 2000.
220. Hassan TB, Jagger C, Barnett DB. A randomised trial to investigate the efficacy of magnesium sulphate for refractory ventricular fibrillation. *Emergency Med J* 19: 57–62, 2002.
221. Haury VG. Blood serum magnesium in bronchial asthma and its treatment by the administration of magnesium sulfate. *J Lab Clin Med* 26: 340–344, 1940.
222. He K, Liu K, Daviglus ML, Morris SJ, Loria CM, Van Horn L, Jacobs DR Jr, Savage PJ. Magnesium intake and incidence of metabolic syndrome among young adults. *Circulation* 113: 1675–1682, 2006.
223. World Health Organization, Department of Reproductive Health and Research. *Managing Complications in Pregnancy and Childbirth: A Guide for Midwives and Doctors*. World Health Organization, 2003.
224. Heaton F, Pyrah L, Beresford C, Bryson R, Martin D. Hypomagnesaemia in chronic alcoholism. *Lancet* 280: 802–805, 1962.
225. Hershko A, Mamont P, Shields R, Tomkins GM. "Pleiotypic response." *Nature New Biol* 232: 206–211, 1971.
226. Hess MW, Hoenderop JG, Bindels RJ, Drenth JP. Systematic review: hypomagnesaemia induced by proton pump inhibition. *Alimentary Pharmacol Ther* 36: 405–413, 2012.
227. Himmerkus N, Shan Q, Goerke B, Hou J, Goodenough DA, Bleich M. Salt and acid-base metabolism in claudin-16 knockdown mice: impact for the pathophysiology of FHHNC patients. *Am J Physiol Renal Physiol* 295: F1641–F1647, 2008.

228. Hirose M, Kobayashi M, Sudo S, Nakanishi K, Noda Y. Hemodialysis for toxic hypermagnesemia caused by intravenous magnesium in a woman with eclampsia and renal insufficiency. A case report. *J Reprod Med* 47: 1050–1052, 2002.
229. Hirota K, Sato T, Hashimoto Y, Yoshioka H, Ohtomo N, Ishihara H, Matsuki A. Relaxant effect of magnesium and zinc on histamine-induced bronchoconstriction in dogs. *Crit Care Med* 27: 1159–1163, 1999.
231. Hirschfelder AD, Haury VG. Clinical manifestations of high and low plasma magnesium: dangers of epsom salt purgation in nephritis. *JAMA* 102: 1138–1141, 1934.
232. Ho KM. Intravenous magnesium for cardiac arrhythmias: jack of all trades. *Magnesium Res* 21: 65–68, 2008.
233. Hoane MR. Assessment of cognitive function following magnesium therapy in the traumatically injured brain. *Magnesium Res* 20: 229–236, 2007.
234. Hoenderop JG, Bindels RJ. Calcitropic and magnesiotropic TRP channels. *Physiology* 23: 32–40, 2008.
235. Hoenderop JG, Bindels RJ. Epithelial Ca^{2+} and Mg^{2+} channels in health and disease. *J Am Soc Nephrol* 16: 15–26, 2005.
236. Hoenderop JG, Nilius B, Bindels RJ. ECaC: the gatekeeper of transepithelial Ca^{2+} transport. *Biochim Biophys Acta* 1600: 6–11, 2002.
237. Holland S, Silberstein SD, Freitag F, Dodick DW, Argoff C, Ashman E. Evidence-based guideline update: NSAIDs and other complementary treatments for episodic migraine prevention in adults: report of the Quality Standards Subcommittee of the American Academy of Neurology and the American Headache Society. *Neurology* 78: 1346–1353, 2012.
238. Hollifield JW. Thiazide treatment of hypertension. Effects of thiazide diuretics on serum potassium, magnesium, and ventricular ectopy. *Am J Med* 80: 8–12, 1986.
239. Holroyde MJ, Potter JD, Solaro RJ. The calcium binding properties of phosphorylated and unphosphorylated cardiac and skeletal myosins. *J Biol Chem* 254: 6478–6482, 1979.
240. Hong BZ, Kang HS, So JN, Kim HN, Park SA, Kim SJ, Kim KR, Kwak YG. Vascular endothelial growth factor increases the intracellular magnesium. *Biochem Biophys Res Commun* 347: 496–501, 2006.
241. Hong BZ, Park SA, Kim HN, Ma TZ, Kim HG, Kang HS, Kim HG, Kwak YG. Basic fibroblast growth factor increases intracellular magnesium concentration through the specific signaling pathways. *Molecules Cells* 28: 13–17, 2009.
242. Hoorn EJ, Walsh SB, McCormick JA, Furstenberg A, Yang CL, Roeschel T, Paliege A, Howie AJ, Conley J, Bachmann S, Unwin RJ, Ellison DH. The calcineurin inhibitor tacrolimus activates the renal sodium chloride cotransporter to cause hypertension. *Nature Med* 17: 1304–1309, 2011.
243. Hoshino K, Ogawa K, Hishitani T, Isobe T, Eto Y. Optimal administration dosage of magnesium sulfate for torsades de pointes in children with long QT syndrome. *J Am Coll Nutr* 23: 497S–500S, 2004.
244. Hoshino K, Ogawa K, Hishitani T, Isobe T, Etoh Y. Successful uses of magnesium sulfate for torsades de pointes in children with long QT syndrome. *Pediatrics Int* 48: 112–117, 2006.
245. Hou J, Paul DL, Goodenough DA. Paracellin-1 and the modulation of ion selectivity of tight junctions. *J Cell Sci* 118: 5109–5118, 2005.
246. Hou J, Renigunta A, Gomes AS, Hou M, Paul DL, Waldegger S, Goodenough DA. Claudin-16 and claudin-19 interaction is required for their assembly into tight junctions and for renal reabsorption of magnesium. *Proc Natl Acad Sci USA* 106: 15350–15355, 2009.
247. Hou J, Renigunta A, Konrad M, Gomes AS, Schneeberger EE, Paul DL, Waldegger S, Goodenough DA. Claudin-16 and claudin-19 interact and form a cation-selective tight junction complex. *J Clin Invest* 118: 619–628, 2008.
248. Huang CL, Kuo E. Mechanism of hypokalemia in magnesium deficiency. *J Am Soc Nephrol* 18: 2649–2652, 2007.
249. Hubbard SR. Crystal structure of the activated insulin receptor tyrosine kinase in complex with peptide substrate and ATP analog. *EMBO J* 16: 5572–5581, 1997.
250. Hubner CA, Holthoff K. Anion transport and GABA signaling. *Front Cell Neurosci* 7: 177, 2013.
251. Hurd TW, Otto EA, Mishima E, Gee HY, Inoue H, Inazu M, Yamada H, Halbritter J, Seki G, Konishi M, Zhou W, Yamane T, Murakami S, Caridi G, Ghiggeri G, Abe T, Hildebrandt F. Mutation of the Mg^{2+} transporter SLC41A1 results in a nephronophthisis-like phenotype. *J Am Soc Nephrol* 24: 967–977, 2013.
252. Hutchison AJ, Wilkie M. Use of magnesium as a drug in chronic kidney disease. *Clin Kidney J* 5: i62–i70, 2012.
253. Huyck MM, Naguib MT, Stroemmel MM, Blick K, Monti K, Martin-Munley S, Kaufman C. A double-blind placebo-controlled crossover trial of intravenous magnesium sulfate for foscarnet-induced ionized hypocalcemia and hypomagnesemia in patients with AIDS and cytomegalovirus infection. *Antimicrobial Agents Chemother* 44: 2143–2148, 2000.
254. Ikari A, Kano T, Suketa Y. Magnesium influx enhanced by nitric oxide in hypertensive rat proximal tubule cells. *Biochem Biophys Res Commun* 294: 710–713, 2002.
255. Ikari A, Sanada A, Sawada H, Okude C, Tonegawa C, Sugatani J. Decrease in transient receptor potential melastatin 6 mRNA stability caused by rapamycin in renal tubular epithelial cells. *Biochim Biophys Acta* 1808: 1502–1508, 2011.
256. Ikonomidou C, Turski L. Why did NMDA receptor antagonists fail clinical trials for stroke and traumatic brain injury? *Lancet Neurol* 1: 383–386, 2002.
257. Iseri LT, French JH. Magnesium: nature's physiologic calcium blocker. *Am Heart J* 108: 188–193, 1984.
258. Ishimura E, Okuno S, Kitatani K, Tsuchida T, Yamakawa T, Shioi A, Inaba M, Nishizawa Y. Significant association between the presence of peripheral vascular calcification and lower serum magnesium in hemodialysis patients. *Clin Nephrol* 68: 222–227, 2007.
259. Ishimura E, Okuno S, Yamakawa T, Inaba M, Nishizawa Y. Serum magnesium concentration is a significant predictor of mortality in maintenance hemodialysis patients. *Magnesium Res* 20: 237–244, 2007.
260. Itoh K, Kawasaka T, Nakamura M. The effects of high oral magnesium supplementation on blood pressure, serum lipids and related variables in apparently healthy Japanese subjects. *Br J Nutr* 78: 737–750, 1997.
261. Jeck N, Konrad M, Peters M, Weber S, Bonzel KE, Seyberth HW. Mutations in the chloride channel gene, CLCNKB, leading to a mixed Bartter-Gitelman phenotype. *Pediatr Res* 48: 754–758, 2000.
262. Jee SH, Miller ER, 3rd Guallar E, Singh VK, Appel LJ, Klag MJ. The effect of magnesium supplementation on blood pressure: a meta-analysis of randomized clinical trials. *Am J Hypertens* 15: 691–696, 2002.
263. Jen JC, Graves TD, Hess EJ, Hanna MG, Griggs RC, Baloh RW. Primary episodic ataxias: diagnosis, pathogenesis and treatment. *Brain* 130: 2484–2493, 2007.
264. Jiang J, Li MH, Inoue K, Chu XP, Seeds J, Xiong ZG. Transient receptor potential melastatin 7-like current in human head and neck carcinoma cells: role in cell proliferation. *Cancer Res* 67: 10929–10938, 2007.
265. Jiang SP, Wu YM, Guo SE, Lv ZQ. Decreased renal mRNA expression of TRPM6 is associated with hypomagnesemia in C57BL/6 asthmatic mice. *Eur Rev Med Pharmacol Sci* 14: 935–940, 2010.
266. Jin J, Desai BN, Navarro B, Donovan A, Andrews NC, Clapham DE. Deletion of *Trpm7* disrupts embryonic development and thymopoiesis without altering Mg^{2+} homeostasis. *Science* 322: 756–760, 2008.
267. Joffres MR, Reed DM, Yano K. Relationship of magnesium intake and other dietary factors to blood pressure: the Honolulu heart study. *Am J Clin Nutr* 45: 469–475, 1987.
268. Kahraman S, Ozgurtas T, Kayali H, Atabay C, Kutluay T, Timurkaynak E. Monitoring of serum ionized magnesium in neurosurgical intensive care unit: preliminary results. *Clin Chim Acta* 334: 211–215, 2003.
269. Kang HS, Kerstan D, Dai L, Ritchie G, Quamme GA. Aminoglycosides inhibit hormone-stimulated Mg^{2+} uptake in mouse distal convoluted tubule cells. *Can J Physiol Pharmacol* 78: 595–602, 2000.
