MAGNESIUM IN MAN: IMPLICATIONS FOR HEALTH AND DISEASE

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De Baaij JHF, Hoenderop JGJ, Bindels RJM. Magnesium in Man: Implications for Health and Disease. Physiol Rev 95: 1–46, 2015; doi:10.1152/physrev.00012.2014.—Magnesium ([Mg\(^{2+}\)]) is an essential ion to the human body, playing an instrumental role in supporting and sustaining health and life. As the second most abundant intracellular cation after potassium, it is involved in over 600 enzymatic reactions including energy metabolism and protein synthesis. Although Mg\(^{2+}\) availability has been proven to be disturbed during several clinical situations, serum Mg\(^{2+}\) values are not generally determined in patients. This review aims to provide an overview of the function of Mg\(^{2+}\) in human health and disease. In short, Mg\(^{2+}\) plays an important physiological role particularly in the brain, heart, and skeletal muscles. Moreover, Mg\(^{2+}\) supplementation has been shown to be beneficial in treatment of, among others, preeclampsia, migraine, depression, coronary artery disease, and asthma. Over the last decade, several hereditary forms of hypomagnesemia have been deciphered, including mutations in transient receptor potential melastatin type 6 (TRPM6), claudin 16, and cyclin M2 (CNNM2). Recently, mutations in Mg\(^{2+}\) transporter 1 (MagT1) were linked to T-cell deficiency underlining the important role of Mg\(^{2+}\) in cell viability. Moreover, hypomagnesemia can be the consequence of the use of certain types of drugs, such as diuretics, epidermal growth factor receptor inhibitors, calcineurin inhibitors, and proton pump inhibitors. This review provides an extensive and comprehensive overview of Mg\(^{2+}\) research over the last few decades, focusing on the regulation of Mg\(^{2+}\) homeostasis in the intestine, kidney, and bone and disturbances which may result in hypomagnesemia.

I. INTRODUCTION

Magnesium (Mg\(^{2+}\)) is an essential ion for health. Mg\(^{2+}\) plays an important role in the physiological function of the brain, heart, and skeletal muscles. Mg\(^{2+}\) has anti-inflammatory properties and acts as Ca\(^{2+}\) antagonist. The United States Food and Nutrition Board recommends a daily intake of 420 mg for men and 320 mg for women (1). However, recent reports estimate that at least 60% of Americans do not consume the recommended daily amount of Mg\(^{2+}\) (281). Part of the problem stems from the soil used for agriculture, which is becoming increasingly deficient in essential minerals. Over the last 60 years the Mg\(^{2+}\) content in fruit and vegetables decreased by 20–30% (570). Moreover, the Western diet contains more refined grains and processed food. Estimates are that 80–90% of Mg\(^{2+}\) is lost during food processing. As a result, a significant number of people are Mg\(^{2+}\) deficient, which may comprise up to 60% of critically ill patients (84, 145). Mg\(^{2+}\) deficiency is commonly determined by measuring total serum Mg\(^{2+}\) concentrations, which ranges between 0.7 and 1.05 mM in a healthy person (323). However, serum Mg\(^{2+}\) values reflect only 1% of the body Mg\(^{2+}\) content, since most of the body’s Mg\(^{2+}\) is stored in bone, muscle, and soft tissues. Therefore, although serum values are within the normal range, the body can be in a severely Mg\(^{2+}\)-depleted state. Consequently, the clinical impact of Mg\(^{2+}\) deficiency may be largely underestimated.

The first use of Mg\(^{2+}\) in human medicine can be traced back to 1697 when Dr. Nehemiah Grew identified magnesium sulfate (MgSO\(_4\)) as the major ingredient of Epsom salt (195). Epsom salt was extracted from a well in Epsom, England and was used over the years to treat abdominal pain, constipation, sprains, muscle strains, hyaline membrane disease, and cerebral edema. Subsequently, Mg\(^{2+}\) was recognized as an element (Mg) by Joseph Black in 1755 and first isolated by Sir Humphrey Davy from magnesia [Mg\(_3\)SO\(_4\)O\(_{10}\)(OH)\(_2\)] and mercury in 1808 (102). The role of Mg\(^{2+}\) in the human body emerged once Mg\(^{2+}\) was described in blood plasma by Willey Glover Denis in 1920 (113). In 1926, Jehan Leroy demonstrated that Mg\(^{2+}\) is essential for life in mice (309). These findings were trans-
lated to humans, and the first report of Mg$^{2+}$ deficiency in humans was by Arthur Hirschfelder and Victor Haury in 1934 (231). Since then, Mg$^{2+}$ has been implicated in and used for treatment of a variety of diseases, including migraines, cardiovascular diseases, and diabetes. Although the importance of Mg$^{2+}$ is widely acknowledged, serum Mg$^{2+}$ values are not generally determined in clinical medicine. Therefore, Mg$^{2+}$ is often referred to as the “forgotten” cation in human health.

This review provides an overview of the role of Mg$^{2+}$ in human health and disease. Mg$^{2+}$ has important cellular functions in enzymatic reactions and in the synthesis and structure of proteins and polynucleotides, which are described in section II. The important regulation of Mg$^{2+}$ homeostasis is discussed in depth in section III. The role of Mg$^{2+}$ in organ function and related diseases is discussed in section IV. An overview is presented of the most important diseases in which Mg$^{2+}$ disturbances have been implicated or in which Mg$^{2+}$ has been considered as a potential treatment. In last part of the review, special attention is awarded to disturbances of intestinal Mg$^{2+}$ uptake and renal Mg$^{2+}$ excretion (sect. V). All together, this review emphasizes the importance of a controlled Mg$^{2+}$ balance in the human body. Increasing the awareness and understanding of Mg$^{2+}$ homeostasis may give more clinical attention to the important role of Mg$^{2+}$ in health and disease.

II. MAGNESIUM IN CELLULAR PHYSIOLOGY

Within the periodic table of elements, Mg has the atomic number 12 and is classed as an alkaline earth element (group 2). Mg occurs in three stable isotopes: $^{24}$Mg, $^{25}$Mg, and $^{26}$Mg. $^{24}$Mg is the most common isotope (78.99%) and has a relative atomic mass of 24.305 Da, a melting point of 648.8°C, and a boiling point of 1,090°C (350). Mg$^{2+}$ is highly soluble and the second most abundant cation in seawater (95). In the dissolved state, Mg$^{2+}$ has two hydration shells, making its hydrated radius ~400 times larger than its dehydrated radius, larger than that of other cations like Na$^+$, K$^+$, and even Ca$^{2+}$ (95). Consequently, Mg$^{2+}$ needs to be dehydrated before passing through channels and transporters, a process that requires a lot of energy. Mg$^{2+}$ is a powerful Ca$^{2+}$ antagonist, despite both having similar charge and chemical properties.

Mg$^{2+}$ is the second most abundant intracellular cation with typical concentrations of ~10–30 mM. However, since most of the intracellular Mg$^{2+}$ is bound to ribosomes, polynucleotides and ATP, the concentration of freely available Mg$^{2+}$ falls within the low millimolar range (0.5–1.2 mM) (133). In contrast to other abundant ions, for which cells maintain considerable transmembrane gradients, the free Mg$^{2+}$ concentrations in the cell and in the extracellular fluid are comparable. Mg$^{2+}$ is a versatile ion that is involved in practically every major metabolic and biochemical process within the cell. Although it extends beyond the purpose of this review to give a comprehensive overview of all biochemical reactions and structural processes involving Mg$^{2+}$, the following paragraphs will highlight the most prominent cellular processes in which Mg$^{2+}$ is involved.

A. Nucleotide Binding

Mg$^{2+}$ forms an essential component of the RNA and DNA tertiary structures, as it binds the negatively charged O and N molecules within the polynucleotide chains. Polynucleotide binding is a complex biophysical process that mainly depends on the level of Mg$^{2+}$ dehydration and the electrostatic potential at the binding site (for extensive review, see Ref. 347). The most studied Mg$^{2+}$-RNA interaction is tRNA, where Mg$^{2+}$ stabilizes the structure. The role of Mg$^{2+}$ became evident in 1966, when it was shown that Mg$^{2+}$ could restore denaturated tRNA molecules (319). Crystallographic structures of tRNAs from yeast identified five Mg$^{2+}$-binding sites, three in the core region around the bend of the L-shaped molecule and two additional sites in the major groove of the anticodon stem (412, 556). Additionally, there may be a few dozen Mg$^{2+}$ in close vicinity of tRNA molecules that may bind weakly to the exterior of the structure (347). Still, the importance of Mg$^{2+}$ binding for the tRNA tertiary structure has been contested over the years. This discussion was mainly triggered by studies showing the importance of nonspecific diffuse binding of Mg$^{2+}$ and other divalent and monovalent cations, questioning the specificity of the Mg$^{2+}$ interactions. However, the role of Mg$^{2+}$ in RNA structure extends beyond tRNAs. For instance, Mg$^{2+}$ is also crucial to the interactions that stabilize the pseudoknot conformation (191), tertiary RNA structures that are present in mRNA, ribosomal RNA, transfer-messenger RNA, catalytic self-splicing RNA, and viral genomic RNA.

In DNA, Mg$^{2+}$ forms hydrogen bonds with the electronegative elements (O, N) to stabilize the natural DNA conformation, referred to as B-DNA (85, 549). Moreover, Mg$^{2+}$ plays a role in the secondary and tertiary structure of DNA by competing with monovalent ions (394). Mg$^{2+}$ binds the minor groove of B-DNA structures, thus protecting it. In Mg$^{2+}$-deficient conditions, DNA is more accessible to free oxygen radicals and more prone to oxidative stress (406). However, at higher Mg$^{2+}$ concentrations, Mg$^{2+}$ may covalently bind DNA, locally distorting the double helix (22). Therefore, maintaining the cellular Mg$^{2+}$ concentration within the physiological range is essential for DNA stability.

B. Enzymatic Activity

In medical textbooks and scientific literature, Mg$^{2+}$ is often described as a cofactor for ~300 enzymes. Theodor
Günther introduced the number 300 as a rough estimate in 1980 and this has been in use ever since (133). However, in the decades after 1980 many new Mg\(^{2+}\)-dependent enzymes have been described, and the number 300 is, therefore, an underestimation. Currently, enzymatic databases list over 600 enzymes for which Mg\(^{2+}\) serves as cofactor, and an additional 200 in which Mg\(^{2+}\) may act as activator (32, 73). An overview of these Mg\(^{2+}\)-dependent enzymes can be found at MetaCyc (http://www.metacyc.org; Ref. 73). Many of the enzymes that require Mg\(^{2+}\) as coactivator are vital for life.

Mg\(^{2+}\) is necessary for the proper structure and activity of DNA and RNA polymerases (56, 500). DNA polymerases have two Mg\(^{2+}\) binding sites, which are hypothesized to play a key role in the conformational changes in the polymerase enzyme during the catalytic reaction (56). This model was further enhanced by studies reporting that the release of one of the Mg\(^{2+}\) ions is necessary for opening the catalytic site for new nucleotides (577). In addition, Mg\(^{2+}\) acts as cofactor for almost every enzyme involved in basically every step of NER (68). In BER, Mg\(^{2+}\) is elemental for the activity of endonuclease, which incise the DNA after DNA damage, and the DNA polymerases and ligases, which repair the gap (41, 473). The third repair pathway, MMR, is also affected by Mg\(^{2+}\) availability since several enzymes involved require Mg\(^{2+}\) and ATP for activity (33). Other enzymes requiring Mg\(^{2+}\) are topoisomerases, helicases, exonucleases, protein kinases, cyclases, and large groups of ATPases, meaning that Mg\(^{2+}\) is an essential component of DNA replication, RNA transcription, amino acid synthesis, and protein formation. Altogether, Mg\(^{2+}\) is a key factor in the maintenance of genomic and genetic stability. The consequences of low Mg\(^{2+}\) availability on the development of cancer is discussed in section IIIE1.

Mg\(^{2+}\) is also an important regulator of many enzymes involved in glycolysis, because it is a cofactor for adenine nucleotides. Mg-ATP is required for the activity of hexokinase, phosphofructokinase, aldolase, phosphoglycerate kinase, and pyruvate kinase (166, 550). Consequently, Mg\(^{2+}\) availability is of major importance for glucose metabolism, which may explain its role in diabetes mellitus type 2 (see sect. IV.E1).

The role of Mg\(^{2+}\), however, extends far beyond DNA and protein synthesis, DNA repair, and glycolysis. Since kinases, ATPases, guanylyl cyclases, and adenyllyl cyclases all depend on Mg-ATP for proper function, Mg\(^{2+}\) plays a role in virtually every process in the cell.