270. Kao WH, Folsom AR, Nieto FJ, Mo JP, Watson RL, Brancati FL. Serum and dietary magnesium and the risk for type 2 diabetes mellitus: the Atherosclerosis Risk in Communities Study. *Arch Intern Med* 159: 2151–2159, 1999.

271. Karandish M, Tamimi M, Shayesteh AA, Haghighizadeh MH, Jalali MT. The effect of magnesium supplementation and weight loss on liver enzymes in patients with non-alcoholic fatty liver disease. *J Res Med Sci* 18: 2013.
272. Karbach U. Cellular-mediated and diffusive magnesium transport across the descending colon of the rat. *Gastroenterology* 96: 1282–1289, 1989.
273. Karbach U, Rummel W. Cellular and paracellular magnesium transport across the terminal ileum of the rat and its interaction with the calcium transport. *Gastroenterology* 98: 985–992, 1990.
274. Katzberg HD, Khan AH, So YT. Assessment: symptomatic treatment for muscle cramps (an evidence-based review): report of the therapeutics and technology assessment subcommittee of the American academy of neurology. *Neurology* 74: 691–696, 2010.
275. Kawasaki T, Itoh K, Kawasaki M. Reduction in blood pressure with a sodium-reduced, potassium- and magnesium-enriched mineral salt in subjects with mild essential hypertension. *Hypertension Res* 21: 235–243, 1998.
276. Kazaks AG, Uriu-Adams JY, Albertson TE, Stern JS. Multiple measures of magnesium status are comparable in mild asthma and control subjects. *J Asthma* 43: 783–788, 2006.
277. Kelsen S, Hall JE, Chade AR. Endothelin-A receptor blockade slows the progression of renal injury in experimental renovascular disease. *Am J Physiol Renal Physiol* 301: F218–F225, 2011.
278. Kim BJ, Park EJ, Lee JH, Jeon JH, Kim SJ, So I. Suppression of transient receptor potential melastatin 7 channel induces cell death in gastric cancer. *Cancer Sci* 99: 2502–2509, 2008.
279. Kim JH, Gelbard AS, Djordjevic B, Kim SH, Perez AG. Action of daunomycin on the nucleic acid metabolism and viability of HeLa cells. *Cancer Res* 28: 2437–2442, 1968.
280. Kimble RB, Srivastava S, Ross FP, Matayoshi A, Pacifici R. Estrogen deficiency increases the ability of stromal cells to support murine osteoclastogenesis via an interleukin-1 and tumor necrosis factor-mediated stimulation of macrophage colony-stimulating factor production. *J Biol Chem* 271: 28890–28897, 1996.
281. King DE, Mainous AG, 3rd Geesey ME, Woolson RF. Dietary magnesium and C-reactive protein levels. *J Am Coll Nutr* 24: 166–171, 2005.
282. Kircelli F, Peter ME, Sevinc Ok E, Celenk FG, Yilmaz M, Steppan S, Ascì G, Ok E, Passlick-Deetjen J. Magnesium reduces calcification in bovine vascular smooth muscle cells in a dose-dependent manner. *Nephrol Dialysis Transplant* 27: 514–521, 2012.
283. Kleyman TR, Roberts C, Ling BN. A mechanism for pentamidine-induced hyperkalemia: inhibition of distal nephron sodium transport. *Ann Intern Med* 122: 103–106, 1995.
284. Knoers NV, Levchenko EN. Gitelman syndrome. *Orphanet J Rare Dis* 3: 22, 2008.
285. Kohvakka A, Heinonen L, Pietinen P, Salo H, Eisalo A. Potassium and magnesium balance in thiazide-treated cardiac patients with special reference to diet. *Acta Med Scand Suppl* 668: 102–109, 1982.
286. Koivisto M, Valta P, Hockerstedt K, Lindgren L. Magnesium depletion in chronic terminal liver cirrhosis. *Clin Transplant* 16: 325–328, 2002.
287. Kokko JP. Proximal tubule potential difference. Dependence on glucose on glucose, HCO₃, and amino acids. *J Clin Invest* 52: 1362–1367, 1973.
288. Kolisek M, Launay P, Beck A, Sponder G, Serafini N, Brenkus M, Froschauer EM, Martens H, Fleig A, Schweigel M. SLC41A1 is a novel mammalian Mg²⁺ carrier. *J Biol Chem* 283: 16235–16247, 2008.
289. Kolisek M, Nestler A, Vormann J, Schweigel-Rontgen M. Human gene SLC41A1 encodes for the Na⁺/Mg²⁺ exchanger. *Am J Physiol Cell Physiol* 302: C318–C326, 2012.
290. Kolisek M, Sponder G, Mastrototaro L, Smorodchenko A, Launay P, Vormann J, Schweigel-Rontgen M. Substitution p A350V in Na⁺/Mg²⁺ exchanger SLC41A1, potentially associated with Parkinson's Disease, is a gain-of-function mutation. *PLoS One* 8: e71096, 2013.
291. Komaki F, Akiyama T, Yamazaki T, Kitagawa H, Nosaka S, Shirai M. Effects of intravenous magnesium infusion on in vivo release of acetylcholine and catecholamine in rat adrenal medulla. *Auton Neurosci Basic Clin* 177: 123–128, 2013.
292. Konishi M. Cytoplasmic free concentrations of Ca²⁺ and Mg²⁺ in skeletal muscle fibers at rest and during contraction. *Jpn J Physiol* 48: 421–438, 1998.
293. Konrad M, Schaller A, Seelow D, Pandey AV, Waldegger S, Lesslauer A, Vitzthum H, Suzuki Y, Luk JM, Becker C, Schlingmann KP, Schmid M, Rodriguez-Soriano J, Ariceta G, Cano F, Enriquez R, Juppner H, Bakkaloglu SA, Hediger MA, Gallati S, Neuhaus SC, Nurnberg P, Weber S. Mutations in the tight-junction gene claudin 19 (CLDN19) are associated with renal magnesium wasting, renal failure, and severe ocular involvement. *Am J Hum Genet* 79: 949–957, 2006.
294. Koseoglu E, Talaslioglu A, Gonul AS, Kula M. The effects of magnesium prophylaxis in migraine without aura. *Magnesium Res* 21: 101–108, 2008.
295. Kuipers MT, Thang HD, Arntzenius AB. Hypomagnesaemia due to use of proton pump inhibitors—a review. *Netherlands J Med* 67: 169–172, 2009.
296. Kuramoto T, Kuwamura M, Tokuda S, Izawa T, Nakane Y, Kitada K, Akao M, Guenet JL, Serikawa T. A mutation in the gene encoding mitochondrial Mg²⁺ channel MRS2 results in demyelination in the rat. *PLoS Genet* 7: e1001262, 2011.
297. Lainez S, Schlingmann KP, van der Wijst J, Dworniczak B, van Zeeland F, Konrad M, Bindels RJ, Hoenderop JG. New TRPM6 missense mutations linked to hypomagnesemia with secondary hypocalcemia. *Eur J Hum Genet* 22: 497–504, 2014.
298. Lajer H, Daugaard G. Cisplatin and hypomagnesemia. *Cancer Treatment Rev* 25: 47–58, 1999.
299. Lambers TT, Bindels RJ, Hoenderop JG. Coordinated control of renal Ca²⁺ handling. *Kidney Int* 69: 650–654, 2006.
300. Lameris AL, Hess MW, van Kruijsbergen I, Hoenderop JG, Bindels RJ. Omeprazole enhances the colonic expression of the Mg²⁺ transporter TRPM6. *Pflügers Arch* 65: 1613–1620, 2013.
301. Lameris AL, Huybers S, Kaukinen K, Makela TH, Bindels RJ, Hoenderop JG, Nevalainen PI. Expression profiling of claudins in the human gastrointestinal tract in health and during inflammatory bowel disease. *Scand J Gastroenterol* 48: 58–69, 2013.
302. Larsson SC, Bergkvist L, Wolk A. Magnesium intake in relation to risk of colorectal cancer in women. *JAMA* 293: 86–89, 2005.
303. Laurant P, Berthelot A. Endothelin-I-induced contraction in isolated aortae from normotensive and DOCA-salt hypertensive rats: effect of magnesium. *Br J Pharmacol* 119: 1367–1374, 1996.
304. Le Grimellec C. Micropuncture study along the proximal convoluted tubule. Electrolyte reabsorption in first convolutions. *Pflügers Arch* 354: 133–150, 1975.
305. Ledeganck KJ, Boulet GA, Bogers JJ, Verpooten GA, De Winter BY. The TRPM6/EGF pathway is downregulated in a rat model of cisplatin nephrotoxicity. *PLoS One* 8: e57016, 2013.
306. Ledeganck KJ, Boulet GA, Horvath CA, Vinckx M, Bogers JJ, Van Den Bossche R, Verpooten GA, De Winter BY. Expression of renal distal tubule transporters TRPM6 and NCC in a rat model of cyclosporine nephrotoxicity and effect of EGF treatment. *Am J Physiol Renal Physiol* 301: F486–F493, 2011.
307. Lee CT, Chen HC, Ng HY, Lai LW, Lien YH. Renal adaptation to gentamicin-induced mineral loss. *Am J Nephrol* 35: 279–286, 2012.
308. Leo AAP. Spreading depression of activity in the cerebral cortex. *J Neurophysiol* 7: 359–390, 1944.
309. Leroy J. Necessite du magnesium pour la croissance de la souris. *CR Soc Biol* 94: 431, 1926.
310. Levine J, Stein D, Rapoport A, Kurtzman L. High serum and cerebrospinal fluid Ca/Mg ratio in recently hospitalized acutely depressed patients. *Neuropsychobiology* 39: 63–70, 1999.
311. Li FY, Chaigne-Delalande B, Kanellopoulou C, Davis JC, Matthews HF, Douek DC, Cohen JL, Uzel G, Su HC, Lenardo MJ. Second messenger role for Mg²⁺ revealed by human T-cell immunodeficiency. *Nature* 475: 471–476, 2011.
312. Li J, Zhang Q, Zhang M, Egger M. Intravenous magnesium for acute myocardial infarction. *Cochrane Database Systematic Rev* CD002755, 2007.
313. Li M, Du J, Jiang J, Ratzan W, Su LT, Runnels LW, Yue L. Molecular determinants of Mg²⁺ and Ca²⁺ permeability and pH sensitivity in TRPM6 and TRPM7. *J Biol Chem* 282: 25817–25830, 2007.

314. Li M, Jiang J, Yue L. Functional characterization of homo- and heteromeric channel kinases TRPM6 and TRPM7. *J Gen Physiol* 127: 525–537, 2006.
315. Liang RY, Wu W, Huang J, Jiang SP, Lin Y. Magnesium affects the cytokine secretion of CD4(+) T lymphocytes in acute asthma. *J Asthma* 49: 1012–1015, 2012.
316. Liao F, Folsom AR, Brancati FL. Is low magnesium concentration a risk factor for coronary heart disease? The Atherosclerosis Risk in Communities (ARIC) Study. *Am Heart J* 136: 480–490, 1998.