### C. Cellular Mg\(^{2+}\) Handling

Since protein and DNA synthesis are highly dependent on intracellular Mg\(^{2+}\) availability, intracellular Mg\(^{2+}\) concentrations are tightly regulated. Over the last 20 years, elucidating the molecular identities of the transporters involved in Mg\(^{2+}\) homeostasis has been the main focus of research within the Mg\(^{2+}\) field. Genetic screenings on human diseases and microarray-based expression studies have resulted in the identification of numerous Mg\(^{2+}\)-transporting proteins (Table 1 and Figure 1). Although the exact role of many of these proteins needs further investigation, re-

#### Table 1. Mg\(^{2+}\) Transporters

<table>
<thead>
<tr>
<th>Name</th>
<th>Membrane</th>
<th>Expression</th>
<th>Permeability</th>
<th>Mechanism</th>
<th>Disease</th>
<th>Reference Nos.</th>
</tr>
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<tbody>
<tr>
<td><strong>General Mg(^{2+}) transporters</strong></td>
<td></td>
<td></td>
<td></td>
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<td></td>
<td></td>
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<tr>
<td>TRPM7</td>
<td>Plasma membrane</td>
<td>Ubiquitous</td>
<td>Ba-&gt;Ni&gt;Mg&gt;Ca</td>
<td>Channel</td>
<td></td>
<td>314, 349</td>
</tr>
<tr>
<td>MagT1</td>
<td>Plasma membrane</td>
<td>Ubiquitous</td>
<td>Mg&gt;Ba&gt;Fe&gt;Cu</td>
<td>Channel</td>
<td>X-MEN syndrome</td>
<td>187, 311</td>
</tr>
<tr>
<td>SLC41A1</td>
<td>Plasma membrane</td>
<td>Ubiquitous</td>
<td>Mg&gt;Sr&gt;Fe&gt;Ba&gt;Cu</td>
<td>Exchanger</td>
<td>Nephronphthisis-like</td>
<td>185, 251, 289</td>
</tr>
<tr>
<td>SLC41A2</td>
<td>Golgi membrane</td>
<td>Ubiquitous</td>
<td>Mg&gt;Ca&gt;Ba&gt;Ni</td>
<td>Exchanger</td>
<td></td>
<td>442</td>
</tr>
<tr>
<td>CNNM3</td>
<td>Plasma membrane</td>
<td>Ubiquitous</td>
<td>Mg&gt;Cu&gt;Ba&gt;Co</td>
<td>Transporter?</td>
<td></td>
<td>545</td>
</tr>
<tr>
<td>MRS2</td>
<td>Mitochondrial membrane</td>
<td>Ubiquitous</td>
<td>Mg&gt;Ni</td>
<td>Channel</td>
<td></td>
<td>399</td>
</tr>
<tr>
<td><strong>Tissue-specific Mg(^{2+}) transporters</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
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<td></td>
</tr>
<tr>
<td>TRPM6</td>
<td>Apical plasma membrane</td>
<td>Kidney, intestine</td>
<td>Ba-&gt;Ni&gt;Mg&gt;Ca</td>
<td>Channel</td>
<td>Hypomagnesemia secondary</td>
<td>314</td>
</tr>
<tr>
<td>CNNM1</td>
<td>?</td>
<td>Brain</td>
<td>Cu&gt;Mg?</td>
<td>?</td>
<td>hypocalcemia</td>
<td>13, 545</td>
</tr>
<tr>
<td>CNNM2</td>
<td>Basolateral plasma membrane</td>
<td>Kidney</td>
<td>Mg&gt;Sr&gt;Zn&gt;Cd</td>
<td>Transporter?</td>
<td>Sensor?</td>
<td>184, 497</td>
</tr>
<tr>
<td>CNNM4</td>
<td>Basolateral plasma membrane</td>
<td>Intestine</td>
<td>Mg</td>
<td>Exchanger?</td>
<td>Jallili syndrome</td>
<td>387, 402, 575</td>
</tr>
</tbody>
</table>

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searchers have identified several proteins critical to cellular Mg$^{2+}$ homeostasis. Within this part of the review, we will focus on the ubiquitous transporters transient receptor potential melastatin type 7 (TRPM7), Mg$^{2+}$ transporter 1 (MagT1), and solute carrier family 41 member 1 (SLC41A1). Tissue-specific Mg$^{2+}$ transporters such as transient receptor potential melastatin type 6 (TRPM6; kidney, colon), cyclin M2 (CNNM2; kidney) and cyclin M4 (CNNM4; colon) are discussed in section III.

1. **TRPM7**

TRPM7 is a ubiquitously expressed divalent cation channel that is responsible for much of the Mg$^{2+}$ flux in the cell. TRPM7 activity is generally regarded as a prerequisite for cell viability (360, 455). However, recent reports with tissue-specific TRPM7 KO mice suggest that TRPM7-deficient T cells are still viable and have normal intracellular Mg$^{2+}$ concentrations (266). TRPM7 constitutes a tetrameric channel, where each subunit consists of six transmembrane regions with a pore region between the fifth and sixth transmembrane domain (42). The intracellular COOH terminus contains a kinase domain that regulates autophosphorylation of the channel, although its mechanism is poorly understood, as TRPM7 channel function is not dependent on its kinase activity (504). Initially, the kinase was reported to exist as separate entity (438), and indeed, recently it was shown that the kinase is cleaved from TRPM7 by caspase-8, although the exact function of the cleaved kinase remains unknown (115).

2. **MagT1**

Originally identified in MDCT cells, Mg$^{2+}$ transporter 1 (MagT1) has been described as a ubiquitously expressed Mg$^{2+}$ channel (187). Survival and growth of TRPM7-defi-
cient cells can be partially rescued by MagT1 overexpression (110). Although identification of MagT1 dates back almost 10 years, the functional characteristics of MagT1 are still undetermined. MagT1 mediates highly specific Mg^{2+} currents in Xenopus laevis oocytes, but these results could not be reproduced in mammalian cells (187, 589). Recent studies in T cells suggest that MagT1 mediates a rapid Mg^{2+} influx upon receptor activation (79, 311). Since T cells do not require TRPM7 for maintaining normal intracellular Mg^{2+} concentrations (266), this suggests that MagT1 has a similar function as TRPM7 in certain cell types.

3. Extrusion

In 1984, Theodor Gunther et al. (214) proposed that the main route of Mg^{2+} efflux from the cell is Na^{+} dependent. A large body of evidence obtained in a wide range of cell types supports this notion (reviewed in Ref. 424). Over the last decades, the mechanism has been further characterized in a variety of cell types, demonstrating inhibition by Na^{+} channel blockers such as amiloride, imipramine, and quinidine. The stoichiometry of this exchange mechanism is still not fully elucidated; Na^{+}-dependent Mg^{2+} extrusion is activated by cAMP in several cell models and conditions (424), but Na^{+}-independent Mg^{2+} extrusion has also been proposed. Ebel et al. (134) reported the presence of a choline-dependent Mg^{2+} transporter in erythrocytes. However, the molecular identity of this proposed Mg^{2+} efflux mechanism remains controversial.

4. SLC41A1

Recent reports by Kolisek and co-workers (288, 289) suggested that solute carrier family 41 member 1 (SLC41A1) functions as a Na^{+}/Mg^{2+} exchanger with a 2:1 stoichiometry. SLC41A1 contains 11 transmembrane domains and was originally described as a Mg^{2+} transporter mediating Mg^{2+} currents in Xenopus laevis oocytes (185). Although electrophysiological analysis could not confirm these measurements in mammalian cells, Mg^{2+} efflux studies using mag-fura 2 show Na^{+}-dependent Mg^{2+} extrusion (251, 289). Gain-of-function SNPs have been associated with Parkinson’s disease, and one mutation in SLC41A1 was identified in a patient with a nephronophthisis-like phenotype (251, 290, 576). SLC41A1 is part of a larger protein family including two additional members, SLC41A2 and SLC41A3, which are studied less extensively. Although SLC41A2 was initially described as a plasma membrane protein, it has a topology opposite to SLC41A1 (442). This finding suggests that SLC41A2 might be expressed on the membranes of organelles and may be involved in subcellular Mg^{2+} transport.

5. CNNM3

Members of the Cyclin M (CNNM) family have been proposed to function as Mg^{2+} transporters (184, 545). CNNM1 is mainly expressed in brain, CNNM2 expression is high in kidney, and CNNM4 is primarily expressed in intestine (105). In contrast, CNNM3 has a ubiquitous expression pattern and may play a role in the maintenance of cellular Mg^{2+} homeostasis. A recent study shows that CNNM3 transports Mg^{2+}, and its activity is regulated by oncogene PRL2 (217). The interaction between PRL2 and CNNM3 is essential for Mg^{2+} influx that drives tumor growth. Therefore, CNNM3 should be considered in future studies on cellular Mg^{2+} handling in nonpathological conditions.

6. MRS2

Although most studied in yeast, MRS2 (mitochondrial RNA splicing 2) is considered to be the primary Mg^{2+} channel on the mitochondrial membrane (592). Knockdown of MRS2 results in reduced Mg^{2+} uptake in mitochondria and cell death (399). Using the newly developed mitochondrial Mg^{2+} fluorescent probe KMG-301, Shindo et al. (475) revealed that MRS2 regulates intramitochondrial Mg^{2+} concentrations. This finding is interesting, since it indicates that mitochondria may store intracellular Mg^{2+}. Given that Mg^{2+} is of major importance for ATP binding, intramitochondrial Mg^{2+} concentrations may indirectly influence the progression of the citric acid cycle. Recently, it was shown that MRS2 mutations cause demyelination. The relevance of this observation to Mg^{2+} homeostasis still remains to be determined (296).

7. Others

In addition to the aforementioned Mg^{2+} transporters, several other proteins have been proposed to transport Mg^{2+}. However, these claims are based mainly on overexpression in the Xenopus oocytes model, and functional evidence for these proteins is scarce. For example, the nonimprinted in Prader-Willi/Angelman syndrome (NIPA) family of proteins has been proposed to transport Mg^{2+}, based on Mg^{2+} currents in Xenopus laevis oocytes (182), but recent studies indicate that NIPA proteins have a role in bone morphogenetic protein (BMP) signaling (525). Likewise, Huntingtin-interacting protein 14 (HIP14) was thought to mediate Mg^{2+} fluxes at the Golgi membrane (183). Now it has become apparent that its main function consists of palmitoyl acyltransferase activity, specifically involved in the palmitoylation of Huntington (129, 582). Therefore, the role of NIPA proteins and HIP14 in Mg^{2+} transport should be questioned. Additionally, members of the membrane Mg^{2+} transporter (MMgT) family have been shown to transport divalent cations in Xenopus oocytes (186). However, as they have only one transmembrane domain after signal peptide cleavage, it is unlikely that they form functional Mg^{2+} transporters themselves. It is possible that MMgT proteins may form subunits of other Mg^{2+} channels, and as a consequence, future studies should be directed to the identification of its protein partners.
D. Cell Signaling

Mg$^{2+}$ acts as a physiological Ca$^{2+}$ antagonist within cells, and as a result, the Mg$^{2+}$/Ca$^{2+}$ ratio is of major importance for the activity of Ca$^{2+}$-ATPases and other Ca$^{2+}$ transporting proteins (257). Small changes in the Mg$^{2+}$ availability within the cell may therefore cause disturbed Ca$^{2+}$ signaling or Ca$^{2+}$ toxicity.

Since 1974, when Mg$^{2+}$ influx was detected upon insulin stimulation, several groups have suggested a second messenger role for Mg$^{2+}$ (311, 322, 503). Most recently in a study on T-cell activation, MagT1 channels were shown to mediate Mg$^{2+}$ influx upon T-cell receptor activation and EGF stimulation (311). In these T cells, Mg$^{2+}$ activates phospholipase C-γ (PLCγ1), resulting in reduced phosphorylation of protein kinase C (PKC) and inositol trisphosphate (IP$_3$) generation downstream, eventually leading to reduced Ca$^{2+}$ influx (FIGURE 1). In contrast, other reports suggest that PLCγ activation precedes Mg$^{2+}$ influx (240, 241). The proposition of Mg$^{2+}$ as a dynamic second messenger raises many questions. How do MagT1 or other Mg$^{2+}$ transporters facilitate rapid Mg$^{2+}$ influx when the intracellular and extracellular Mg$^{2+}$ concentrations are almost equal? What mechanism is involved in managing Mg$^{2+}$ after the initial influx, given the absence of Mg$^{2+}$ pumps and major Mg$^{2+}$ binding proteins? Follow-up studies demonstrated that MagT1-deficient cells have severely reduced basal intracellular Mg$^{2+}$ concentrations (79). These results suggest that the effects seen on PLCγ1 are dependent on general intracellular Mg$^{2+}$ availability and further question the physiological role of variable fluxes that are proposed in the second messenger theory. Studying Mg$^{2+}$ dynamics within the cell using fluorescent probes may help to draw definitive conclusions on this matter.

E. Cell Proliferation

Given its effect on RNA, DNA, and protein synthesis, Mg$^{2+}$ is an important factor in the control of cell proliferation. Over the last 40 years, the role of Mg$^{2+}$ in cell cycle control, protein synthesis, and growth factor response has been extensively studied, pioneered by several groundbreaking studies from the group of Harry Rubin (428, 430). Cell proliferation is largely dependent on protein synthesis, more than DNA or RNA synthesis. Inhibition of protein synthesis directly shuts down DNA synthesis, whereas there is a 2-h delay to achieve the same effect using RNA synthesis inhibitors (279). Protein synthesis is highly dependent on intracellular Mg$^{2+}$ concentrations; increasing the Mg$^{2+}$ content amplifies protein synthesis within 60 min, whereas DNA synthesis is only enhanced after 10 h (429, 514). Activation of proliferation is initiated by growth factors that increase glucose uptake and protein synthesis within minutes (225). Interestingly, Mg$^{2+}$ is tightly regulated during these intracellular processes. Initial studies in cultured cells showed that applying insulin induced 20% higher intracellular Mg$^{2+}$ concentrations after 16 h (445). Later studies with EGF using the fluorescent probe mag-fura 2 showed an impressive fourfold increase of intracellular Mg$^{2+}$ from 0.3 to 1.4 mM after 20 min of epidermal growth factor (EGF) stimulation (199). The authors state that a rise in Mg$^{2+}$ precedes DNA synthesis, but coincides with and thus may contribute to increases in protein synthesis. Recent studies identifying the molecular mediators of Mg$^{2+}$-dependent cell proliferation have resulted in the membrane, magnesium, mitosis (MMM) model (432). The MMM model proposes that, upon growth factor binding, Mg$^{2+}$ enters the cell or is released from phospholipids in the cell membranes (FIGURE 1). Increased cytosolic Mg$^{2+}$ levels contribute to ribosomal activity and protein synthesis, eventually leading to DNA replication and mitosis. The mammalian target of rapamycin (mTOR) complex is a critical component of the MMM model, as it is the master regulator of cell cycle progression and proliferation (551). Growth factors binding to their receptors leads to phosphoinositol 3-kinase (PI3K) phosphorylation, which activates the mTOR complex (431, 548). Activation of mTOR is MgATP$^{2-}$ dependent, and ATP has been suggested as the main regulator of mTOR activity. However, ATP levels do not change upon growth factor stimulation, whereas Mg$^{2+}$ levels do (445, 537). Therefore, the MMM model proposes Mg$^{2+}$ as the primary regulator of mTOR dynamics and cell proliferation.

1. Cancer

Tumor cells contain high concentrations of intracellular Mg$^{2+}$ (508). In a mammary tumor cell line, Mg$^{2+}$ can be transported into the cell even when extracellular Mg$^{2+}$ concentrations were below physiological levels (203, 264, 566). Mg$^{2+}$ uptake via divalent cation channel TRPM7 has been suggested to stimulate tumor cell proliferation (203, 264). TRPM7 expression is upregulated in hepatoma, pancreatic adenocarcinoma, gastric cancer, and breast cancer tissue (203, 278, 346, 581). Although TRPM7 has been primarily described as a Mg$^{2+}$ channel, it is also permeable for other divalent cations (349). Given the involvement of Mg$^{2+}$ in cell proliferation, the influx of Mg$^{2+}$ via TRPM7 has been proposed as the main regulator of tumor growth. However, recent studies using prostate cancer cells suggest that TRPM7-mediated Ca$^{2+}$ uptake may also play an important role in tumor growth (501). The expression of Mg$^{2+}$ transporter CNNM3 is increased in human breast cancer tissue (217). CNNM3 binds oncogene PRL2 and facilitates the entry of Mg$^{2+}$ in the tumor cell to drive cell proliferation. Elevated intracellular Mg$^{2+}$ concentrations have been suggested to be beneficial for tumor growth because Mg$^{2+}$ regulates several cancer-associated enzymes including telomerase and protein phosphatase 1D, which are involved in the glycolytic cycle and BER (74). However, the regulatory role of Mg$^{2+}$ on these enzymes during the pathogenic state of tumor cell proliferation has never been inves-
tigated and, therefore, the exact role of Mg\(^{2+}\) in enzymatic regulation in cancer remains speculative.

In contrast to the proliferative phase of tumor growth, in which tumor cells have high intracellular Mg\(^{2+}\) concentrations, low intracellular Mg\(^{2+}\) concentrations are associated with increased rates of carcinogenesis and metastasis (74). Low Mg\(^{2+}\) conditions and impaired activity of DNA repair mechanisms reduces DNA protection against oxidative stress. Indeed, low dietary Mg\(^{2+}\) intake has been associated with the risk of several types of cancers. Epidemiological studies have established a correlation between low Mg\(^{2+}\) intake and colon cancer risk (160, 302, 534). In addition, in a study with 1,200 lung cancer patients and a similar number of controls, low dietary Mg\(^{2+}\) intake was associated with reduced lung cancer risk (326). However, these results could not be reproduced in other patient cohorts (325, 502).

III. REGULATION OF MAGNESIUM HOMEOSTASIS

Mg\(^{2+}\) serum concentrations range between 0.7 and 1.1 mM in healthy people (323). To maintain constant plasma Mg\(^{2+}\) levels, the United States Food and Nutrition Board recommends a daily Mg\(^{2+}\) intake of 420 mg for men and 320 mg for women (1). Mg\(^{2+}\) homeostasis depends on the collaborative actions of the intestine, responsible for Mg\(^{2+}\) uptake from food, the bone, which stores Mg\(^{2+}\) in its hydroxy-apatite form, and the kidneys, regulating urinary Mg\(^{2+}\) excretion (FIGURE 2).

A. Magnesium in Intestine

Given a daily Mg\(^{2+}\) intake of 370 mg, ~30–50% is absorbed in the intestine, resulting in a net uptake of ±100 mg. However, if Mg\(^{2+}\) intake is low, early reports suggest that up to 80% of dietary Mg\(^{2+}\) can be absorbed (189). Mg\(^{2+}\) absorption in the gut depends on two separate pathways; paracellular transport is responsible for bulk Mg\(^{2+}\) absorption and takes place mostly in the small intestine, whereas fine-tuning occurs in the cecum and colon via transcellular transport (FIGURE 3). In spite of this, the intestine seems to have a limited role in regulation of the Mg\(^{2+}\) balance. In contrast to other minerals, intestinal Mg\(^{2+}\) absorption is poorly regulated and depends mainly on Mg\(^{2+}\) intake (216, 461). Thus the kidneys presumably primarily regulate the maintenance of Mg\(^{2+}\) homeostasis.

1. Small intestine

Mg\(^{3+}\) absorption in the small intestine is hypothesized to be exclusively of a paracellular nature, since Mg\(^{2+}\) absorption...
in this region of the intestine correlates linearly to luminal Mg\(^{2+}\) concentrations (273, 409). Moreover, the epithelial Mg\(^{2+}\) channel TRPM6 is not expressed in the small intestine (196). Mg\(^{2+}\) is poorly absorbed in the duodenum, where unfavorable electrochemical gradients may even result in a limited amount of paracellular Mg\(^{2+}\) excretion (389). In more distal parts of the small intestine, such as late jejunum and ileum, the driving force for passive Mg\(^{2+}\) transport is established by the high luminal Mg\(^{2+}\) concentration and the lumen-positive transepithelial voltage of ~15 mV (164). The \(K_m\) for Mg\(^{2+}\) transport in the distal small intestine has been reported to be in the range of 4–12 mM (343, 461). These results suggest that NaCl and water absorption are prerequisites for Mg\(^{2+}\) uptake, since water absorption concentrates luminal Mg\(^{2+}\). Tight junction permeability underlying paracellular Mg\(^{2+}\) transport is still poorly understood. The small intestine is described as the most ion-permeable part of the intestine because of the relatively low expression of “tightening” claudins 1, 3, 4, 5, and 8 (20, 301). Claudins 16 and 19, which are linked to Mg\(^{2+}\) transport, are not expressed in the intestine (20, 246). The exact composition of the tight junction complex facilitating intestinal Mg\(^{2+}\) absorption remains to be elucidated.

2. Large intestine

Mg\(^{2+}\) absorption in cecum and colon is thought to be transcellular of nature and is mediated by TRPM6 and TRPM7 on the luminal side of the enterocyte (FIGURE 3). Intestinal expression of TRPM6 is located in cecum and colon (196, 300). In a study with rat colon epithelium, 37% of Mg\(^{2+}\) was transported transcellularly (272). This suggests signif-
icant paracellular transport of Mg\(^{2+}\) in the colon, which would be unlikely given the expression of tightening Claudins 3, 4, and 8 in this segment (301). In contrast to Ca\(^{2+}\), Mg\(^{2+}\) transport in colon is independent of 1,25-dihydroxyvitamin D\(_3\) [1,25(OH)\(_2\)D\(_3\)] signaling, nor is TRPM6 expression dependent on 1,25(OH)\(_2\)D\(_3\) (196, 272). It has been suggested that the basolateral Mg\(^{2+}\) extrusion mechanism of the enterocyte is coupled to the Na\(^{+}\) gradient (424). Indeed, the results of a recent study using CNNM4 KO mice suggest that CNNM4 may act as a Na\(^{+}/Mg\(^{2+}\) exchanger at the basolateral membrane of enterocytes (575). CNNM4 KO mice suffer from hypomagnesemia, and functional analysis using Magnesium Green showed that CNNM4 overexpression increased Mg\(^{2+}\) efflux in HEK293 cells. However, patients with CNNM4 mutations do not suffer from hypomagnesemia (387, 402).

### B. Magnesium in Bone

Approximately 50–60% of the total body Mg\(^{2+}\) content is stored in bone. Serum Mg\(^{2+}\) concentrations are closely related to bone metabolism; bone surface Mg\(^{2+}\) is continuously exchanged with blood Mg\(^{2+}\) (14). In bone, Mg\(^{2+}\) ions bind at the surface of the hydroxyapatite crystals. Mg\(^{2+}\) increases the solubility of P, and Ca\(^{2+}\) hydroxyapatite and thereby acts on the crystal size and formation (443). Mg\(^{2+}\) increases the secretion of proinflammatory cytokines such as tumor necrosis factor (TNF)-\(\alpha\), interleukin (IL)-1\(\beta\), and substance P (434, 552), all of which have been implicated in the release of proinflammatory molecules such as IL-1\(\beta\), TNF-\(\alpha\), and SP, which stimulate osteoclast activity. In bone development, Mg\(^{2+}\) stimulates chondrocyte column formation. IL-1\(\beta\), interleukin-1\(\beta\); TNF-\(\alpha\), tumor necrosis factor-\(\alpha\); SP, substance P. 

![FIGURE 4](image)

**FIGURE 4.** Magnesium storage in bone. Mg\(^{2+}\) stimulates osteoblast proliferation in bone and inhibits the release of proinflammatory molecules such as IL-1\(\beta\), TNF-\(\alpha\), and SP, which stimulate osteoclast activity. In bone development, Mg\(^{2+}\) stimulates chondrocyte column formation. IL-1\(\beta\), interleukin-1\(\beta\); TNF-\(\alpha\), tumor necrosis factor-\(\alpha\); SP, substance P.

expression study in the Mg\(^{2+}\)-transporting segment of the kidney, SOX9 mRNA was the most increased in the low-Mg\(^{2+}\) diet group (104). These findings suggest that SOX9 is an important transcription factor for bone and kidney Mg\(^{2+}\) homeostasis.

### C. Magnesium in Kidney

Approximately 2,400 mg of Mg\(^{2+}\) is filtered by the glomerulus on a daily basis. The nephron recovers 95–99% of this; the remaining 100 mg leaves the body via the urine (FIGURE 2).

1. **Proximal tubule**

The mechanisms of proximal tubule (PT) Mg\(^{2+}\) reabsorption are poorly understood, but early micropuncture studies showed that \(\sim 10–25\%\) of Mg\(^{2+}\) is reabsorbed by the proximal convoluted tubule segment of the nephron (304, 411). In the glomeruli, 70% of the serum Mg\(^{2+}\) is freely filterable, suggesting that the concentration in the glomerular filtrates and thus at the start of the PT ranges between 0.5–0.7 mM. The transepithelial potential difference ranges from slightly lumen negative (–6 mV) in the early parts of the PT to positive (3 mV) in later parts (287). Micropuncture studies have shown that a 1.9 ratio between the concentrations of Mg\(^{2+}\) in the tubular fluid and the interstitial fluid is necessary to initiate Mg\(^{2+}\) transport (304). This finding could be explained by the poor tight junction permeability for Mg\(^{2+}\) in PT. As a result, water uptake via aquaporin 1 (AQP1) precedes Mg\(^{2+}\) reabsorption (410) (FIGURE 5).