317. Lin CY, Tsai PS, Hung YC, Huang CJ. L-type calcium channels are involved in mediating the anti-inflammatory effects of magnesium sulphate. *Br J Anaesthesia* 104: 44–51, 2010.
318. Lin JY, Chung SY, Lin MC, Cheng FC. Effects of magnesium sulfate on energy metabolites and glutamate in the cortex during focal cerebral ischemia and reperfusion in the gerbil monitored by a dual-probe microdialysis technique. *Life Sci* 71: 803–811, 2002.
319. Lindahl T, Adams A, Fresco JR. Renaturation of transfer ribonucleic acids through site binding of magnesium. *Proc Natl Acad Sci USA* 55: 941–948, 1966.
320. Liu C, Yeh J, Aloia J. Magnesium directly stimulates osteoblast proliferation. *J Bone Miner Res* 3: S104, 1988.
321. Loffing J, Vallon V, Loffing-Cueni D, Aregger F, Richter K, Pietri L, Bloch-Faure M, Hoenderop JGJ, Shull GE, Meneton P, Kaissling B. Altered renal distal tubule structure and renal Na⁺ and Ca²⁺ handling in a mouse model for Gitelman's Syndrome. *J Am Soc Nephrol* 15: 2276–2288, 2004.
322. Lostrah AJ, Krahl ME. Magnesium, a second messenger for insulin: ion translocation coupled to transport activity. *Adv Enzyme Regul* 12: 73–81, 1974.
323. Lowenstein FW, Stanton MF. Serum magnesium levels in the United States, 1971–1974. *J Am Coll Nutr* 5: 399–414, 1986.
324. Ma J, Folsom AR, Melnick SL, Eckfeldt JH, Sharrett AR, Nabulsi AA, Hutchinson RG, Metcalf PA. Associations of serum and dietary magnesium with cardiovascular disease, hypertension, diabetes, insulin, and carotid arterial wall thickness: the ARIC study. Atherosclerosis Risk in Communities Study. *J Clin Epidemiol* 48: 927–940, 1995.
325. Mahabir S, Forman MR, Dong YQ, Park Y, Hollenbeck A, Schatzkin A. Mineral intake and lung cancer risk in the NIH-American Association of Retired Persons Diet and Health study. *Cancer Epidemiol Biomarkers Prevention* 19: 1976–1983, 2010.
326. Mahabir S, Wei Q, Barrera SL, Dong YQ, Etzel CJ, Spitz MR, Forman MR. Dietary magnesium and DNA repair capacity as risk factors for lung cancer. *Carcinogenesis* 29: 949–956, 2008.
327. Maier JA, Bernardini D, Rayssiguier Y, Mazur A. High concentrations of magnesium modulate vascular endothelial cell behaviour in vitro. *Biochim Biophys Acta* 1689: 6–12, 2004.
328. Maizels M, Blumenfeld A, Burchette R. A combination of riboflavin, magnesium, and feverfew for migraine prophylaxis: a randomized trial. *Headache* 44: 885–890, 2004.
329. Mandon B, Siga E, Roinel N, de Rouffignac C. Ca²⁺, Mg²⁺ and K⁺ transport in the cortical and medullary thick ascending limb of the rat nephron: influence of transepithelial voltage. *Pflügers Arch* 424: 558–560, 1993.
330. Martin BJ, Milligan K. Diuretic-associated hypomagnesemia in the elderly. *Arch Internal Med* 147: 1768–1771, 1987.
331. Massa F, Garbay S, Bouvier R, Sugitani Y, Noda T, Gubler MC, Heidet L, Pontoglio M, Fischer E. Hepatocyte nuclear factor 1beta controls nephron tubular development. *Development* 140: 886–896, 2013.
332. Mauskop A, Altura BT, Cracco RQ, Altura BM. Intravenous magnesium sulfate relieves cluster headaches in patients with low serum ionized magnesium levels. *Headache* 35: 597–600, 1995.
333. Mauskop A, Altura BT, Cracco RQ, Altura BM. Intravenous magnesium sulphate relieves migraine attacks in patients with low serum ionized magnesium levels: a pilot study. *Clin Sci* 89: 633–636, 1995.
334. Mauskop A, Varughese J. Why all migraine patients should be treated with magnesium. *J Neural Transm* 119: 575–579, 2012.
335. Mayer ML, Westbrook GL, Guthrie PB. Voltage-dependent block by Mg²⁺ of NMDA responses in spinal cord neurones. *Nature* 309: 261–263, 1984.
336. McCollister RJ, Flink EB, Lewis MD. Urinary excretion of magnesium in man following the ingestion of ethanol. *Am J Clin Nutr* 12: 415–420, 1963.
337. McDonald SD, Lutsiv O, Dzaja N, Duley L. A systematic review of maternal and infant outcomes following magnesium sulfate for pre-eclampsia/eclampsia in real-world use. *Int J Gynaecol Obstet* 118: 90–96, 2012.
338. McNeill DA, Herbein JH, Ritchey SJ. Hepatic gluconeogenic enzymes, plasma insulin and glucagon response to magnesium deficiency and fasting. *J Nutr* 112: 736–743, 1982.
339. Meij IC, Koenderink JB, van Bokhoven H, Assink KF, Groenestege WT, de Pont JJ, Bindels RJ, Monnens LA, van den Heuvel LP, Knoers NV. Dominant isolated renal magnesium loss is caused by misrouting of the Na⁺,K⁺-ATPase gamma-subunit. *Nature Genet* 26: 265–266, 2000.
340. Meij IC, van den Heuvel LP, Hemmes S, van der Vliet WA, Willems JL, Monnens LA, Knoers NV. Exclusion of mutations in FXYP2, CLDN16 and SLC12A3 in two families with primary renal Mg²⁺ loss. *Nephrol Dialysis Transplantation* 18: 512–516, 2003.
341. Melicher B, Kralickova P, Hyspler R, Kalabova H, Cerman J Jr, Holecckova P, Studen-tova H, Malirova E. Hypomagnesaemia in patients with metastatic colorectal carcinoma treated with cetuximab. *Hepatogastroenterology* 59: 366–371, 2012.
342. Michailova AP, Belik ME, McCulloch AD. Effects of magnesium on cardiac excitation-contraction coupling. *J Am Coll Nutr* 23: 514S–517S, 2004.
343. Milla PJ, Aggett PJ, Wolff OH, Harries JT. Studies in primary hypomagnesaemia: evidence for defective carrier-mediated small intestinal transport of magnesium. *Gut* 20: 1028–1033, 1979.
344. Milner RD, Hales CN. The role of calcium and magnesium in insulin secretion from rabbit pancreas studied in vitro. *Diabetologia* 3: 47–49, 1967.
345. Mishra NK, Peleg Y, Cirri E, Belogus T, Lifshitz Y, Voelker DR, Apell HJ, Garty H, Karlish SJ. FXYP proteins stabilize Na,K-ATPase: amplification of specific phosphatidylserine-protein interactions. *J Biol Chem* 286: 9699–9712, 2011.
346. Mishra R, Rao V, Ta R, Shobeiri N, Hill CE. Mg²⁺- and MgATP-inhibited and Ca²⁺/calmodulin-sensitive TRPM7-like current in hepatoma and hepatocytes. *Am J Physiol Gastrointest Liver Physiol* 297: G687–G694, 2009.
347. Misra VK, Draper DE. On the role of magnesium ions in RNA stability. *Biopolymers* 48: 113–135, 1998.
348. Mohammed S, Goodacre S. Intravenous and nebulised magnesium sulphate for acute asthma: systematic review and meta-analysis. *Emergency Med J* 24: 823–830, 2007.
349. Monteilh-Zoller MK, Hermosura MC, Nadler MJ, Scharenberg AM, Penner R, Fleig A. TRPM7 provides an ion channel mechanism for cellular entry of trace metal ions. *J Gen Physiol* 121: 49–60, 2003.
350. Mordike BL, Ebert T. Magnesium: Properties-applications-potential. *Materials Sci Eng* 302: 37–45, 2001.
351. Morris ME. Brain and CSF magnesium concentrations during magnesium deficit in animals and humans: neurological symptoms. *Magnesium Res* 5: 303–313, 1992.
352. Morton BC, Nair RC, Smith FM, McKibbin TG, Poznanski WJ. Magnesium therapy in acute myocardial infarction: a double-blind study. *Magnesium* 3: 346–352, 1984.
353. Moykkynen T, Uusi-Oukari M, Heikkilä J, Lovinger DM, Luddens H, Korpi ER. Magnesium potentiation of the function of native and recombinant GABA(A) receptors. *Neuroreport* 12: 2175–2179, 2001.
354. Mubagwa K, Gwanyanya A, Zakharov S, Macianskiene R. Regulation of cation channels in cardiac and smooth muscle cells by intracellular magnesium. *Arch Biochem Biophys* 458: 73–89, 2007.
355. Muir KW. Magnesium for neuroprotection in ischaemic stroke: rationale for use and evidence of effectiveness. *CNS Drugs* 15: 921–930, 2001.
356. Muir KW, Lees KR, Ford I, Davis S. Magnesium for acute stroke (Intravenous Magnesium Efficacy in Stroke trial): randomised controlled trial. *Lancet* 363: 439–445, 2004.
357. Murakami M, Ishizuka J, Sumi S, Nickols GA, Cooper CW, Townsend CM Jr, Thompson JC. Role of extracellular magnesium in insulin secretion from rat insulinoma cells. *Proc Soc Exp Biol Med* 200: 490–494, 1992.

358. Muroi C, Terzic A, Fortunati M, Yonekawa Y, Keller E. Magnesium sulfate in the management of patients with aneurysmal subarachnoid hemorrhage: a randomized, placebo-controlled, dose-adapted trial. *Surg Neurol* 69: 33–39, 2008.
359. Myrdal U, Leppert J, Edvinsson L, Ekman R, Hedner T, Nilsson H, Ringqvist I. Magnesium sulphate infusion decreases circulating calcitonin gene-related peptide (CGRP) in women with primary Raynaud's phenomenon. *Clin Physiol* 14: 539–546, 1994.
360. Nadler MJ, Hermosura MC, Inabe K, Perraud AL, Zhu Q, Stokes AJ, Kurosaki T, Kinet JP, Penner R, Scharenberg AM, Fleig A. LTRPC7 is a Mg-ATP-regulated divalent cation channel required for cell viability. *Nature* 411: 590–595, 2001.
361. Nair AV, Hocher B, Verkaar S, van Zeeland F, Pfab T, Slowinski T, Chen YP, Schlingmann KP, Schaller A, Gallati S, Bindels RJ, Konrad M, Hoenderop JG. Loss of insulin-induced activation of TRPM6 magnesium channels results in impaired glucose tolerance during pregnancy. *Proc Natl Acad Sci USA* 109: 11324–11329, 2012.
362. Nair RR, Nair P. Alteration of myocardial mechanics in marginal magnesium deficiency. *Magnesium Res* 15: 287–306, 2002.
363. Nicar MJ, Pak CY. Oral magnesium load test for the assessment of intestinal magnesium absorption. Application in control subjects, absorptive hypercalcaemia, primary hyperparathyroidism, and hypoparathyroidism. *Miner Electrolyte Metab* 8: 44–51, 1982.
364. Nicholls DG, Sihra TS. Synaptosomes possess an exocytotic pool of glutamate. *Nature* 321: 772–773, 1986.
365. Nijenhuis T, Hoenderop JG, Bindels RJ. Downregulation of Ca^{2+} and Mg^{2+} transport proteins in the kidney explains tacrolimus (FK506)-induced hypercalcaemia and hypomagnesaemia. *J Am Soc Nephrol* 15: 549–557, 2004.
366. Nijenhuis T, Hoenderop JG, Loffing J, van der Kemp AW, van Os CH, Bindels RJ. Thiazide-induced hypocalcaemia is accompanied by a decreased expression of Ca^{2+} transport proteins in kidney. *Kidney Int* 64: 555–564, 2003.
367. Nijenhuis T, Renkema KY, Hoenderop JG, Bindels RJ. Acid-base status determines the renal expression of Ca^{2+} and Mg^{2+} transport proteins. *J Am Soc Nephrol* 17: 617–626, 2006.
368. Nijenhuis T, Vallon V, van der Kemp AW, Loffing J, Hoenderop JG, Bindels RJ. Enhanced passive Ca^{2+} reabsorption and reduced Mg^{2+} channel abundance explains thiazide-induced hypocalcaemia and hypomagnesaemia. *J Clin Invest* 115: 1651–1658, 2005.
369. Noormohamed FH, Youle MS, Tang B, Martin-Munley S, Gazzard BG, Lant AF. Foscarnet-induced changes in plasma concentrations of total and ionized calcium and magnesium in HIV-positive patients. *Antiviral Ther* 1: 172–179, 1996.
370. Nouria S, Bouida W, Grissa MH, Beltaief K, Trimech MN, Boubaker H, Marghli S, Letaief M, Boukef R. Magnesium sulfate versus ipratropium bromide in chronic obstructive pulmonary disease exacerbation: a randomized trial. *Am J Ther* 21: 152–158, 2014.
371. Novelli F, Malagrino L, Dente FL, Paggiaro P. Efficacy of anticholinergic drugs in asthma. *Expert Rev Respir Med* 6: 309–319, 2012.
372. Nowak L, Bregestovski P, Ascher P, Herbet A, Prochiantz A. Magnesium gates glutamate-activated channels in mouse central neurones. *Nature* 307: 462–465, 1984.
373. Okahira M, Kubota M, Iguchi K, Usui S, Hirano K. Regulation of aquaporin 3 expression by magnesium ion. *Eur J Pharmacol* 588: 26–32, 2008.
374. Okayama H, Aikawa T, Okayama M, Sasaki H, Mue S, Takishima T. Bronchodilating effect of intravenous magnesium sulfate in bronchial asthma. *JAMA* 257: 1076–1078, 1987.
375. Oladipo OO, Ajala MO, Okubadejo N, Danesi MA, Afonja OA. Plasma magnesium in adult Nigerian patients with epilepsy. *Nigerian Postgrad Med J* 10: 234–237, 2003.
376. Olesen J. The role of nitric oxide (NO) in migraine, tension-type headache and cluster headache. *Pharmacol Ther* 120: 157–171, 2008.
377. Olinger E, Schwaller B, Loffing J, Gailly P, Devuyst O. Parvalbumin: calcium and magnesium buffering in the distal nephron. *Nephrol Dialysis Transplantation* 27: 3988–3994, 2012.
378. Onishi S, Yoshino S. Cathartic-induced fatal hypermagnesaemia in the elderly. *Intern Med* 45: 207–210, 2006.
379. Oren S, Rapoport J, Zlotnik M, Brami JL, Heimer D, Chaimovitz C. Extreme hypermagnesaemia due to ingestion of Dead Sea water. *Nephron* 47: 199–201, 1987.
380. Orenstein SR, Orenstein DM. Magnesium deficiency in cystic fibrosis. *Southern Med J* 76: 1586, 1983.
381. Oyanagi K, Kawakami E, Kikuchi-Horie K, Ohara K, Ogata K, Takahama S, Wada M, Kihira T, Yasui M. Magnesium deficiency over generations in rats with special references to the pathogenesis of the Parkinsonism-dementia complex and amyotrophic lateral sclerosis of Guam. *Neuropathology* 26: 115–128, 2006.
382. Pages N, Gogly B, Godeau G, Igonjo-Tchen S, Maurois P, Durlach J, Bac P. Structural alterations of the vascular wall in magnesium-deficient mice. A possible role of gelatinases A (MMP-2) and B (MMP-9). *Magnesium Res* 16: 43–48, 2003.
383. Pandey M, Gupta A, Baduni N, Vijfadar H, Sinha S, Jain A. Refractory status epilepticus—magnesium as rescue therapy. *Anaesthesia Intensive Care* 38: 962, 2010.
384. Paoletti P, Bellone C, Zhou Q. NMDA receptor subunit diversity: impact on receptor properties, synaptic plasticity and disease. *Nature Rev Neurosci* 14: 383–400, 2013.
385. Paolisso G, Sgambato S, Pizzi G, Passariello N, Varricchio M, D'Onofrio F. Improved insulin response and action by chronic magnesium administration in aged NIDDM subjects. *Diabetes Care* 12: 265–269, 1989.
386. Pardutz A, Vecsei L. Should magnesium be given to every migraineur? No. *J Neural Transm* 119: 581–585, 2012.
387. Parry DA, Mighell AJ, El-Sayed W, Shore RC, Jalili IK, Dollfus H, Bloch-Zupan A, Carlos R, Carr IM, Downey LM, Blain KM, Mansfield DC, Shahrabi M, Heidari M, Aref P, Abbasi M, Michaelides M, Moore AT, Kirkham J, Inglehearn CF. Mutations in CNNM4 cause Jalili syndrome, consisting of autosomal-recessive cone-rod dystrophy and amelogenesis imperfecta. *Am J Hum Genet* 84: 266–273, 2009.
388. Parsons AA. Cortical spreading depression: its role in migraine pathogenesis and possible therapeutic intervention strategies. *Curr Pain Headache Rep* 8: 410–416, 2004.
389. Partridge IG. Studies on digestion and absorption in the intestines of growing pigs. 3. Net movements of mineral nutrients in the digestive tract. *Br J Nutr* 39: 527–537, 1978.
390. Paunier L, Radde IC, Kooh SW, Conen PE, Fraser D. Primary hypomagnesaemia with secondary hypocalcaemia in an infant. *Pediatrics* 41: 385–402, 1968.
391. Peacock JM, Ohira T, Post W, Sotoodehnia N, Rosamond W, Folsom AR. Serum magnesium and risk of sudden cardiac death in the Atherosclerosis Risk in Communities (ARIC) Study. *Am Heart J* 160: 464–470, 2010.
392. Pearson PJ, Evora PR, Seccombe JF, Schaff HV. Hypomagnesaemia inhibits nitric oxide release from coronary endothelium: protective role of magnesium infusion after cardiac operations. *Ann Thoracic Surg* 65: 967–972, 1998.
393. Peikert A, Wilimzig C, Kohne-Volland R. Prophylaxis of migraine with oral magnesium: results from a prospective, multi-center, placebo-controlled and double-blind randomized study. *Cephalalgia* 16: 257–263, 1996.
394. Pelletier H, Sawaya MR, Kumar A, Wilson SH, Kraut J. Structures of ternary complexes of rat DNA polymerase beta, a DNA template-primer, and ddCTP. *Science* 264: 1891–1903, 1994.
395. Perticone F, Adinolfi L, Bonaduce D. Efficacy of magnesium sulfate in the treatment of torsade de pointes. *Am Heart J* 112: 847–849, 1986.
396. Petrelli F, Borgonovo K, Cabiddu M, Ghilardi M, Barni S. Risk of anti-EGFR monoclonal antibody-related hypomagnesaemia: systematic review and pooled analysis of randomized studies. *Expert Opin Drug Safety* 11 Suppl 1: S9–19, 2012.
397. Pfaffenrath V, Wessely P, Meyer C, Isler HR, Evers S, Grottemeyer KH, Taneri Z, Soyka D, Gobel H, Fischer M. Magnesium in the prophylaxis of migraine—a double-blind placebo-controlled study. *Cephalalgia* 16: 436–440, 1996.
398. Pieper MP. The non-neuronal cholinergic system as novel drug target in the airways. *Life Sci* 91: 1113–1118, 2012.
399. Piskacek M, Zotova L, Zsurka G, Schweyen RJ. Conditional knockdown of hMRS2 results in loss of mitochondrial Mg^{2+} uptake and cell death. *J Cell Mol Med* 13: 693–700, 2009.

400. Pittenger C, Sanacora G, Krystal JH. The NMDA receptor as a therapeutic target in major depressive disorder. *CNS Neurol Disorders Drug Targets* 6: 101–115, 2007.
401. Pokan R, Hofmann P, von Duvillard SP, Smekal G, Wonisch M, Lettner K, Schmid P, Shechter M, Silver B, Bachl N. Oral magnesium therapy, exercise heart rate, exercise tolerance, and myocardial function in coronary artery disease patients. *Br J Sports Med* 40: 773–778, 2006.
402. Polok B, Escher P, Ambresin A, Chouery E, Bolay S, Meunier I, Nan F, Hamel C, Munier FL, Thilo B, Megarbane A, Schorderet DF. Mutations in CNNM4 cause recessive cone-rod dystrophy with amelogenesis imperfecta. *Am J Hum Genet* 84: 259–265, 2009.
403. Potter JD, Gergely J. The calcium and magnesium binding sites on troponin and their role in the regulation of myofibrillar adenosine triphosphatase. *J Biol Chem* 250: 4628–4633, 1975.
404. Powell C, Dwan K, Milan SJ, Beasley R, Hughes R, Knopp-Sihota JA, Rowe BH. Inhaled magnesium sulfate in the treatment of acute asthma. *Cochrane Database Systematic Rev* 12: CD003898, 2012.
405. Powell C, Kolamunnage-Dona R, Lowe J, Boland A, Petrou S, Doull I, Hood K, Williamson P. MAGNESium Trial In Children (MAGNETIC): a randomised, placebo-controlled trial and economic evaluation of nebulised magnesium sulphate in acute severe asthma in children. *Health Technol Assess* 17: 1–216, 2013.
406. Price MA, Tullius TD. II. Using hydroxyl radical to probe DNA structure. In: *Methods in Enzymology*, edited by David M. J. Lilley. New York: Academic, 1992, p. 194–219.
407. Pritchard JA, Pritchard SA. Standardized treatment of 154 consecutive cases of eclampsia. *Am J Obstet Gynecol* 123: 543–552, 1975.
408. Quamme GA. Effect of furosemide on calcium and magnesium transport in the rat nephron. *Am J Physiol Renal Fluid Electrolyte Physiol* 241: F340–F347, 1981.
409. Quamme GA. Recent developments in intestinal magnesium absorption. *Curr Opin Gastroenterol* 24: 230–235, 2008.
410. Quamme GA, Smith CM. Magnesium transport in the proximal straight tubule of the rabbit. *Am J Physiol Renal Fluid Electrolyte Physiol* 246: F544–F550, 1984.