Consequently, Mg\(^{2+}\) reabsorption mainly occurs in the late parts of the PT, where the transepithelial chemical Mg\(^{2+}\) gradient is sufficient to favor Mg\(^{2+}\) transport. PT Mg\(^{2+}\) reabsorption is generally considered to be a passive paracellular process, but there might be some transcellular Mg\(^{2+}\) transport via a poorly characterized amiloride-sensitive mechanism (254). In both cases, sufficient Na\(^{+}\) transport is required to drive water transport that is a prerequisite for Mg\(^{2+}\) reabsorption. Hormonal effects on Na\(^{+}\) reabsorption in the PT will therefore also affect Mg\(^{2+}\) reabsorption in this segment. However, disturbances of proximal tubular
FIGURE 5. Magnesium reabsorption in the kidney. The glomerulus filters the blood, and along the nephron 95% is reabsorbed. In the late proximal tubule (PT), Na⁺ and H₂O reabsorption via NHE3 and AQP1 are prerequisites for paracellular Mg\(^{2+}\) transport. Approximately 10–25% of Mg\(^{2+}\) is reabsorbed in the proximal tubule. Bulk Mg\(^{2+}\) reabsorption (50–70%) takes place in the thick ascending limb of Henle’s Loop (TAL). In TAL, Mg\(^{2+}\) reabsorption take place paracellular and depends on the uptake of Na⁺ and K⁺ via NKCC2. Fine-tuning (10%) of Mg\(^{2+}\) transport takes place transcellular in the distal convoluted tubule (DCT). In DCT, TRPM6 facilitates Mg\(^{2+}\) uptake from the pro-urine, which depends on the voltage gradient set by backleak of K⁺ via ROMK and Kv1.1 potassium channels. At the basolateral membrane, Mg\(^{2+}\) is extruded via an unknown mechanism, which may be regulated by CNNM2 acting as Mg\(^{2+}\) sensor: Mg\(^{2+}\) extrusion depends on the Na⁺ gradient, set by the Na⁺-K⁺-ATPase. The activity of the Na⁺-K⁺-ATPase is in turn dependent on K⁺ recycling via Kir4.1, FXYD2 transcription encoding the γ-subunit of the Na⁺-K⁺-ATPase is regulated by HNF1β and PCBD1. Regulation of Mg\(^{2+}\) transport in DCT depends on EGF and insulin. Upon activation of the EGFR and IR, an intracellular signaling cascade including PI3K, Akt, and Rac1 results in an increased TRPM6 membrane expression and increased channel activity. Additionally, estrogens have been shown to increase TRPM6 expression. PT, proximal tubule; TAL, thick ascending limb of Henle’s loop; DCT, distal convoluted tubule; CNT, connecting tubule; CD, collecting duct; NHE3, Na⁺-H⁺ exchanger type 3; AQP1, aquaporin 1; NKCC2, Na⁺-K⁺-2Cl⁻ cotransporter; ROMK, renal outer medulla K⁺ channel; ClC-Kb, chloride channel Kb; Kv1.1, voltage-gated K⁺ channel 1.1; TRPM6, transient receptor potential melastatin type 6; NCC, Na⁺-Cl⁻ cotransporter; CNNM2, cyclin M2; FXYD2, FXYD-domain containing 2; HNF1β, hepatocyte nuclear factor 1β; PCBD1, pterin-4 alpha-carbinolamine dehydratase 1; EGF, epidermal growth factor; EGFR, epidermal growth factor receptor; IR, insulin receptor; PI3K, phosphoinositide 3-kinase; Rac1, Ras-related C3 botulinum toxin substrate 1; Cdk5, cyclin-dependent kinase 5.


Mg\(^{2+}\) reabsorption generally do not result in clinical symptoms, since more distal segments will compensate for reduced Mg\(^{2+}\) uptake in PT.

2. Thick ascending limb of Henle’s loop

Whereas most electrolytes are majorly transported in the PT, the thick ascending limb of Henle’s loop (TAL) is the main location for Mg\(^{2+}\) reabsorption (236, 299, 420). Due to the unique properties of this segment, ~50–70% of filtered Mg\(^{2+}\) is reabsorbed here. Most of the Mg\(^{2+}\) is reabsorbed by the cortical part of the TAL, since medullary Mg\(^{2+}\) reabsorption is negligible (468). Paracellular bulk Mg\(^{2+}\) transport is dependent on the lumen-positive transepithelial voltage (+10 mV) that is determined by the activity of the Na\(^{+}\)-K\(^{+}\)-2Cl\(^{-}\) cotransporter (NKCC2) and the subsequent secretion of K\(^{+}\) at the apical membrane (192). Inhibition of NKCC2 by furosemide diuretics therefore decreases TAL Mg\(^{2+}\) reabsorption (see sect. VB3). Further contributors to the transepithelial membrane voltage are K\(^{+}\) secretion via renal outer medullary potassium channel (ROMK) and paracellular backflux of Na\(^{+}\) ions as a result of decreasing luminal Na\(^{+}\) concentrations (329).

The reabsorption of Mg\(^{2+}\) in the TAL follows the paracellular pathway and therefore depends on the tight junction permeability (FIGURE 5). Tight junctions form a physical and chemical barrier between the epithelial cells. Their major components are proteins of the claudin family. Currently, 26 claudins have been described in humans (209). Tight junction permeability is determined by the individual claudins in each tight junction complex. TAL tubuli are known to express claudins 3, 10, 11, 14, 16, and 19. Claudins 16 and 19 are considered to be the main claudins influencing Mg\(^{2+}\) permeability, since mutations in these proteins result in renal Mg\(^{2+}\) wasting (293) (FIGURE 5).

However, the role of claudin 16 in Mg\(^{2+}\) reabsorption is controversial. Claudin 16 was initially considered to act as a paracellular Mg\(^{2+}\) channel, but this hypothesis has not been unequivocally confirmed (207, 245). Reports using the claudin 16 knockdown (KD) mouse model and the LLC-PK\(_1\) cell model suggest that claudin 16 increases Na\(^{+}\) permeability (227, 245, 466). This would imply that claudin 16 is mainly involved in the regulation of the transepithelial voltage gradient by controlling the paracellular Na\(^{+}\) back-leak. In MDCK-C7 cells overexpressing claudin 16, Na\(^{+}\) permeability yet remained stable, whereas Mg\(^{2+}\) permeability increased significantly (207). Claudin 16 KD mice demonstrate a twofold lower permeability ratio for Na\(^{+}\) over Cl\(^{-}\) without a change in paracellular conductance. Consequently, the transepithelial voltage collapsed, reducing the driving force for Mg\(^{2+}\) reabsorption in TAL (466).

Claudin 19 has been studied less extensively, but has been suggested to increase the tight junction barrier function (24). The claudin 19 KD mouse exhibits highly increased urinary excretion of K\(^{+}\), Mg\(^{2+}\), and Ca\(^{2+}\), but Mg\(^{2+}\) is the only electrolyte altered at the serum level (246). The discrepancy between studies with claudin 16 and claudin 19 isoforms might be explained by their interdependence in forming functional tight junction barriers (246, 247). Both in vitro and in vivo studies demonstrated that claudins 16 and 19 need to interact for proper insertion in the tight junction to become functionally active. Further differences in experimental results may depend on the endogenous expression of other claudin isoforms in the specific cell types used in these experiments.

Claudin 14 reduces the cation specificity of tight junction barriers, when coexpressed with claudin 16, or with claudin 16-claudin 19 complexes (180). Consequently, claudin 14 KO mice exhibit increased serum Mg\(^{2+}\) values and decreased urinary Mg\(^{2+}\) excretion. This agrees with previous findings in MDCK cells showing that claudin 14 acts as a nonspecific cation blocker (45, 560). Studies on claudin 14 KO mice have mainly focused on Ca\(^{2+}\) homeostasis, since claudin 14 expression is highly Ca\(^{2+}\) sensitive (121, 180). The CaSR regulates claudin 14 expression and Ca\(^{2+}\) reabsorption in the TAL by downregulation of two microRNAs, miR-9 and miR-374. Since CaSR is also activated by Mg\(^{2+}\), although to a lesser extent than Ca\(^{2+}\), it would be interesting to address the effect of elevated serum Mg\(^{2+}\) levels on claudin 14 expression in future studies.

Recently, claudin 10 has been identified as an important factor in cation selectivity in TAL, as demonstrated in a mouse model where claudin 10 was deleted specifically in this segment (57). Claudin 10 TAL-KO mice show hypermagnesemia, nephrocalcinosis, and impaired paracellular Na\(^{+}\) permeability. In the absence of claudin 10, TAL tight junctions became more permeable to Ca\(^{2+}\) and Mg\(^{2+}\) and the transepithelial voltage increased. These results are in line with in vitro studies overexpressing claudin 10b, suggesting that this splice variant is mainly expressed in TAL (57, 208).

3. Distal convoluted tubule

The distal convoluted tubule (DCT) determines the final urinary Mg\(^{2+}\) concentration, since no reabsorption of Mg\(^{2+}\) takes place beyond this segment. Approximately 10% of the total Mg\(^{2+}\) is reabsorbed by tightly regulated transcellular transport mechanisms (64). DCT cells form a high-resistance epithelium with a lumen-negative voltage of approximately −5 mV (192, 571). In DCT, TRPM6 divalent cation channels mediate luminal Mg\(^{2+}\) uptake (234, 235) (FIGURE 5). Within the kidney, TRPM6 is specifically expressed in DCT, and its activity is regulated by intracellular Mg\(^{2+}\) (540). TRPM6 contains six transmembrane spanning domains with a pore region between the fifth and sixth segment and a large kinase domain fused to the channel’s intracellular COOH terminus. TRPM6 may function in homo- and heteromeric tetramers with TRPM7, al-
MAGNESIUM IN MAN

Though there is some controversy about the necessity of TRPM7 for TRPM6 function (314, 588).

TRPM6 is regulated by numerous factors at the level of transcription, plasma membrane availability, and activity (69). EGF and insulin act on TRPM6 by a PI3K-Akt-Rac1 dependent mechanism, increasing the insertion of TRPM6 in the membrane (361, 515) (FIGURE 5). Insulin may directly affect TRPM6 activity through cytin-dependent kinase 5 (cdk5)-dependent phosphorylation of the channel. Patients with reduced EGFR or insulin receptor (IR) activity are therefore more susceptible to hypomagnesemia (361, 459). Additionally, estrogens increase TRPM6 mRNA expression (196). Over the last decade, several important interactors of TRPM6, including receptor for activated C-kinase 1 (RACK1) and prohibitin2 (PHB2/REA), have been identified (70, 71). RACK1 interacts with the a-kinase domain of TRPM6 in the autophosphorylated state, thereby reducing TRPM6 activity (70). Other modulators of TRPM6 activity include dietary Mg²⁺, pH, and ATP (516). Interestingly, acidification-induced current potentiation is dependent on residues p.Glu1024 and p.Glu1029, which also determine the pore selectivity for Mg²⁺ (313, 367). Moreover, recent findings indicate that TRPM6 is inhibited by low concentrations of intracellular ATP (IC₅₀ 29 μM), questioning the physiological activity of monomeric TRPM6 channels (588). Extracellular ATP also inhibits TRPM6 activity via the purinergic receptor P2X4 (103).

A chemical gradient for Mg²⁺ entry in DCT cells is almost absent. The luminal Mg²⁺ concentrations vary between 0.2 and 0.7 mM, and the intracellular Mg²⁺ levels are typically in the range of 0.5–1 mM. Therefore, luminal Mg²⁺ entry is purely dependent on the negative membrane potential in the DCT cell. Luminal K⁺ channels are indispensable for maintaining the necessary driving force for Mg²⁺ uptake. The voltage-gated K⁺ channel Kv1.1 has been suggested to provide efflux K⁺ currents resulting in hyperpolarization of the luminal membrane, although expression levels in the DCT are limited (104, 176). To prevent Mg²⁺ overload and hyperpolarization of the luminal membrane, intracellular Mg²⁺ blocks Kv1.1 (179). Interestingly, recent studies suggest that other potassium channels in the luminal membrane of DCT cells may have comparable roles. ROMK is prominently expressed in DCT, and its expression in this segment is regulated by dietary Mg²⁺ (104, 572). Similar to Kv1.1, intracellular Mg²⁺ blocks ROMK currents, suggesting a regulatory function on Mg²⁺ homeostasis (578). Moreover, indirect inhibition of ROMK by aldosterone or epithelial Na⁺ channel (ENaC) blockers represent the only effective approach to prevent renal Mg²⁺ wasting in most clinical situations (140).