411. Quamme GA, Wong NL, Dirks JH, Roinel N, De Rouffignac C, Morel F. Magnesium handling in the dog kidney: a micropuncture study. *Pflügers Arch* 377: 95–99, 1978.
412. Quigley GJ, Teeter MM, Rich A. Structural analysis of spermine and magnesium ion binding to yeast phenylalanine transfer RNA. *Proc Natl Acad Sci USA* 75: 64–68, 1978.
413. Qureshi T, Melonakos TK. Acute hypermagnesemia after laxative use. *Ann Emergency Med* 28: 552–555, 1996.
414. Ramadan NM, Halvorson H, Vande-Linde A, Levine SR, Helpert JA, Welch KM. Low brain magnesium in migraine. *Headache* 29: 590–593, 1989.
415. Rasmussen HS, McNair P, Norregard P, Backer V, Lindeneg O, Balslev S. Intravenous magnesium in acute myocardial infarction. *Lancet* 1: 234–236, 1986.
416. Regan RF, Jasper E, Guo Y, Panter SS. The effect of magnesium on oxidative neuronal injury in vitro. *J Neurochem* 70: 77–85, 1998.
417. Reichold M, Zdebek AA, Lieberer E, Rapedius M, Schmidt K, Bandulik S, Sterner C, Tegtmeier I, Penton D, Baukowitz T, Hulton SA, Witzgall R, Ben-Zeev B, Howie AJ, Kleta R, Bockenauer D, Warth R. KCNJ10 gene mutations causing EAST syndrome (epilepsy, ataxia, sensorineural deafness, and tubulopathy) disrupt channel function. *Proc Natl Acad Sci USA* 107: 14490–14495, 2010.
418. Reis MA, Latorraca MQ, Carneiro EM, Boschero AC, Saad MJ, Velloso LA, Reyes FG. Magnesium deficiency improves glucose homeostasis in the rat: studies in vivo and in isolated islets in vitro. *Br J Nutr* 85: 549–552, 2001.
419. Reis MA, Reyes FG, Saad MJ, Velloso LA. Magnesium deficiency modulates the insulin signaling pathway in liver but not muscle of rats. *J Nutr* 130: 133–138, 2000.
420. Renkema KY, Alexander RT, Bindels RJ, Hoenderop JG. Calcium and phosphate homeostasis: concerted interplay of new regulators. *Ann Med* 40: 82–91, 2008.
421. Rodriguez-Hernandez H, Cervantes-Huerta M, Rodriguez-Moran M, Guerrero-Romero F. Oral magnesium supplementation decreases alanine aminotransferase levels in obese women. *Magnesium Res* 23: 90–96, 2010.
422. Rodriguez-Moran M, Guerrero-Romero F. Insulin secretion is decreased in non-diabetic individuals with hypomagnesaemia. *Diabetes/Metab Res Rev* 27: 590–596, 2011.
423. Rodriguez-Moran M, Guerrero-Romero F. Oral magnesium supplementation improves insulin sensitivity and metabolic control in type 2 diabetic subjects: a randomized double-blind controlled trial. *Diabetes Care* 26: 1147–1152, 2003.
424. Romani A. Regulation of magnesium homeostasis and transport in mammalian cells. *Arch Biochem Biophys* 458: 90–102, 2007.
425. Rosanoff A, Plesset MR. Oral magnesium supplements decrease high blood pressure (SBP > 155 mmHg) in hypertensive subjects on anti-hypertensive medications: a targeted meta-analysis. *Magnesium Res* 26: 93–99, 2013.
426. Rosenacker J, Naundorf S, Rudolph C. Airway surface liquid contains endogenous DNase activity which can be activated by exogenous magnesium. *Eur J Med Res* 14: 304–308, 2009.
427. Rowe BH, Camargo CA Jr. The role of magnesium sulfate in the acute and chronic management of asthma. *Curr Opin Pulmon Med* 14: 70–76, 2008.
428. Rubin AH, Terasaki M, Sanui H. Magnesium reverses inhibitory effects of calcium deprivation on coordinate response of 3T3 cells to serum. *Proc Natl Acad Sci USA* 75: 4379–4383, 1978.
429. Rubin AH, Terasaki M, Sanui H. Major intracellular cations and growth control: correspondence among magnesium content, protein synthesis, and the onset of DNA synthesis in BALB/c3T3 cells. *Proc Natl Acad Sci USA* 76: 3917–3921, 1979.
430. Rubin H. Central role for magnesium in coordinate control of metabolism and growth in animal cells. *Proc Natl Acad Sci USA* 72: 3551–3555, 1975.
431. Rubin H. The logic of the Membrane, Magnesium, Mitosis (MMM) model for the regulation of animal cell proliferation. *Arch Biochem Biophys* 458: 16–23, 2007.
432. Rubin H. The membrane, magnesium, mitosis (MMM) model of cell proliferation control. *Magnesium Res* 18: 268–274, 2005.
433. Rude RK. Magnesium depletion and hypermagnesemia. *Primer Metab Bone Dis Disorders Miner Metab* 1: 2006.
434. Rude RK, Gruber HE, Norton HJ, Wei LY, Frausto A, Mills BG. Bone loss induced by dietary magnesium reduction to 10% of the nutrient requirement in rats is associated with increased release of substance P and tumor necrosis factor- α . *J Nutr* 134: 79–85, 2004.
435. Rude RK, Oldham SB, Sharp CF Jr, Singer FR. Parathyroid hormone secretion in magnesium deficiency. *J Clin Endocrinol Metab* 47: 800–806, 1978.
436. Rude RK, Olerich M. Magnesium deficiency: possible role in osteoporosis associated with gluten-sensitive enteropathy. *Osteoporosis Int* 6: 453–461, 1996.
437. Rukshin V, Shah PK, Cercek B, Finkelstein A, Tsang V, Kaul S. Comparative antithrombotic effects of magnesium sulfate and the platelet glycoprotein IIb/IIIa inhibitors tirofiban and eptifibatid in a canine model of stent thrombosis. *Circulation* 105: 1970–1975, 2002.
438. Runnels LW, Yue L, Clapham DE. TRP-PLIK, a bifunctional protein with kinase and ion channel activities. *Science* 291: 1043–1047, 2001.
439. Sabbagh F, El Tawil Z, Lecerf F, Hulin A, Maurois P, Dartevelle P, Bac P, German-Fattal M. Impact of cyclosporine A on magnesium homeostasis: clinical observation in lung transplant recipients and experimental study in mice. *Transplantation* 86: 436–444, 2008.
440. Sabbagh F, Lecerf F, Maurois P, Bac P, German-Fattal M. Allogeneic activation is attenuated in a model of mouse lung perfused with magnesium-deficient blood. *Transplant Immunol* 16: 200–207, 2006.
441. Sahin G, Ertem U, Duru F, Birgen D, Yuksek N. High prevalence of chronic magnesium deficiency in T cell lymphoblastic leukemia and chronic zinc deficiency in children with acute lymphoblastic leukemia and malignant lymphoma. *Leukemia Lymphoma* 39: 555–562, 2000.
442. Sahni J, Nelson B, Scharenberg AM. SLC41A2 encodes a plasma-membrane Mg²⁺ transporter. *Biochem J* 401: 505–513, 2007.
443. Salimi MH, Heughebaert JC, Nancollas GH. Crystal growth of calcium phosphates in the presence of magnesium ions. *Langmuir* 1: 119–122, 1985.

444. Sanders NN, Franckx H, De Boeck K, Haustraete J, De Smedt SC, Demeester J. Role of magnesium in the failure of rhDNase therapy in patients with cystic fibrosis. *Thorax* 61: 962–968, 2006.
445. Sanui H, Rubin AH. Membrane bound and cellular cationic changes associated with insulin stimulation of cultured cells. *J Cell Physiol* 96: 265–278, 1978.
446. Sassen MC, Kim SW, Kwon TH, Knepper MA, Miller RT, Frokiaer J, Nielsen S. Dysregulation of renal sodium transporters in gentamicin-treated rats. *Kidney Int* 70: 1026–1037, 2006.
447. Satake K, Lee JD, Shimizu H, Uzui H, Mitsuke Y, Yue H, Ueda T. Effects of magnesium on prostacyclin synthesis and intracellular free calcium concentration in vascular cells. *Magnesium Res* 17: 20–27, 2004.
448. Saver JL. Targeting the brain: neuroprotection and neurorestoration in ischemic stroke. *Pharmacotherapy* 30: 62S–69S, 2010.
449. Saver JL, Kidwell C, Eckstein M, Starkman S. Prehospital neuroprotective therapy for acute stroke: results of the Field Administration of Stroke Therapy-Magnesium (FAST-MAG) pilot trial. *Stroke* 35: e106–108, 2004.
450. Scanlan BJ, Tuft B, Elfrey JE, Smith A, Zhao A, Morimoto M, Chmielinska JJ, Tejero-Taldo MI, Mak lu T, Weglicki WB, Shea-Donohue T. Intestinal inflammation caused by magnesium deficiency alters basal and oxidative stress-induced intestinal function. *Mol Cell Biochem* 306: 59–69, 2007.
451. Schelling JR. Fatal hypermagnesemia. *Clin Nephrol* 53: 61–65, 2000.
452. Schiffrin EL, Lipman ML, Mann JF. Chronic kidney disease: effects on the cardiovascular system. *Circulation* 116: 85–97, 2007.
453. Schilsky RL, Anderson T. Hypomagnesemia and renal magnesium wasting in patients receiving cisplatin. *Ann Internal Med* 90: 929–931, 1979.
454. Schlingmann KP, Weber S, Peters M, Niemann Nejsum L, Vitzthum H, Klingel K, Kratz M, Haddad E, Ristoff E, Dinour D, Syrrou M, Nielsen S, Sassen M, Waldegger S, Seyberth HW, Konrad M. Hypomagnesemia with secondary hypocalcemia is caused by mutations in TRPM6, a new member of the TRPM gene family. *Nature Genet* 31: 166–170, 2002.
455. Schmitz C, Perraud AL, Johnson CO, Inabe K, Smith MK, Penner R, Kurosaki T, Fleig A, Scharenberg AM. Regulation of vertebrate cellular Mg^{2+} homeostasis by TRPM7. *Cell* 114: 191–200, 2003.
456. Schneeberger PR, Heizmann CW. Parvalbumin in rat kidney. Purification and localization. *FEBS Lett* 201: 51–56, 1986.
457. Schoenen J, Sianard-Gainko J, Lenaerts M. Blood magnesium levels in migraine. *Cephalalgia* 11: 97–99, 1991.
458. Scholl UI, Choi M, Liu T, Ramaekers VT, Hausler MG, Grimmer J, Tobe SW, Farhi A, Nelson-Williams C, Lifton RP. Seizures, sensorineural deafness, ataxia, mental retardation, and electrolyte imbalance (SeSAME syndrome) caused by mutations in KCNJ10. *Proc Natl Acad Sci USA* 106: 5842–5847, 2009.
459. Schrag D, Chung KY, Flombaum C, Saltz L. Cetuximab therapy and symptomatic hypomagnesemia. *J Natl Cancer Inst* 97: 1221–1224, 2005.
460. Schultheis PJ, Lorenz JN, Meneton P, Nieman ML, Riddle TM, Flagella M, Duffy JJ, Doetschman T, Miller ML, Shull GE. Phenotype resembling Gitelman's Syndrome in mice lacking the apical $Na^{+}-Cl^{-}$ cotransporter of the distal convoluted tubule. *J Biol Chem* 273: 29150–29155, 1998.
461. Schweigel M, Martens H. Magnesium transport in the gastrointestinal tract. *Front Biosci* 5: D666–677, 2000.
462. Secil Y, Unde C, Beckmann YY, Bozkaya YT, Ozerkan F, Basoglu M. Blood pressure changes in migraine patients before, during and after migraine attacks. *Pain Pract* 10: 222–227, 2010.
463. Seelig MS, Elin RJ, Antman EM. Magnesium in acute myocardial infarction: still an open question. *Can J Cardiol* 14: 745–749, 1998.
464. Sha Q, Pearson W, Burcea LC, Wigfall DA, Schlesinger PH, Nichols CG, Mercer RW. Human FXVD2 G4 IR mutation responsible for renal hypomagnesemia behaves as an inward-rectifying cation channel. *Am J Physiol Renal Physiol* 295: F91–F99, 2008.
465. Shah GM, Alvarado P, Kirschenbaum MA. Symptomatic hypocalcemia and hypomagnesemia with renal magnesium wasting associated with pentamidine therapy in a patient with AIDS. *Am J Med* 89: 380–382, 1990.
466. Shan Q, Himmerkus N, Hou J, Goodenough DA, Bleich M. Insights into driving forces and paracellular permeability from claudin-16 knockdown mouse. *Ann NY Acad Sci* 1165: 148–151, 2009.
468. Shareghi GR, Agus ZS. Magnesium transport in the cortical thick ascending limb of Henle's loop of the rabbit. *J Clin Invest* 69: 759–769, 1982.
469. Shechter M, Hod H, Chouraqui P, Kaplinsky E, Rabinowitz B. Magnesium therapy in acute myocardial infarction when patients are not candidates for thrombolytic therapy. *Am J Cardiol* 75: 321–323, 1995.
470. Shechter M, Hod H, Marks N, Behar S, Kaplinsky E, Rabinowitz B. Beneficial effect of magnesium sulfate in acute myocardial infarction. *Am J Cardiol* 66: 271–274, 1990.
471. Shechter M, Merz CN, Paul-Labrador M, Meisel SR, Rude RK, Molloy MD, Dwyer JH, Shah PK, Kaul S. Oral magnesium supplementation inhibits platelet-dependent thrombosis in patients with coronary artery disease. *Am J Cardiol* 84: 152–156, 1999.
472. Shechter M, Sharir M, Labrador MJ, Forrester J, Silver B, Bairey Merz CN. Oral magnesium therapy improves endothelial function in patients with coronary artery disease. *Circulation* 102: 2353–2358, 2000.
473. Shen B, Nolan JP, Sklar LA, Park MS. Essential amino acids for substrate binding and catalysis of human flap endonuclease I. *J Biol Chem* 271: 9173–9176, 1996.
474. Shiga T, Wajima Z, Inoue T, Ogawa R. Magnesium prophylaxis for arrhythmias after cardiac surgery: a meta-analysis of randomized controlled trials. *Am J Med* 117: 325–333, 2004.
475. Shindo Y, Fujii T, Komatsu H, Citterio D, Hotta K, Suzuki K, Oka K. Newly developed Mg^{2+} -selective fluorescent probe enables visualization of Mg^{2+} dynamics in mitochondria. *PLoS One* 6: e23684, 2011.
476. Shorvon S, Ferlisi M. The treatment of super-refractory status epilepticus: a critical review of available therapies and a clinical treatment protocol. *Brain* 134: 2802–2818, 2011.
477. Simon DB, Bindra RS, Mansfield TA, Nelson-Williams C, Mendonca E, Stone R, Schurman S, Nayir A, Alpay H, Bakkaloglu A, Rodriguez-Soriano J, Morales JM, Sanjad SA, Taylor CM, Pilz D, Brem A, Trachtman H, Griswold W, Richard GA, John E, Lifton RP. Mutations in the chloride channel gene, CLCNKB, cause Bartter's syndrome type III. *Nature Genet* 17: 171–178, 1997.
478. Simon DB, Karet FE, Hamdan JM, DiPietro A, Sanjad SA, Lifton RP. Bartter's syndrome, hypokalaemic alkalosis with hypercalciuria, is caused by mutations in the Na-K-2Cl cotransporter NKCC2. *Nature Genet* 13: 183–188, 1996.
479. Simon DB, Karet FE, Rodriguez-Soriano J, Hamdan JH, DiPietro A, Trachtman H, Sanjad SA, Lifton RP. Genetic heterogeneity of Bartter's syndrome revealed by mutations in the K^{+} channel, ROMK. *Nature Genet* 14: 152–156, 1996.
480. Simon DB, Lu Y, Choate KA, Velazquez H, Al-Sabban E, Praga M, Casari G, Bettinelli A, Colussi G, Rodriguez-Soriano J, McCredie D, Milford D, Sanjad S, Lifton RP. Paracellin-1, a renal tight junction protein required for paracellular Mg^{2+} resorption. *Science* 285: 103–106, 1999.
481. Simon DB, Nelson-Williams C, Bia MJ, Ellison D, Karet FE, Molina AM, Vaara I, Iwata F, Cushner HM, Koolen M, Gainza FJ, Gittleman HJ, Lifton RP. Gitelman's variant of Bartter's syndrome, inherited hypokalaemic alkalosis, is caused by mutations in the thiazide-sensitive Na-Cl cotransporter. *Nature Genet* 12: 24–30, 1996.
482. Sinert R, Zehtabchi S, Desai S, Peacock P, Altura BT, Altura BM. Serum ionized magnesium and calcium levels in adult patients with seizures. *Scand J Clin Lab Invest* 67: 317–326, 2007.
483. Singh J, Wisdom DM. Second messenger role of magnesium in pancreatic acinar cells of the rat. *Mol Cell Biochem* 149–150: 175–182, 1995.
484. Singh RB, Pella D, Neki NS, Chandel JP, Rastogi S, Mori H, Otsuka K, Gupta P. Mechanisms of acute myocardial infarction study (MAMIS). *Biomed Pharmacotherapy* 58 Suppl 1: S111–115, 2004.

485. Skorodin MS, Tenholder MF, Yetter B, Owen KA, Waller RF, Khandelwahi S, Maki K, Rohail T, D'Alfonso N. Magnesium sulfate in exacerbations of chronic obstructive pulmonary disease. *Arch Intern Med* 155: 496–500, 1995.
486. Smith DA, Connick JH, Stone TW. Effect of changing extracellular levels of magnesium on spontaneous activity and glutamate release in the mouse neocortical slice. *Br J Pharmacol* 97: 475–482, 1989.
487. Smith LF, Heagerty AM, Bing RF, Barnett DB. Intravenous infusion of magnesium sulphate after acute myocardial infarction: effects on arrhythmias and mortality. *Int J Cardiol* 12: 175–183, 1986.
488. Song Y, He K, Levitan EB, Manson JE, Liu S. Effects of oral magnesium supplementation on glycaemic control in Type 2 diabetes: a meta-analysis of randomized double-blind controlled trials. *Diabetic Med* 23: 1050–1056, 2006.
489. Song Y, Hsu YH, Niu T, Manson JE, Buring JE, Liu S. Common genetic variants of the ion channel transient receptor potential membrane melastatin 6 and 7 (TRPM6 and TRPM7), magnesium intake, and risk of type 2 diabetes in women. *BMC Med Genet* 10: 4, 2009.
490. Song Y, Li TY, van Dam RM, Manson JE, Hu FB. Magnesium intake and plasma concentrations of markers of systemic inflammation and endothelial dysfunction in women. *Am J Clin Nutr* 85: 1068–1074, 2007.
491. Song Y, Manson JE, Buring JE, Liu S. Dietary magnesium intake in relation to plasma insulin levels and risk of type 2 diabetes in women. *Diabetes Care* 27: 59–65, 2004.
492. Sood AK, Handa R, Malhotra RC, Gupta BS. Serum, CSF, RBC & urinary levels of magnesium & calcium in idiopathic generalised tonic clonic seizures. *Indian J Med Res* 98: 152–154, 1993.
493. Steinert JR, Chernova T, Forsythe ID. Nitric oxide signaling in brain function, dysfunction, and dementia. *Neuroscientist* 16: 435–452, 2010.
494. Stendig-Lindberg G, Tepper R, Leichter I. Trabecular bone density in a two year controlled trial of peroral magnesium in osteoporosis. *Magnesium Res* 6: 155–163, 1993.
495. Stohr W, Paulides M, Bielack S, Jurgens H, Koscielniak E, Rossi R, Langer T, Beck JD. Nephrotoxicity of cisplatin and carboplatin in sarcoma patients: a report from the late effects surveillance system. *Pediatr Blood Cancer* 48: 140–147, 2007.
496. Storchheim F. Status epilepticus treated by magnesium sulphate, injected intravenously. *JAMA* 101: 1313–1314, 1933.
497. Stuiver M, Lainez S, Will C, Terryn S, Gunzel D, Debaix H, Sommer K, Kopplin K, Thumfart J, Kampik NB, Querfeld U, Willnow TE, Nemeč V, Wagner CA, Hoenderop JG, Devuyt O, Knoers NV, Bindels RJ, Meij IC, Muller D. CNNM2, encoding a basolateral protein required for renal Mg²⁺ handling, is mutated in dominant hypomagnesemia. *Am J Hum Genet* 88: 333–343, 2011.
498. Suarez A, Pulido N, Casla A, Casanova B, Arrieta FJ, Rovira A. Impaired tyrosine-kinase activity of muscle insulin receptors from hypomagnesaemic rats. *Diabetologia* 38: 1262–1270, 1995.
499. Sudo GZ, Sanguinetti MC. Intracellular [Mg²⁺] determines specificity of K⁺ channel block by a class III antiarrhythmic drug. *J Pharmacol Exp Ther* 276: 951–957, 1996.
500. Suh WC, Leirmo S, Record MT Jr. Roles of Mg²⁺ in the mechanism of formation and dissociation of open complexes between *Escherichia coli* RNA polymerase and the lambda PR promoter: kinetic evidence for a second open complex requiring Mg²⁺. *Biochemistry* 31: 7815–7825, 1992.
501. Sun Y, Selvaraj S, Varma A, Derry S, Sahnoun AE, Singh BB. Increase in serum Ca²⁺/Mg²⁺ ratio promotes proliferation of prostate cancer cells by activating TRPM7 channels. *J Biol Chem* 288: 255–263, 2013.
502. Takata Y, Shu XO, Yang G, Li H, Dai Q, Gao J, Cai Q, Gao YT, Zheng W. Calcium intake and lung cancer risk among female nonsmokers: a report from the Shanghai Women's Health Study. *Cancer Epidemiol Biomarkers Prevention* 22: 50–57, 2013.
503. Takaya J, Higashino H, Kobayashi Y. Can magnesium act as a second messenger? Current data on translocation induced by various biologically active substances. *Magnesium Res* 13: 139–146, 2000.