Several proteins have been proposed to mediate Mg²⁺ extrusion to the bloodstream, but general consensus of the extrusion mechanism has not been reached (424). Due to the absence of a representative DCT cell model, the properties of Mg²⁺ extrusion have not been elucidated. Nevertheless, over the last decade several groups claimed to have identified Mg²⁺ extrusion proteins. Originally described in 2002, cyclin M2 (CNNM2, previously known as ACDP2) is exclusively expressed at the basolateral membrane of DCT and CNT cells within the kidney (105, 497, 545). Moreover, expression of CNNM2 is sensitive to dietary Mg²⁺ availability (104, 497). CNNM2 was initially depicted as Mg²⁺ transporter, since overexpression in Xenopus laevis oocytes allows uptake of a variety of divalent cations, with highest affinity for Mg²⁺ (184). However, these results could not be confirmed in mammalian cell lines (497). Alternatively, a Mg²⁺-sensing function has been proposed, since CNNM2 harbors a Mg-ATP binding site in its cystathionine-β-synthase (CBS) domains (105). CNNM2 increases Mg²⁺ uptake in HEK293 cells (28). Nevertheless, it remains unclear whether CNNM2 mediates Mg²⁺ uptake directly or activates other Mg²⁺ carriers.

Recently mutations in the SLC41A1 Mg²⁺ transporter were described to cause a nephronophthisis-like phenotype (251). Immunohistological studies showed expression in DCT, but the stainings were not conclusive about the subcellular localization (apical or basolateral) of SLC41A1 proteins (251). By the use of Mag-Fura, SLC41A1 was demonstrated to increase both Mg²⁺ absorption and Mg²⁺ extrusion (251, 289). These results suggest that SLC41A1 plays a role in DCT Mg²⁺ reabsorption, although further studies are necessary to elucidate the mechanisms by which SLC41A1 mediates Mg²⁺ transport.

Parvalbumin is exclusively expressed in DCT within the kidney, where it may function as a Ca²⁺/Mg²⁺ buffer (456). Although parvalbumin has much higher affinity for Ca²⁺ than for Mg²⁺ (dissociation constants are ~5–10 mM for Ca²⁺ and ~30 μM for Mg²⁺), the cation binding sites of parvalbumin will be mainly occupied by Mg²⁺ (377). This can be explained by the fact that the intracellular concentration of Mg²⁺ (0.5–1 mM) vastly exceeds that of Ca²⁺ (50–100 nM). In mouse and human kidney, parvalbumin is exclusively expressed in the early DCT (44). In late DCT and CNT, calbindin-D₂₈K is the main Ca²⁺-binding protein. The exact role of parvalbumin in DCT remains to be investigated. Parvalbumin KO mice do not have altered serum or urine Mg²⁺ levels under basal conditions; these mice have reduced NCC expression, but display normal tubule morphology (44). DCT parvalbumin expression is highly sensitive to dietary Mg²⁺ availability, suggesting an important role for parvalbumin in Mg²⁺ reabsorption in DCT (104).

IV. MAGNESIUM IN PHYSIOLOGY AND PATHOPHYSIOLOGY

The human body contains ~24g Mg²⁺, of which 99% is stored in bone, muscle, and other soft tissues. Mg²⁺ is crit-
ical to the function of basically every organ in the human body. Moreover, Mg\(^{2+}\) deficiency is associated with a wide range of diseases, and as a result Mg\(^{2+}\) supplementation is considered as potential treatment in many of them (TABLE 2). This part of the review focuses on the organ-specific functions of Mg\(^{2+}\) and provides an overview of all major diseases in which Mg\(^{2+}\) may play a role.

A. Magnesium in Brain

Low serum Mg\(^{2+}\) values are associated with a wide range of neurological diseases such as migraine, depression, and epilepsy. Neuronal Mg\(^{2+}\) concentrations are of major importance in the regulation of N-methyl-D-aspartate (NMDA) receptor excitability. NMDA receptors are essential for excitatory synaptic transmission, neuronal plasticity, and excitotoxicity and therefore play an important role in developmental plasticity, learning, and memory (384). NMDA receptors are activated upon glutamate binding and mediate the influx of Ca\(^{2+}\) and Na\(^{+}\) ions and the efflux of K\(^{+}\) ions. Every NMDA receptor consists of four subunits, each with different biochemical properties (97). In Mg\(^{2+}\) deficiency, NMDA receptors become hyperexcitable, which can be explained by inhibitory function of extracellular Mg\(^{2+}\) on the receptors (335, 372) (FIGURE 6). Glutamate from the presynaptic neuron will bind both the ionotropic 2-amino-3-(3-hydroxy-5-methyl-isoxazol-4-yl)propanoic acid (AMPA) and NMDA receptors on the postsynaptic neuron. At a normal membrane potential of −70 mV, Mg\(^{2+}\) ions block NMDA receptors. Therefore, only AMPA receptors will be activated and consequently facilitate an influx of cations. Only when the membrane potential rises above −60 mV the Mg\(^{2+}\) block is relieved and NMDA receptors are opened upon glutamate binding. The unlocking mechanism consists of a slow and a fast component, which depend on the relative expression of subunits that compose the channel (87, 536). Upon reduced extracellular Mg\(^{2+}\) concentrations, less NMDA channels will be blocked, and more NMDA channels can be opened at relatively low membrane potentials (351). This increased excitatory postsynaptic potential causes hyperexcitability of the neurons. In the adult brain, this process is further amplified by the action of inhibitory \(\gamma\)-aminobutyric acid (GABA) receptors, whose function is also regulated by Mg\(^{2+}\). GABA\(_A\) receptors are ionotropic anion channels that open upon GABA binding and facilitate Cl\(^−\) influx (250). Since the equilibrium potential of Cl\(^−\) is 10–20 mV lower than the membrane potential, this influx contributes to hyperpolarization of the neuronal cells. Extracellular Mg\(^{2+}\) stimulates GABA\(_A\) receptors resulting in hyperpolarized neuronal cells (353). When Mg\(^{2+}\) concentrations in the central nervous system (CNS) are low, GABA\(_A\) receptors are less stimulated. Consequently, the membrane potential will be higher.

<table>
<thead>
<tr>
<th>Disease</th>
<th>Cochrane Review</th>
<th>Large-Scale Clinical Studies</th>
<th>Guidelines</th>
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<tbody>
<tr>
<td><strong>First drug of choice</strong></td>
<td></td>
<td></td>
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<tr>
<td>Preeclampsia</td>
<td>RR: 0.41, 95% CI: 0.29–0.58 (130)</td>
<td></td>
<td>(223)</td>
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<tr>
<td>Arrhythmia–Torsades des Pointes</td>
<td></td>
<td></td>
<td>(591)</td>
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<tr>
<td><strong>Alternative drug of choice</strong></td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Migraine</td>
<td>RR: 0.53, 95% CI: 0.05–5.31 (404)</td>
<td>Magnetic</td>
<td>(3)</td>
</tr>
<tr>
<td>Asthma</td>
<td>RR: 0.59, 95% CI: 0.49–0.70* (312)</td>
<td></td>
<td>(476)</td>
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<tr>
<td>Super-refractory status epilepticus</td>
<td></td>
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<tr>
<td>Muscle cramps</td>
<td>No. cramps: −3.93%, 95% CI: −21.12 to 13.26% (168)</td>
<td></td>
<td>(274)</td>
</tr>
<tr>
<td><strong>Experimental</strong></td>
<td></td>
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<tr>
<td>Stroke</td>
<td></td>
<td>FAST-MAG Images</td>
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<tr>
<td>Subarachnoid hemorrhage</td>
<td>RR: 0.75, 95% CI: 0.57–1.00* (125)</td>
<td>IMASH MASH-II</td>
<td></td>
</tr>
<tr>
<td>Myocardial infarction</td>
<td>OR: 0.59, 95% CI: 0.49–0.70* (312)</td>
<td>LIMIT-2 ISIS-4</td>
<td></td>
</tr>
<tr>
<td>Hypertension</td>
<td>DBP: −2.2 mmHg, 95% CI: −3.4 to −0.9 (119)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Traumatic brain injury</td>
<td>GS: 0.02, 95% CI: −0.38 to 0.041 (25)</td>
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</tr>
</tbody>
</table>

This table summarizes all diseases in which Mg\(^{2+}\) has been considered as treatment by the official American guidelines. Moreover, it lists all large-scale clinical studies and meta-analyses of small-scale clinical studies by the Cochrane collaboration. RR, risk ratio; OR, odds ratio; CI, confidence interval; DBP, diastolic blood pressure; GS, Glasgow score. Reference numbers are given in parentheses. *In addition to standard nifedipine treatment. Early mortality in patients treated with <75 mmol of magnesium compared with placebo groups.
which in turn relieves the Mg$^{2+}$ block of the NMDA receptor and contributes to hyperexcitability of the neurons.

The final mechanism contributing to the hyperexcitability of NMDA-receptor rich neurons is inhibiting glutamate release from the presynaptic neuron. Release of glutamate can be inhibited by high extracellular Mg$^{2+}$ concentrations (257, 318, 486). Although the exact mechanism by which Mg$^{2+}$ reduces glutamate release is still unknown, it could be related to the inhibition of voltage-gated Ca$^{2+}$ channels, as glutamate release is triggered by an influx of Ca$^{2+}$ after an action potential (364).

As a result of the described mechanisms, low extracellular Mg$^{2+}$ levels in the CNS contribute to the hyperexcitability of NDMA receptor. The excessive intracellular Ca$^{2+}$ in the neurons may lead to the production of toxic reactive oxygen species (ROS) and eventually to neuronal cell death.

In addition to increasing the hyperexcitability of excitatory neuronal pathways, Mg$^{2+}$ has an important role in the regulation of oxidative stress and the release of neuropeptides such as calcitonin gene-related peptide (CGRP) and substance P. CGRP is secreted from sensory neurons and has a vasodilatory effect (48). Mg$^{2+}$ may increase CGRP expression and secretion, as has been shown in women with preeclampsia (26, 151), although an opposite effect was reported in women with Raynaud’s phenomenon (359). Mg$^{2+}$ deficiency increases the release of substance P, which is a neuroinflammatory tachykinin (554), stimulating the release of neuropeptides such as calcitonin gene-related peptide (CGRP) and substance P. CGRP is secreted from sensory neurons and has a vasodilatory effect (48). Mg$^{2+}$ may increase CGRP expression and secretion, as has been shown in women with preeclampsia (26, 151), although an opposite effect was reported in women with Raynaud’s phenomenon (359). Mg$^{2+}$ deficiency increases the release of substance P, which is a neuroinflammatory tachykinin (554), stimulating the release of neuropeptides such as calcitonin gene-related peptide (CGRP) and substance P. CGRP is secreted from sensory neurons and has a vasodilatory effect (48). Mg$^{2+}$ may increase CGRP expression and secretion, as has been shown in women with preeclampsia (26, 151), although an opposite effect was reported in women with Raynaud’s phenomenon (359). Mg$^{2+}$ deficiency increases the release of substance P, which is a neuroinflammatory tachykinin (554), stimulating the release of neuropeptides such as calcitonin gene-related peptide (CGRP) and substance P. CGRP is secreted from sensory neurons and has a vasodilatory effect (48). Mg$^{2+}$ may increase CGRP expression and secretion, as has been shown in women with preeclampsia (26, 151), although an opposite effect was reported in women with Raynaud’s phenomenon (359). Mg$^{2+}$ deficiency increases the release of substance P, which is a neuroinflammatory tachykinin (554), stimulating the release of neuropeptides such as calcitonin gene-related peptide (CGRP) and substance P. CGRP is secreted from sensory neurons and has a vasodilatory effect (48). Mg$^{2+}$ may increase CGRP expression and secretion, as has been shown in women with preeclampsia (26, 151), although an opposite effect was reported in women with Raynaud’s phenomenon (359). Mg$^{2+}$ deficiency increases the release of substance P, which is a neuroinflammatory tachykinin (554), stimulating the release of neuropeptides such as calcitonin gene-related peptide (CGRP) and substance P. CGRP is secreted from sensory neurons and has a vasodilatory effect (48). Mg$^{2+}$ may increase CGRP expression and secretion, as has been shown in women with preeclampsia (26, 151), although an opposite effect was reported in women with Raynaud’s phenomenon (359). Mg$^{2+}$ deficiency increases the release of substance P, which is a neuroinflammatory tachykinin (554), stimulating the
secretion of inflammatory mediators such as IL-1, -2, -4, -5, -10, -12, and -13 as well as TNF-α (552, 554). Moreover, Mg\(^{2+}\) enhances the activity of nitric oxide synthases (NOS) through a NMDA receptor-dependent mechanism (80, 392). Nitric oxide (NO) has multiple functions in the brain including vasodilation, regulation of gene transcription, channel activity, and neurotransmitter release (493). Altogether, the role of Mg\(^{2+}\) in the regulation of neuropeptide release may have serious consequences in neuronal disease.

1. Migraine

Migraine has been linked to low levels of magnesium in serum and cerebrospinal fluid (CSF) (414, 457). Migraine headaches are the consequence of cortical spreading depression (CSD), which consists of an intense membrane depolarization and repolarization in neurons and glial cells (308, 388). CSD can be evoked by NMDA receptor activation (181). Therefore, patients with an increase in neuronal excitability due to low CSF Mg\(^{2+}\) levels are more susceptible to migraine attacks. Moreover, patients suffering from migraine are often hypotensive during attacks (462), which can be explained by increased NO levels. NO is an important vasodilator and modulator of brain blood flow (376). As an inhibitor of NO production, reduced Mg\(^{2+}\) values may result in decreased NO levels (80, 392).