504. Takezawa R, Schmitz C, Demeuse P, Scharenberg AM, Penner R, Fleig A. Receptor-mediated regulation of the TRPM7 channel through its endogenous protein kinase domain. *Proc Natl Acad Sci USA* 101: 6009–6014, 2004.
505. Taniguchi CM, Emanuelli B, Kahn CR. Critical nodes in signalling pathways: insights into insulin action. *Nature Rev Mol Cell Biol* 7: 85–96, 2006.
506. Tashiro M, Inoue H, Konishi M. Magnesium homeostasis in cardiac myocytes of mg-deficient rats. *PLoS One* 8: e73171, 2013.
507. Taubert K. [Magnesium in migraine. Results of a multicenter pilot study]. *Fortschritte der Medizin* 112: 328–330, 1994.
508. Taylor JS, Vigneron DB, Murphy-Boesch J, Nelson SJ, Kessler HB, Coia L, Curran W, Brown TR. Free magnesium levels in normal human brain and brain tumors: ³¹P chemical-shift imaging measurements at 1.5 T. *Proc Natl Acad Sci USA* 88: 6810–6814, 1991.
509. Tejero-Taldo MI, Chmielinska JJ, Gonzalez G, Mak IT, Weglicki WB. N-methyl-D-aspartate receptor blockade inhibits cardiac inflammation in the Mg²⁺-deficient rat. *J Pharmacol Exp Ther* 311: 8–13, 2004.
510. Tejpar S, Piessevaux H, Claes K, Piront P, Hoenderop JG, Verslype C, Van Cutsem E. Magnesium wasting associated with epidermal-growth-factor receptor-targeting antibodies in colorectal cancer: a prospective study. *Lancet Oncol* 8: 387–394, 2007.
511. Temkin NR, Anderson GD, Winn HR, Ellenbogen RG, Britz GW, Schuster J, Lucas T, Newell DW, Mansfield PN, Machamer JE, Barber J, Dikmen SS. Magnesium sulfate for neuroprotection after traumatic brain injury: a randomised controlled trial. *Lancet Neurol* 6: 29–38, 2007.
512. Teragawa H, Kato M, Yamagata T, Matsuura H, Kajiyama G. Magnesium causes nitric oxide independent coronary artery vasodilation in humans. *Heart* 86: 212–216, 2001.
513. Teragawa H, Matsuura H, Chayama K, Oshima T. Mechanisms responsible for vasodilation upon magnesium infusion in vivo: clinical evidence. *Magnesium Res* 15: 241–246, 2002.
514. Terasaki M, Rubin H. Evidence that intracellular magnesium is present in cells at a regulatory concentration for protein synthesis. *Proc Natl Acad Sci USA* 82: 7324–7326, 1985.
515. Thebault S, Alexander RT, Tiel Groenestege WM, Hoenderop JG, Bindels RJ. EGF increases TRPM6 activity and surface expression. *J Am Soc Nephrol* 20: 78–85, 2009.
516. Thebault S, Cao G, Venselaar H, Xi Q, Bindels RJ, Hoenderop JG. Role of the alpha-kinase domain in transient receptor potential melastatin 6 channel and regulation by intracellular ATP. *J Biol Chem* 283: 19999–20007, 2008.
517. Thompson CB, June CH, Sullivan KM, Thomas ED. Association between cyclosporin neurotoxicity and hypomagnesaemia. *Lancet* 2: 1116–1120, 1984.
518. Thony B, Neuheiser F, Kierat L, Blaskovics M, Arn PH, Ferreira P, Rebrin I, Ayling J, Blau N. Hyperphenylalaninemia with high levels of 7-biopterin is associated with mutations in the PCBD gene encoding the bifunctional protein pterin-4a-carbinolamine dehydratase and transcriptional coactivator (DCoH). *Am J Hum Genet* 62: 1302–1311, 1998.
519. Thony B, Neuheiser F, Kierat L, Rolland MO, Guibaud P, Schluter T, Germann R, Heidenreich RA, Duran M, de Klerk JB, Ayling JE, Blau N. Mutations in the pterin-4a-carbinolamine dehydratase (PCBD) gene cause a benign form of hyperphenylalaninemia. *Hum Genet* 103: 162–167, 1998.
520. Tietz NW. *Clinical Guide to Laboratory Tests*. Philadelphia, PA: Saunders, 1995.
521. Tofil NM, Benner KW, Winkler MK. Fatal hypermagnesemia caused by an Epsom salt enema: a case illustration. *Southern Medical J* 98: 253–256, 2005.
522. Tonelli M, Wiebe N, Culletto B, House A, Rabbat C, Fok M, McAlister F, Garg AX. Chronic kidney disease and mortality risk: a systematic review. *J Am Soc Nephrol* 17: 2034–2047, 2006.
523. Topf JM, Murray PT. Hypomagnesemia and hypermagnesemia. *Rev Endocr Metab Disorders* 4: 195–206, 2003.
524. Topol EJ, Lerman BB. Hypomagnesemic torsades de pointes. *Am J Cardiol* 52: 1367–1368, 1983.
525. Tsang HT, Edwards TL, Wang X, Connell JW, Davies RJ, Durrington HJ, O'Kane CJ, Luzio JP, Reid E. The hereditary spastic paraplegia proteins NIPA1, spastin and spartin are inhibitors of mammalian BMP signalling. *Hum Mol Genet* 18: 3805–3821, 2009.

526. Turecky L, Kupcova V, Szantova M, Uhlíkova E, Viktorinova A, Czifrusz A. Serum magnesium levels in patients with alcoholic and non-alcoholic fatty liver. *Bratislavske Lekarske Listy* 107: 58–61, 2006.
527. Tzivoni D, Banai S, Schuger C, Benhorin J, Keren A, Gottlieb S, Stern S. Treatment of torsades de pointes with magnesium sulfate. *Circulation* 77: 392–397, 1988.
528. Tzivoni D, Keren A, Cohen AM, Loebel H, Zahavi I, Chenzbraun A, Stern S. Magnesium therapy for torsades de pointes. *Am J Cardiol* 53: 528–530, 1984.
529. Van Angelen AA, Glaudemans B, van der Kemp AW, Hoenderop JG, Bindels RJ. Cisplatin-induced injury of the renal distal convoluted tubule is associated with hypomagnesaemia in mice. *Nephrol Dialysis Transplantation* 28: 879–889, 2013.
530. Van Angelen AA, van der Kemp AW, Hoenderop JG, Bindels RJ. Increased expression of renal TRPM6 compensates for Mg²⁺ wasting during furosemide treatment. *Clin Kidney J* 5: 535–544, 2012.
531. Van den Bergh WM, Algra A, Rinkel GJ. Electrocardiographic abnormalities and serum magnesium in patients with subarachnoid hemorrhage. *Stroke* 35: 644–648, 2004.
532. Van den Bergh WM, Algra A, van der Sprenkel JW, Tulleken CA, Rinkel GJ. Hypomagnesaemia after aneurysmal subarachnoid hemorrhage. *Neurosurgery* 52: 276–281, 2003.
533. Van den Bergh WM, Algra A, van Kooten F, Dirven CM, van Gijn J, Vermeulen M, Rinkel GJ. Magnesium sulfate in aneurysmal subarachnoid hemorrhage: a randomized controlled trial. *Stroke* 36: 1011–1015, 2005.
534. Van den Brandt PA, Smits KM, Goldbohm RA, Weijnenberg MP. Magnesium intake and colorectal cancer risk in the Netherlands Cohort Study. *Br J Cancer* 96: 510–513, 2007.
535. Van Laecke S, Van Biesen W, Verbeke F, De Bacquer D, Peeters P, Vanholder R. Posttransplantation hypomagnesaemia and its relation with immunosuppression as predictors of new-onset diabetes after transplantation. *Am J Transplant* 9: 2140–2149, 2009.
536. Vargas-Caballero M, Robinson HP. Fast and slow voltage-dependent dynamics of magnesium block in the NMDA receptor: the asymmetric trapping block model. *J Neurosci* 24: 6171–6180, 2004.
537. Vidair C, Rubin H. Evaluation of Mg²⁺ as an intracellular regulator of uridine uptake. *J Cell Physiol* 108: 317–325, 1981.
538. Vink R, Nechifor M. *Magnesium in the Central Nervous System*. Adelaide, Australia: Univ. of Adelaide Press, 2011.
539. Vink R, van den Heuvel C. Substance P antagonists as a therapeutic approach to improving outcome following traumatic brain injury. *Neurotherapeutics* 7: 74–80, 2010.
540. Voets T, Nilius B, Hoefs S, van der Kemp AW, Droogmans G, Bindels RJ, Hoenderop JG. TRPM6 forms the Mg²⁺ influx channel involved in intestinal and renal Mg²⁺ absorption. *J Biol Chem* 279: 19–25, 2004.
541. Volpe SL. Magnesium, the metabolic syndrome, insulin resistance, and type 2 diabetes mellitus. *Crit Rev Food Sci Nutr* 48: 293–300, 2008.
542. Vosgerau H. [Migraine therapy with magnesium glutamate]. *Therapie der Gegenwart* 112: 640, 1973.
543. Walder RY, Landau D, Meyer P, Shalev H, Tsolia M, Borochoowitz Z, Boettger MB, Beck GE, Englehardt RK, Carmi R, Sheffield VC. Mutation of TRPM6 causes familial hypomagnesaemia with secondary hypocalcaemia. *Nature Genet* 31: 171–174, 2002.
544. Walder RY, Shalev H, Brennan TM, Carmi R, Elbedour K, Scott DA, Hanauer A, Mark AL, Patil S, Stone EM, Sheffield VC. Familial hypomagnesaemia maps to chromosome 9q, not to the X chromosome: genetic linkage mapping and analysis of a balanced translocation breakpoint. *Hum Mol Genet* 6: 1491–1497, 1997.
545. Wang CY, Shi JD, Yang P, Kumar PG, Li QZ, Run QG, Su YC, Scott HS, Kao KJ, She JX. Molecular cloning and characterization of a novel gene family of four ancient conserved domain proteins (ACDP). *Gene* 306: 37–44, 2003.
546. Wang JJ, Freeman AI, Gaeta JF, Sinks LF. Unusual complications of pentamidine in the treatment of *Pneumocystis carinii* pneumonia. *J Pediatr* 77: 311–314, 1970.
547. Wang M, Tashiro M, Berlin JR. Regulation of L-type calcium current by intracellular magnesium in rat cardiac myocytes. *J Physiol* 555: 383–396, 2004.
548. Wang X, Proud CG. Nutrient control of TORC1, a cell-cycle regulator. *Trends Cell Biol* 19: 260–267, 2009.
549. Watson JD, Crick FH. Molecular structure of nucleic acids: a structure for deoxyribose nucleic acid. *Nature* 171: 737–738, 1953.
550. Weber G, Lea MA, Convery HJ, Stamm NB. Regulation of gluconeogenesis and glycolysis: studies of mechanisms controlling enzyme activity. *Adv Enzyme Regul* 5: 257–300, 1967.