The first reports of Mg\(^{2+}\) treatment for migraine patients appeared in the 1960s and 1970s (542), and since then the pharmacological role of Mg\(^{2+}\) has been slowly recognized. Although presently the effectiveness of Mg\(^{2+}\) treatment for the majority of patients is still debated (334, 386), Mg\(^{2+}\) is a second line drug for migraine patients (148, 237). Over the last decades several double-blind placebo-controlled randomized trials have provided evidence for a beneficial effect of oral Mg\(^{2+}\) supplementation on the number of migraine attacks (294, 393, 507) as well as the intensity of the pain during these attacks (149, 294). One clinical trial failed to show any favorable effect (397), but patients in this study suffered from diarrhea. Although Mg\(^{2+}\) is generally well tolerated, supplementation with certain Mg\(^{2+}\) salts often results in diarrhea and malabsorption of the Mg\(^{2+}\). Intravenous Mg\(^{2+}\) supplementation in acute migraine and cluster headache treatment provided pain relief in several cases (47, 112, 332, 333). In contrast, other studies failed to demonstrate such an effect (78). A recent meta-analysis combining five studies on the effect of intravenous Mg\(^{2+}\) administration on migraine did not show significant improvements on pain relief [Risk ratio (RR): −0.07, 95% confidence interval (CI): −0.23 to 0.09]. However, the relatively low number of patients included (n = 295) is one of the major limits of this analysis (86). Efficacy of Mg\(^{2+}\) treatment in combination with other drugs is doubtful; Mg\(^{2+}\) combined with metoclopramide and riboflavin did not demonstrate any efficacy in treating migraine (94, 328).

2. Depression

In 1921, Weston (557) reported the beneficial role of Mg\(^{2+}\) in treatment of patients with depression. Nevertheless, large-scale, placebo-controlled double blind clinical trials assessing the efficacy of Mg\(^{2+}\) supplementation on depression are still lacking. Studies examining the association between serum Mg\(^{2+}\) concentrations and depression severity are not conclusive; some studies report altered blood Mg\(^{2+}\) levels, while others do not find differences (reviewed in Ref. 114). Since serum Mg\(^{2+}\) levels do not necessarily reflect neuronal Mg\(^{2+}\) availability, determining Mg\(^{2+}\) levels in CSF may be more relevant for patients with depression. Only three cross-sectional studies have addressed this issue, and none of them found altered CSF Mg\(^{2+}\) concentrations in depressed patients compared with healthy controls (35, 171, 310). Despite this, several investigations have proposed that Mg\(^{2+}\) may relieve depression by blocking the NMDA receptor, whose dysfunction is a major causative factor in depression pathology (135, 143, 400, 557). To date, two interventional studies have investigated the role of Mg\(^{2+}\) in treating depression and their results are contradictory. In a randomized trial examining depressed elderly patients with diabetes mellitus type 2 and hypomagnesemia, Mg\(^{2+}\) supplementation was as effective as standard imipramine treatment (37). However, this study lacked a placebo control group. Moreover, it should be noted that imipramine therapy is nowadays largely replaced by selective serotonin reuptake inhibitors (SSRI). Another limit of the studies is the small sample population (23 patients). Therefore, large-scale studies are necessary to delineate a role of Mg\(^{2+}\) in the treatment and prevention of depression.

3. Epilepsy

Seizures are often associated with genetic and acquired forms of hypomagnesemia (see sect. V). Many studies have found that patients suffering from epilepsy display lower blood Mg\(^{2+}\) values (212, 375, 482). The link between Mg\(^{2+}\) status and the development of seizures may be explained by the role of Mg\(^{2+}\) in NMDA receptor blockade. Most studies addressing this issue show a small but significant decrease in CSF Mg\(^{2+}\) levels in epilepsy patients (492). In eclampsia patients, Mg\(^{2+}\) has proven to be successful in reducing the risk of recurrent convulsions (198). For other types of seizures, the evidence is less conclusive. In 1933 the first report of Mg\(^{2+}\) infusions in eight status epilepticus patients, a life-threatening form of epilepsy in which patients suffer from continuous seizures without regaining consciousness, was successful in all cases (496). However, modern reports of Mg\(^{2+}\) infusion treatment of status epilepticus patients have more variable outcomes and are not as conclusive (159, 383). Mg\(^{2+}\) infusions are therefore considered a second line of treatment, when anti-epileptic drugs and anesthetics have proven to be unsuccessful. Absence of large-scale randomized double blind placebo-controlled tri-
als obstructs the implementation of Mg$^{2+}$ as general anti-epileptic treatment (9).

4. Stroke

Stroke is one of the major causes of death in the Western society and has been associated with a drop in serum Mg$^{2+}$ levels (19). There may be multiple roles for Mg$^{2+}$ in the etiology of stroke. Low serum Mg$^{2+}$ levels increase NMDA receptor activity and thus more glutamate and Ca$^{2+}$ influx. Excessive Ca$^{2+}$ and glutamate influx via the NMDA receptor may be the basis of excitotoxicity during stroke (137).

Since clinical trials with NMDA receptor antagonists have proven to be unsuccessful in treatment of stroke (256), it is, however, unlikely that NMDA receptor blockade alone can fully explain the role of Mg$^{2+}$ in the development and onset of stroke. Mg$^{2+}$ also blocks other voltage-gated Ca$^{2+}$ channels that may be involved in Ca$^{2+}$ cytotoxicity. Additionally, Mg$^{2+}$ has a vasodilatory effect, which may be beneficial for patients suffering from ischemic stroke. Although more than 100 neuroprotective agents were tested in animals, not a single agent has been proven successful in a phase 3 clinical trial (111). After several pilot studies showed beneficial effects of Mg$^{2+}$ on clinical outcome parameters, two large randomized controlled trials have been performed to determine the role of Mg$^{2+}$ administration in stroke treatment (355, 356, 449). The Intravenous Magnesium Efficacy in Stroke Trial (IMAGES) enrolled over 2,500 stroke patients and gave a 16 mmol MgSO$_4$ bolus injection within 12 h of a stroke, followed by a maintenance dose of 65 mmol over 24 h. No beneficial effects were reported on the primary outcome, death and disability at 3 mo [odds ratio (OR) 0.95, 95% CI 0.80–1.13]. However, in a subgroup of patients treated within the first 3 h (3.3% of the cohort), a favorable death or disability outcome of 0.66 (95% CI 0.25–1.70) was reported (356). Since animal studies indicate that Mg$^{2+}$ treatment is only successful when applied within 3 h of the onset of the stroke (579), a large-scale follow-up randomized controlled trial is currently running which aims to treat patients with Mg$^{2+}$ within the first hours after stroke in a prehospital and emergency department setting. Although the trial is still running, first reports indicate that they have succeeded in including 72% of ~1,000 patients within the first hour (448). Final results from this FAST-MAG study will help determine the efficiency of Mg$^{2+}$ treatment for stroke patients.

Approximately 5% of all strokes are caused by subarachnoid hemorrhage (SAH) that results from ruptured aneurysms. Delayed cerebral ischemia (DCI) is the major cause of death and disability in patients that survive the first 24 h (126). Interestingly, patients with SAH often present with hypomagnesemia (531, 532). Vasoconstriction is the main cause of DCI, and this may be enhanced when the patient is Mg$^{2+}$ deficient. Over the last decade, several clinical trials have examined the addition of Mg$^{2+}$ administration to the standard nimodipine treatment (358, 533, 535, 568). In 2005, in the Magnesium and Acetylsalicylic acid in Subarachnoid Hemorrhage (MASH-I) study poor outcomes were 23% reduced (RR 0.77; 95% CI 0.54–1.09) and DCI was reduced by 35% (RR 0.65; 95% CI 0.40–1.05) (533). However, more recent studies in larger study-cohorts did not report beneficial effects. In the intravenous magnesium sulfate for aneurysmal subarachnoid hemorrhage (IMASH) double-blind randomized placebo-controlled trial, 6 mo favorable outcome was similar between the patients given MgSO$_4$ and the control, 64 and 63%, respectively (RR 1.0; 95% CI 0.7–1.6) (568). Comparable results were obtained more recently, in the MASH-II multicenter randomized placebo-controlled in a cohort of 1,200 patients; 74% had a favorable outcome in the Mg$^{2+}$ group and 75% in the control group (RR 1.03; 95% CI 0.85–1.25) (124). Overall, these studies suggest that intravenous Mg$^{2+}$ treatment does not improve clinical outcome after aneurysmal subarachnoid hemorrhage.

5. Brain injury

Mg$^{2+}$ deficiency is regularly found in patients with traumatic brain injury (TBI) and spinal cord injury (SCI) (81, 268). Reduced CSF Mg$^{2+}$ levels increase oxidative stress (ROS, NO) and lipid peroxidation, which both contribute to the severity of TBI (77, 416). Additionally, it has been proposed that Mg$^{2+}$ deficiency increases the release of substance P in TBI, resulting in neuronal cell death and edema (539). In animal experiments, Mg$^{2+}$ improved sensorimotor function as well as cognitive function (233). In a small study of 30 TBI patients, Mg$^{2+}$ supplementation improved patient outcomes as measured by the Glasgow outcome scale (OR 4.13; 95% CI 1.39–12.27) (116). A large phase 3 randomized placebo-controlled clinical trial tested the effect of two doses of Mg$^{2+}$ treatment on 6 mo mortality, seizures, functional measures, and neuropsychological tests in 500 TBI patients. Surprisingly, patients receiving Mg$^{2+}$ treatment did significantly perform worse on the primary outcome than control patients (48 vs. 34; 95% CI −10.5 to −2), suggesting an adverse effect of Mg$^{2+}$ (511). A meta-analysis on all clinical trials with TBI patients confirmed that there is no evidence for a neuroprotective role of Mg$^{2+}$ in TBI (25). This demonstrates again the difficulties in translating the results obtained in animal studies to the clinic (111).

6. Parkinson’s disease

Parkinson’s disease is characterized by a loss of dopaminergic neurons. Parkinson’s patients have low Mg$^{2+}$ concentrations in cortex, white matter, basal ganglia, and brain stem (580). Interestingly, rats with chronic low Mg$^{2+}$ intake exhibit a significant loss of dopaminergic neurons (381). In vitro experiments often use differentiated PC12 cells and 1-methyl-4-phenylpyridium ion (MPP$^+$) to model Parkinson’s disease at the cellular level. In this experimental
model, mitochondrial Mg\(^{2+}\) concentrations were decreased as was demonstrated using the mitochondrial KMG-301 fluorescent Mg\(^{2+}\) probe (475). Moreover, Mg\(^{2+}\) transporter SLC41A1 is located on the PARK16 locus that is associated with Parkinson’s disease (576). Recent characterization of the SLC41A1-pA350V single nucleotide polymorphism (SNP) linked to Parkinson’s disease evidenced a gain-of-function effect (290). These studies suggest that Mg\(^{2+}\) supplementation may be beneficial for patients suffering from Parkinson’s disease.

7. Other brain pathologies

Low serum Mg\(^{2+}\) levels have also been associated with a wide range of neurological pathologies including schizophrenia, bipolar disorder, neuroses, addiction, stress, and Alzheimer’s disease (538). Although this suggests that Mg\(^{2+}\) deficiency plays a role in the etiology of these pathologies, all reports to date are of an epidemiological nature. There are currently no reports of clinical trials examining the effect of Mg\(^{2+}\) supplementation on the disease outcome of these diseases.

B. Magnesium in Lung

Dietary Mg\(^{2+}\) intake has been repeatedly associated with lung function, as assessed by forced expiratory volume (FEV) and forced vital capacity (FVC) (60, 174). Lung Mg\(^{2+}\) research suffers from a lack of fundamental studies. As a result, the mechanisms that explain the role of Mg\(^{2+}\) in lung function are poorly understood, and hypotheses are mainly based on studies in other cell types and organs. Nevertheless, the role of Mg\(^{2+}\) in lung function may be explained at three levels: 1) Mg\(^{2+}\) has a strong vasodilator and bronchodilator effect; 2) Mg\(^{2+}\) regulates the release of acetylcholine (ACh) and histamine; and 3) Mg\(^{2+}\) acts as anti-inflammatory agent (FIGURE 7).

From studies on coronary artery related diseases, it is known that Mg\(^{2+}\) has a vasodilatory effect (512, 513). Like many vasodilators, Mg\(^{2+}\) also has a bronchodilating effect (229, 374). Although the mechanisms underlying Mg\(^{2+}\) induced bronchodilation remain to be elucidated, Mg\(^{2+}\) is known to inhibit the release of ACh and histamine, both known to induce bronchoconstriction (88, 291, 398, 427). Moreover, Mg\(^{2+}\) may reduce the airway inflammation that underlies several lung diseases, including chronic obstructive pulmonary disorder (COPD) and cystic fibrosis. In line with this, Mg\(^{2+}\) deficiency has been reported in children with bronchitis, and low Mg\(^{2+}\) levels can induce an inflammatory response in lung allografts (43, 440). Most of our understanding of the role of Mg\(^{2+}\) in inflammation comes from studies in brain, heart, and intestine (450, 509, 553), and only a few studies have examined the anti-inflammar-
tory function of \( \text{Mg}^{2+} \) in lung. Nevertheless, it is generally accepted that \( \text{Mg}^{2+} \) protects against inflammation by reducing oxidative stress, inhibiting substance P release, and preventing \( \text{Ca}^{2+} \) toxicity by inhibiting voltage-gated \( \text{Ca}^{2+} \) channels (450, 553). \( \text{Mg}^{2+} \) also modulates NF\( \kappa \)B activation by influencing lipid peroxidation (17, 153). All together, these characteristics make \( \text{Mg}^{2+} \) a potential therapeutic agent for lung diseases such as asthma and COPD.

1. Asthma

Several studies have reported low serum \( \text{Mg}^{2+} \) levels or low erythrocyte \( \text{Mg}^{2+} \) levels in asthmatic patients (21, 141, 219). However, others could not detect a \( \text{Mg}^{2+} \) deficiency in patients with asthma, suggesting that \( \text{Mg}^{2+} \) levels may depend on the severity of the disease (109, 276). Since \( \text{Mg}^{2+} \) relaxes smooth muscle cells, low \( \text{Mg}^{2+} \) levels cause bronchoconstriction and vasoconstriction, resulting in more asthmatic exacerbations (374). Moreover, \( \text{Mg}^{2+} \) regulates the release of \( \text{ACh} \) and histamine, which have both been implicated in asthma (170, 371). Interestingly, asthmatic C57/Bl6 mice have lower serum and intracellular \( \text{Mg}^{2+} \) concentrations than controls, which could be explained by a decreased renal TRPM6 expression (265). In 1940, Victor Haury (221) was the first to treat bronchial asthma patients with \( \text{Mg}^{2+} \) injections to relieve asthmatic paroxysms. Since then, –25 randomized controlled studies have been published examining the effects of nebulized and intravenous \( \text{Mg}^{2+} \) administration in asthma patients. A recent systematic review failed to demonstrate significant improvement of respiratory function [standardized mean difference (SMD) 0.17, 95% CI –0.02 to 0.36] or hospital admissions (RR 0.68, 95% CI 0.46 to 1.02) for patients with acute asthma who had been given nebulized \( \text{Mg}^{2+} \), although both parameters almost reached statistical significance (348). In studies using intravenous \( \text{Mg}^{2+} \) injection, respiratory function increased slightly in adults, but the most significant improvements were found in children (SMD 1.94, 95% CI 0.80–3.08), and children’s hospital admissions were reduced (RR 0.70, 95% CI 0.54–0.90) (348). A recent systematic review within the Cochrane collaboration addressing nebulized \( \text{Mg}^{2+} \) for treatment of acute asthma concluded that respiratory function is not significantly improved in \( \text{Mg}^{2+} \)-treated patients compared with patients receiving \( \beta_2 \)-agonists (404). However, the patients covered by this systematic review were mainly adult patients. Recently, the outcomes of the MAGNesium Trial In Children (MAGNETIC) randomized controlled study were published, showing improvement of the asthma severity score at 60 min (0.25, 95% CI 0.02–0.48) and 240 min (0.20, 95% CI 0.01–0.40) after inhalation of \( \text{MgSO}_4 \) (405). The MAGNETIC study was not published at the time of the systematic reviews, and future analysis including this trial may further substantiate the beneficial effects of \( \text{Mg}^{2+} \) in treatment of children with asthma.

2. Cystic fibrosis

\( \text{Mg}^{2+} \) deficiency has been repeatedly reported in patients with cystic fibrosis (210, 380). Patients with cystic fibrosis are often treated with recombinant human DNase-I to degrade the viscous mucus. However, the recombinant DNase requires \( \text{Mg}^{2+} \) to function, and efficiency of the treatment depends, therefore, on the \( \text{Mg}^{2+} \) status of the patient (444). It was suggested that \( \text{Mg}^{2+} \) treatment in itself would be sufficient to trigger endogenous DNase activity in the sputum (426). To further examine the potential role of \( \text{Mg}^{2+} \) as treatment for cystic fibrosis patients, a small double-blind, randomized, placebo-controlled crossover study tested the effect of oral \( \text{Mg}^{2+} \) supplementation on respiratory muscle strength and disease severity (444). Patients supplemented with \( \text{Mg}^{2+} \) increased their maximal inspiratory pressure (MIP) and maximal expiratory pressure (MEP). Although these first results are promising, more large-scale follow-up studies are necessary to assess the effects of \( \text{Mg}^{2+} \) in cystic fibrosis patients.

3. COPD

Serum \( \text{Mg}^{2+} \) levels have been associated with disease progression in patients with COPD (31, 46). Given the bronchodilating effects of \( \text{Mg}^{2+} \), several studies have examined whether intravenous or nebulized \( \text{Mg}^{2+} \) may benefit COPD patients. In 1995, a small study of 27 COPD patients examined the effect of 1.2 g \( \text{MgSO}_4 \) infusion after \( \beta_2 \)-agonist administration. Although the sample size was small, \( \text{Mg}^{2+} \)-treated patients demonstrated higher peak expiratory flow values compared with placebo-treated patients (22 ± 29 vs. 6 ± 24%) (485). In another small randomized double-blind controlled trial, \( \text{Mg}^{2+} \) infusion resulted in improved functional respiratory capacity (–0.48 l, 95% CI: –0.96, –0.01), inspiratory capacity (0.21 l, 95% CI: 0.04–0.37), MIP (10 cmH\(_2\)O, 95% CI: 1.6–18.4), and MEP (10.7 cmH\(_2\)O, 95% CI: 0.20–21.2) (123). In contrast, a recent study using combined intravenous and nebulized \( \text{MgSO}_4 \) administration in 62 COPD patients did not detect significant effects on the primary outcome, as measured by hospital admission, intubation, and hospital death rates, nor did they find improved lung function (370). A larger recent study testing nebulized \( \text{MgSO}_4 \) did not detect improved lung function after 90 min, as determined by FEV\(_1\) (136). Patients that received 151 mg \( \text{MgSO}_4 \) in addition to standard salbutamol treatment demonstrated similar FEV\(_1\) (–0.0261, 95% CI –0.15 to 0.095). Altogether, the efficacy of \( \text{Mg}^{2+} \) treatment in COPD remains unclear and may depend on the route of administration and the combination with the use of additional drugs.

C. Magnesium in Heart and Vasculature

\( \text{Mg}^{2+} \) plays an important role in heart function by influencing myocardial metabolism, \( \text{Ca}^{2+} \) homeostasis, vascular
tissue, peripheral vascular resistance, and cardiac output. Mg\(^{2+}\) exerts its effects in three ways: 1) Mg\(^{2+}\) regulates the activity of ion channels in the cardiac cells, thereby affecting the electrical properties of the myocardium (354); 2) Mg\(^{2+}\) regulates myocardial contractility by influencing the intracellular Ca\(^{2+}\) mobility; and 3) Mg\(^{2+}\) has an anti-inflammatory and vasodilatory effect (FIGURE 8).

The cardiac action potential consists of five phases: phase 0 is the rapid depolarization by the influx of Na\(^{+}\). Phase 1 consists of rapid repolarization by efflux of K\(^{+}\). Phase 2, named the plateau phase, is the longest phase and marks Ca\(^{2+}\) entry. Phase 3 allows final repolarization of the cell by restoration of the membrane potential. Phase 4 is the stable phase with a resting potential of ±90 mV (190). Mg\(^{2+}\) is mainly important in phases 2 and 3 of the myocardial action potential, exerting its effect on K\(^{+}\) and Ca\(^{2+}\) channels. In phase 2, Mg\(^{2+}\) inhibits L-type Ca\(^{2+}\) channels (Ca\(_v1.2\)) to prevent Ca\(^{2+}\) overload and cell toxicity (558). Mg\(^{2+}\) can bind a COOH-terminal EF hand motif of the channel and thereby influences the Ca\(^{2+}\) current (63). The effects of Mg\(^{2+}\) on the current through the L-type Ca\(^{2+}\) channels (\(I_{CaL}\)) may depend on the channel’s phosphorylation state, since phosphatase treatment decreases the inhibitory effects of Mg\(^{2+}\) (547). In phase 3, delayed rectifier K\(^{+}\) channels repolarize the cell by rapid-activating (\(I_{Kr}\)) and slow-activating (\(I_{Ks}\)) currents. High [Mg\(^{2+}\)] inhibit \(I_K\) currents in frog and guinea pig cardiomyocytes (128, 563). This effect probably depends on the slow-activating component of the current, since rapid-activating currents seem insensitive to

**FIGURE 8.** Magnesium in heart. Mg\(^{2+}\) influences phase 2 and phase 3 of the cardiac action potential by inhibiting L-type Ca\(^{2+}\) channels (phase 2) and delayed rectifier K currents (phase 3). Moreover, Mg\(^{2+}\) directly influences the cardiac muscle contraction by antagonizing Ca\(^{2+}\) binding of troponin C and calmodulin. It further modifies Ca\(^{2+}\) availability by affecting NCX and SERCA activity. Inset: the cardiac action potential. The numbers indicate the phases of the action potential. Tc, troponin C; CaM, calmodulin; NCX, Na\(^{+}\)-Ca\(^{2+}\)-exchanger; SERCA, sarcoplasmic/endoplasmic reticulum Ca\(^{2+}\)-ATPase; RYR, ryanodine receptor.
Mg\(^{2+}\) inhibition (499). The intracellular block of inward rectifier K\(^{+}\) channels Kir2.1 and Kir2.2 by Mg\(^{2+}\) substantially influences phase 3 and phase 4 of the action potential (536). This block of the \(I_{K1}\) current is relieved by high extracellular K\(^{+}\) concentrations (56).

In recent years an increasing amount of attention has been directed to the role of \([\text{Mg}^{2+}]\) in cardiac excitation-contraction coupling (342). Mg\(^{2+}\) has often been considered as a natural Ca\(^{2+}\) antagonist, since it can compete with Ca\(^{2+}\) for binding sites in proteins and Ca\(^{2+}\) transporters (257). The effect of Mg\(^{2+}\) on cardiomyocytes is mainly explained by its role of Ca\(^{2+}\) mobilization. Mg\(^{2+}\) binds calmodulin, troponin C, and parvalbumin, and therefore a reduced [Mg\(^{2+}\)] may result in alterations in the unbound Ca\(^{2+}\) fraction (15). Mg\(^{2+}\) may also affect the main Ca\(^{2+}\)-transporting proteins in the cardiomyocytes. Mg\(^{2+}\) acts as substrate in a complex with ATP for cardiac Ca\(^{2+}\)-ATPases and alters the affinity of Na\(^{+}\)–Ca\(^{2+}\) exchanger type 1 for Ca\(^{2+}\) (NCX1) (55, 58, 342). There is a dearth of physiological studies on the effect of Mg\(^{2+}\) on NCX1 and SERCA activity, and available studies mainly rely on modeling and in vitro experiments. Nevertheless, tight regulation of [Mg\(^{2+}\)] in cardiac cells is necessary for optimal cardiac function. This is substantiated by the fact that high [Mg\(^{2+}\)] can cause cardiac arrest, and by the impressive capacity of cardiomyocytes to maintain constant [Mg\(^{2+}\)] (506, 521).

An important role of Mg\(^{2+}\) in heart and vasculature function is a substantial vasodilatory effect that has been reported in animal and human studies (18, 161, 512, 513). Although the results from animal studies suggest that Mg\(^{2+}\)-induced vasodilation is due to the regulation of NO synthesis, human studies show that it is independent of NO activity (512, 513). Moreover, Mg\(^{2+}\) deficiency promotes oxidative stress notably in endothelial cells (118, 561), resulting in increased reactive oxygen species (ROS) and cytotoxicity (590). In contrast, high Mg\(^{2+}\) in the cell increases eNOS activity and suppresses the synthesis of vasoconstrictor endothelin-1 (277, 327). In conditions of low Mg\(^{2+}\)-induced oxidative stress, the endothelium develops a state of permanent innamination, which is marked by increased NFKB activity (153). NFKB is the master regulator of transcription of cytokines and pro-inflammatory genes, including IL-1α. As a result of this local inflammation, the vessel wall will recruit monocytes and trigger the proliferation and migration of vascular smooth muscle cells. These processes are facilitated by the increased expression of matrix metalloproteases 2 and 9 in low Mg\(^{2+}\) conditions (153, 382). Eventually low Mg\(^{2+}\) concentrations may, therefore, result in atherosclerosis, vascular calcifications, or thrombosis.