551. Weber JD, Gutmann DH. Deconvoluting mTOR biology. *Cell Cycle* 11: 236–248, 2012.
552. Weglicki WB, Dickens BF, Wagner TL, Chmielinska JJ, Phillips TM. Immunoregulation by neuropeptides in magnesium deficiency: ex vivo effect of enhanced substance P production on circulating T lymphocytes from magnesium-deficient mice. *Magnesium Res* 9: 3–11, 1996.
553. Weglicki WB, Mak IT, Phillips TM. Blockade of cardiac inflammation in Mg²⁺ deficiency by substance P receptor inhibition. *Circ Res* 74: 1009–1013, 1994.
554. Weglicki WB, Phillips TM. Pathobiology of magnesium deficiency: a cytokine/neurogenic inflammation hypothesis. *Am J Physiol Regul Integr Comp Physiol* 263: R734–R737, 1992.
555. Westermaier T, Stetter C, Vince GH, Pham M, Tejon JP, Eriskat J, Kunze E, Matthies C, Ernestus RI, Solymosi L, Roosen K. Prophylactic intravenous magnesium sulfate for treatment of aneurysmal subarachnoid hemorrhage: a randomized, placebo-controlled, clinical study. *Crit Care Med* 38: 1284–1290, 2010.
556. Westhof E, Sundaralingam M. Restrained refinement of the monoclinic form of yeast phenylalanine transfer RNA. Temperature factors and dynamics, coordinated waters, and base-pair propeller twist angles. *Biochemistry* 25: 4868–4878, 1986.
557. Weston PG. Magnesium as a sedative. *Am J Psychiatry* 78: 637–638, 1922.
558. White RE, Hartzell HC. Effects of intracellular free magnesium on calcium current in isolated cardiac myocytes. *Science* 239: 778–780, 1988.
559. Widman L, Wester PO, Stegmayr BK, Wirell M. The dose-dependent reduction in blood pressure through administration of magnesium. A double blind placebo controlled cross-over study. *Am J Hypertens* 6: 41–45, 1993.
560. Wilcox ER, Burton QL, Naz S, Riazuddin S, Smith TN, Ploplis B, Belyantseva I, Ben-Yosef T, Liburd NA, Morell RJ, Kachar B, Wu DK, Griffith AJ, Riazuddin S, Friedman TB. Mutations in the gene encoding tight junction claudin-14 cause autosomal recessive deafness DFNB29. *Cell* 104: 165–172, 2001.
561. Wiles ME, Wagner TL, Weglicki WB. Effect of acute magnesium deficiency (MgD) on aortic endothelial cell (EC) oxidant production. *Life Sci* 60: 221–236, 1997.
562. Wilkinson R, Lucas GL, Heath DA, Franklin IM, Boughton BJ. Hypomagnesaemic tetany associated with prolonged treatment with aminoglycosides. *Br Med J* 292: 818–819, 1986.
563. Williams BA, Beatch GN. Magnesium shifts voltage dependence of activation of delayed rectifier I(K) in guinea pig ventricular myocytes. *Am J Physiol Heart Circ Physiol* 272: H1292–H1301, 1997.
564. Wisdom DM, Salido GM, Baldwin LM, Singh J. The role of magnesium in regulating CCK-8-evoked secretory responses in the exocrine rat pancreas. *Mol Cell Biochem* 154: 123–132, 1996.
565. Witkowski M, Hubert J, Mazur A. Methods of assessment of magnesium status in humans: a systematic review. *Magnesium Res* 24: 163–180, 2011.
566. Wolf FI, Fasanella S, Tedesco B, Torsello A, Sgambato A, Faraglia B, Palozza P, Boninsegna A, Cittadini A. Regulation of magnesium content during proliferation of mammary epithelial cells (HC-11). *Front Biosci* 9: 2056–2062, 2004.
567. Wolf MT, Dotsch J, Konrad M, Boswald M, Rascher W. Follow-up of five patients with FHHNC due to mutations in the Paracellin-1 gene. *Pediatr Nephrol* 17: 602–608, 2002.
568. Wong GK, Poon WS, Chan MT, Boet R, Gin T, Ng SC, Zee BC. Intravenous magnesium sulphate for aneurysmal subarachnoid hemorrhage (IMASH): a randomized, double-blinded, placebo-controlled, multicenter phase III trial. *Stroke* 41: 921–926, 2010.

569. Woods KL, Fletcher S, Roffe C, Haider Y. Intravenous magnesium sulphate in suspected acute myocardial infarction: results of the second Leicester Intravenous Magnesium Intervention Trial (LIMIT-2). *Lancet* 339: 1553–1558, 1992.
570. Worthington V. Nutritional quality of organic versus conventional fruits, vegetables, and grains. *J Altern Complement Med* 7: 161–173, 2001.
571. Wright FS. Increasing magnitude of electrical potential along the renal distal tubule. *Am J Physiol* 220: 624–638, 1971.
572. Xu JZ, Hall AE, Peterson LN, Bienkowski MJ, Eessalu TE, Hebert SC. Localization of the ROMK protein on apical membranes of rat kidney nephron segments. *Am J Physiol Renal Physiol* 273: F739–F748, 1997.
573. Yago MD, Manas M, Singh J. Intracellular magnesium: transport and regulation in epithelial secretory cells. *Front Biosci* 5: D602–618, 2000.
574. Yamamoto M, Yamaguchi T, Yamauchi M, Yano S, Sugimoto T. Acute-onset hypomagnesemia-induced hypocalcemia caused by the refractoriness of bones and renal tubules to parathyroid hormone. *J Bone Miner Metab* 29: 752–755, 2011.
575. Yamazaki D, Funato Y, Miura J, Sato S, Toyosawa S, Furutani K, Kurachi Y, Omori Y, Furukawa T, Tsuda T, Kuwabata S, Mizukami S, Kikuchi K, Miki H. Basolateral Mg^{2+} extrusion via CNNM4 mediates transcellular Mg^{2+} transport across epithelia: a mouse model. *PLoS Genet* 9: e1003983, 2013.
576. Yan Y, Tian J, Mo X, Zhao G, Yin X, Pu J, Zhang B. Genetic variants in the RAB7L1 and SLC41A1 genes of the PARK16 locus in Chinese Parkinson's disease patients. *Int J Neurosci* 121: 632–636, 2011.
577. Yang L, Arora K, Beard WA, Wilson SH, Schlick T. Critical role of magnesium ions in DNA polymerase beta's closing and active site assembly. *J Am Chem Soc* 126: 8441–8453, 2004.
578. Yang L, Frindt G, Palmer LG. Magnesium modulates ROMK channel-mediated potassium secretion. *J Am Soc Nephrol* 21: 2109–2116, 2010.
579. Yang Y, Li Q, Ahmad F, Shuaib A. Survival and histological evaluation of therapeutic window of post-ischemia treatment with magnesium sulfate in embolic stroke model of rat. *Neurosci Lett* 285: 119–122, 2000.
580. Yasui M, Kihira T, Ota K. Calcium, magnesium and aluminum concentrations in Parkinson's disease. *Neurotoxicology* 13: 593–600, 1992.
581. Yee NS, Zhou W, Liang IC. Transient receptor potential ion channel Trpm7 regulates exocrine pancreatic epithelial proliferation by Mg^{2+} -sensitive Socs3a signaling in development and cancer. *Disease Models Mechanisms* 4: 240–254, 2011.
582. Young FB, Franciosi S, Spreeuw A, Deng Y, Sanders S, Tam NC, Huang K, Singaraja RR, Zhang W, Bissada N, Kay C, Hayden MR. Low levels of human HIP14 are sufficient to rescue neuropathological, behavioural, and enzymatic defects due to loss of murine HIP14 in Hip14^{-/-} mice. *PLoS One* 7: e36315, 2012.
583. Young GL, Jewell D. Interventions for leg cramps in pregnancy. *Cochrane Database Systematic Rev* CD000121, 2002.
584. Zafar S, Hussain A, Liu Y, Lewis D, Inesi G. Specificity of ligand binding to transport sites: Ca^{2+} binding to the Ca^{2+} transport ATPase and its dependence on H^{+} and Mg^{2+} . *Arch Biochem Biophys* 476: 87–94, 2008.
585. Zaloga GP, Chernow B, Pock A, Wood B, Zaritsky A, Zucker A. Hypomagnesemia is a common complication of aminoglycoside therapy. *Surg Gynecol Obstet* 158: 561–565, 1984.
586. Zaman F, Abreo K. Severe hypermagnesemia as a result of laxative use in renal insufficiency. *Southern Med J* 96: 102–103, 2003.
587. Zhang W, Iso H, Ohira T, Date C, Tamakoshi A. Associations of dietary magnesium intake with mortality from cardiovascular disease: the JACC study. *Atherosclerosis* 221: 587–595, 2012.
588. Zhang Z, Yu H, Huang J, Faouzi M, Schmitz C, Penner R, Fleig A. The TRPM6 kinase domain determines the MgATP sensitivity of TRPM7/M6 heteromeric ion channels. *J Biol Chem* 289: 5217–5227, 2014.
589. Zhou H, Clapham DE. Mammalian MagT1 and TUSC3 are required for cellular magnesium uptake and vertebrate embryonic development. *Proc Natl Acad Sci USA* 106: 15750–15755, 2009.
590. Zhou Q, Olinescu RM, Kummerow FA. Influence of low magnesium concentrations in the medium on the antioxidant system in cultured human arterial endothelial cells. *Magnesium Res* 12: 19–29, 1999.
591. Zipes DP, Camm AJ, Borggrefe M, Buxton AE, Chaitman B, Fromer M, Gregoratos G, Klein G, Moss AJ, Myerburg RJ, Priori SG, Quinones MA, Roden DM, Silka MJ, Tracy C, Smith SC Jr, Jacobs AK, Adams CD, Antman EM, Anderson JL, Hunt SA, Halperin JL, Nishimura R, Ornato JP, Page RL, Riegel B, Blanc JJ, Budaj A, Dean V, Deckers JW, Despres C, Dickstein K, Lekakis J, McGregor K, Metra M, Morais J, Osterspey A, Tamargo JL, Zamorano JL. ACC/AHA/ESC 2006 Guidelines for Management of Patients With Ventricular Arrhythmias and the Prevention of Sudden Cardiac Death: a report of the American College of Cardiology/American Heart Association Task Force and the European Society of Cardiology Committee for Practice Guidelines (writing committee to develop Guidelines for Management of Patients With Ventricular Arrhythmias and the Prevention of Sudden Cardiac Death): developed in collaboration with the European Heart Rhythm Association and the Heart Rhythm Society. *Circulation* 114: e385–484, 2006.
592. Zsurka G, Gregan J, Schweyen RJ. The human mitochondrial Mrs2 protein functionally substitutes for its yeast homologue, a candidate magnesium transporter. *Genomics* 72: 158–168, 2001.
593. Zumkley H, Bertram HP, Preusser P, Kellinghaus H, Straub C, Vetter H. Renal excretion of magnesium and trace elements during cisplatin treatment. *Clin Nephrol* 17: 254–257, 1982.
594. Zuspan FP. Treatment of severe preeclampsia and eclampsia. *Clin Obstet Gynecol* 9: 954–972, 1966.
595. Zwillinger L. über die Magnesiumwirkung auf das Herz. *Klin Wochenschr* 14: 1429–1433, 1935.