1. Coronary artery disease

Over the last 20 years, an increasing number of studies have demonstrated that low serum Mg\(^{2+}\) levels and low Mg\(^{2+}\) intake are associated with an increased risk of coronary artery disease (CAD), atherosclerosis, and metabolic syndrome (8, 222, 316, 587). Low serum Mg\(^{2+}\) levels have been associated with a higher mortality risk in CAD patients (162). There may be several ways in which Mg\(^{2+}\) supplementation benefits patients with CAD. Since Mg\(^{2+}\) has a strong anti-inflammatory role, Mg\(^{2+}\) results in an improved lipid profile, reduced free oxygen radicals, and improved endothelial function (317, 490). Mg\(^{2+}\) prevents blood clotting by reducing platelet aggregation (437), and it has a strong vasodilator effect (512, 513). These properties make Mg\(^{2+}\) an important factor in the development and management of CAD. Mg\(^{2+}\) improves several aspects of vascular function in CAD. Reduced serum Mg\(^{2+}\) concentrations are associated with an increase in carotid intima-media thickness and risk for sudden cardiac death (218, 324, 391). In a randomized, double-blind, placebo-controlled study of 50 CAD patients, oral Mg\(^{2+}\) supplementation ameliorated endothelial function (472). Moreover, Mg\(^{2+}\) reduced platelet-induced thrombosis in a randomized prospective, double-blind, crossover, and placebo-controlled study in 42 CAD patients (471). Six months of Mg\(^{2+}\) supplementation increased maximal oxygen uptake (\(\text{VO}_{2\text{max}}\)) and left ventricular ejection fraction (LVEF) in 53 CAD patients (401). Taken together, these studies suggest that Mg\(^{2+}\) levels should be closely monitored in CAD patients and propose Mg\(^{2+}\) as potential drug to improve quality of life in CAD patients.

2. Myocardial infarction

In the 1970s, a pioneering paper by Abraham et al. (9) associated myocardial infarction with a significant drop in serum Mg\(^{2+}\). Indeed, low serum Mg\(^{2+}\) levels have been associated with an increased risk of acute myocardial infarction (AMI) (484). The role of Mg\(^{2+}\) in preventing myocardial infarction may be caused by relaxing endothelial and smooth muscle cells in the heart and vasculature (18, 161, 512). Moreover, heart rate variability is a risk factor for AMI and Mg\(^{2+}\) may prevent arrhythmias (474, 484). Several studies have addressed the effect of Mg\(^{2+}\) on myocardial infarction. In the 1980s, studies reported a 20% reduction of infarct size in Mg\(^{2+}\)-treated patients, and decreased mortality after Mg\(^{2+}\) infusion (352, 415). Several follow-up studies suggested that decreased rates of arrhythmias after infarction explain the lower mortality (76, 487). After these initial promising results, several clinical trials have addressed this subject. The Leicester Intravenous Magnesium Intervention Trial 2 (LIMIT-2) included 2,316 AMI patients and found 24% reduced mortality after 28 days in the Mg\(^{2+}\) group (95% CI: 1–43%) (569). Moreover, two smaller studies by Shechter and colleagues reported reduced mortality rates of 50 and 40% (469, 470). Contrary to these findings, in the fourth Infarct Survival and Magnesium in Coronaries (ISIS-4) study, a randomized factorial trial in 58,050 patients showed no beneficial effects of intravenous Mg\(^{2+}\) administration on survival (4). The results from the ISIS-4 study were called into question because
the statistical analysis did not reflect the heterogeneous studies used in the analysis, the low mortality in the placebo-group was indicative of a low-risk comparison group, and the late time-point of Mg²⁺ administration, namely, after and not during reperfusion, was at odds with animal data demonstrating the effectiveness of Mg²⁺ (463). However, also a second large-scale randomized double-blind study with 6,213 patients and an earlier time point of Mg²⁺ infusion (3.8 h compared with 8 h in ISIS-4) also failed to show a decrease on 30-day mortality rates (2). Indeed, a recent meta-analysis within the Cochrane collaboration concludes that there is no beneficial effect of Mg²⁺ on mortality in AMI patients (OR 0.99, 95% CI 0.94 to 1.04) (312). However, it should be noted that the ISIS-4 study provides 72% of the power in this analysis and that the Mg²⁺ doses used within the analyzed studies differ significantly. Nevertheless, current guidelines do not recommend Mg²⁺ administration in AMI patients.

3. Arrhythmia

In 1935, Dr. Zwillinger was the first to report an antiarrhythmic effect of Mg²⁺, and since then sporadic reports of patients treated with Mg²⁺ have appeared in the literature (595). However, the field suffers from a lack of large-scale randomized controlled trials, and therefore the exact clinical benefit of Mg²⁺ in treatment of arrhythmia remains to be determined. Mg²⁺ is known to have a function in regulating cardiac K⁺ and Ca²⁺ channels, so it affects the cardiac action potential. As a result, hypomagnesemia in itself has been proposed as a cause of arrhythmia, specifically in combination with stress or alcoholism (362, 524). Clinical studies have demonstrated that treatment success strongly depends on the type of arrhythmia (232). Atrial fibrillation is one of the most common and dangerous complications after cardiac surgery. In the last few decades, several small-scale studies have examined the effect of Mg²⁺ in preventing these fibrillations (93). Meta-analysis of these studies concluded that Mg²⁺ infusion may prevent atrial fibrillations (201, 474). Therefore, the European Association for Cardiothoracic Surgery and the Canadian Cardiovascular Society recommend prophylaxis with intravenous MgSO₄. However, recent reports point to problems associated with many studies at the level of double-blind, primary outcome, and intention-to-treat analysis. When only high quality studies are included, Mg²⁺ does not have a preventive effect on atrial fibrillations (OR: 0.94, 95% CI: 0.61–1.44) (93). A similar discrepancy between low- and high-quality studies has been noted for studies addressing the effects of Mg²⁺ on supraventricular arrhythmias (108). Interestingly, for treatment of torsades des points, Mg²⁺ has been implemented as a first line of treatment after several studies in the 1980s showed beneficial effects (395, 527, 528). However, due to the absence of large-scale clinical trials, optimal doses for treatment are still under debate (211, 243, 244). Other arrhythmias, including refractory ventricular fibrillation and monomor-phic ventricular tachycardia, are insensitive to Mg²⁺ treatment (150, 220).

4. Preeclampsia

Since the 1950s, intravenous MgSO₄ administration has gradually become the standard treatment for preeclampsia and eclampsia, and nowadays the treatment is widely advocated by the World Health Organization (223, 407, 594). The mechanisms of action underlying the effect of Mg²⁺ in the treatment of these patients are largely unknown. Mg²⁺ may reduce preeclampsia by its effect as vasodilator in the vasculature, but it cannot be excluded that Mg²⁺ also functions as an anticonvulsant through blockade of the NMDA receptor and reduces cerebral edema (147). Recent Cochrane systematic reviews showed that MgSO₄ treatment in preeclamptic women reduced the risk for eclampsia by >50% (RR 0.41, 95% CI 0.29–0.58), and there was a trend towards lower maternal mortality (RR 0.54, 95% CI 0.26–1.10) (130). MgSO₄ demonstrated similar ratios for reduced risk for eclamptic convulsions comparable to anticonvulsant medication, with 59% compared with diazepam (RR 0.41, 95% CI 0.29–0.58) and 66% compared with phenytoin (RR 0.34, 95% CI 0.24–0.49) (131, 132). Moreover, a recent meta-analysis of “real-world” use of MgSO₄ for the treatment of preeclampsia confirmed the results from the clinical trials and showed ~50% reduction of eclampsia risk in most studies (337). Interestingly, although Mg²⁺ was successful in preventing eclampsia, it did not change the risk of death or disability for children at 18 mo (RR 1.06, 95% CI 0.90–1.25) or the risk of death or disability for women at 2 yr (RR 0.84, 95% CI 0.60–1.18) (5, 6).

5. Hypertension

Low serum Mg²⁺ levels are frequently linked to high blood pressure (30, 267, 270). Intracellular Mg²⁺ may reduce the intracellular Ca²⁺ concentration within vascular smooth muscle cells. This Mg²⁺-induced vasodilation is thought to be the mechanism by which Mg²⁺ alters the blood pressure. Moreover, high extracellular Mg²⁺ reduces the endothelin-1 expression and causes an increase in prostacyclin (PGI₂) levels, contributing to vasodilation (27, 59, 303, 447). Additionally, Mg²⁺ inhibits the production of NO (392). Mg²⁺ was first used to lower blood pressure in 1925, when Mg²⁺ infusion was found to lower blood pressure by reducing the peripheral vascular resistance (53). Several studies have shown that oral Mg²⁺ intake reduces both systolic and diastolic blood pressure (260, 275, 559); however, other studies fail to see such an effect (72, 152). A systematic review of the Cochrane Hypertension Group reported a small reduction of diastolic blood pressure (DBP; −2.2 mmHg, 95% CI −3.4 to −0.9), but not of the systolic blood pressure (SBP; −1.3 mmHg, 95% CI −4.0 to 1.5) (119). Another meta-analysis detected a small but dose-
dependent effect of Mg$^{2+}$ on blood pressure, for each 10 mmol/day – 4.3 mmHg SBP (95% CI – 6.3 to –2.2) and –2.3 mmHg DBP (95% CI – 4.9 to 0.0) (262). However, the outcomes may be biased by the inclusion of poor quality studies that tend to overestimate the effects. In contrast, a meta-analysis of a subset of studies, including patients on antihypertensive drugs with high blood pressure (SBP > 155 mmHg) reports much stronger effects of oral Mg$^{2+}$ treatment on SBP (–18.7 mmHg, 95% CI –14.95 to –22.45) and DBP (–10.9 mmHg, 95% CI – 8.73 to –13.1) (425). These results suggest that Mg$^{2+}$ may be beneficial for certain subgroups of hypertensive patients and that comparing the highly heterogeneous studies may underestimate the effects of Mg$^{2+}$ in these groups.

6. Vascular calcification

Vascular calcification is frequently observed in patients suffering from chronic kidney disease (CKD) (452). Calcifications are a major contributor to cardiovascular death that accounts for 50% of all deaths in CKD (522). Vascular calcification is the consequence of a disturbed mineral metabolism including increased serum P,$^{1}$ levels. Increased serum levels of FGF23 and PTH enhance the formation of calcification. Low serum Mg$^{2+}$ levels are associated with vascular calcification, and hemodialysis patients with higher serum Mg$^{2+}$ levels show higher survival (258, 259). Although the mechanisms of action are not completely understood, there are two contributing factors: 1) Mg$^{2+}$ prevents the formation and deposition of Ca/P nanocrystals and the development of appatite structures (83); and 2) Mg$^{2+}$ inhibits the transdifferentiation of the smooth muscle cells in the vessel wall into osteoblast-like cells (282). In both processes Mg$^{2+}$ prevents vascular calcification, and thus hypomagnesemic patients are at risk. Therefore, Mg$^{2+}$ supplementation has been proposed as P,$^{1}$-binder to reduce vascular calcification in CKD patients. Several studies have shown that combined administration of Ca$^{2+}$ and Mg$^{2+}$ is as effective as standard treatment options (252). In the recent CALMAG study comparing MgCO$_3$/Ca(OAc)$_2$ with Sevalamer-HCl in 200 hemodialysis patients, both treatments were effective in reducing serum P,$^{1}$ levels without increasing Ca$^{2+}$ levels (106).

D. Magnesium in Muscle

Mg$^{2+}$ mainly exerts its effects on skeletal muscle function as a Ca$^{2+}$-antagonist on Ca$^{2+}$-permeable channels and Ca$^{2+}$-binding proteins. Muscle contraction is a highly Ca$^{2+}$-dependent process, initiated Ca$^{2+}$ release from the sarcoplasmic reticulum. Ca$^{2+}$ binds to troponin C and myosin to induce the conformational changes of these proteins that will result in contraction (193). Mg$^{2+}$ competes for the Ca$^{2+}$-binding sites on these proteins. Although the affinity of troponin and myosin for Mg$^{2+}$ is much lower than for Ca$^{2+}$ (292), the effects of Mg$^{2+}$ are not negligible (239, 403). In the resting state, Mg$^{2+}$ is present in concentrations 10,000 times higher than Ca$^{2+}$ in muscle cells (292); therefore, Mg$^{2+}$ will occupy all Ca$^{2+}$ binding sites, and it is only after Ca$^{2+}$ is released from the sarcoplasmic reticulum that Mg$^{2+}$ is replaced. However, in Mg$^{2+}$-deficient conditions, not as much Ca$^{2+}$ is required to displace Mg$^{2+}$, resulting in hypercontractibility, which presents as muscle cramps and spasms in the clinic.

Moreover, as a cofactor of ATP, Mg$^{2+}$ availability is essential for the function of the ryanodine receptor (RyR), which allows rapid Ca$^{2+}$ release from the ER, and Ca$^{2+}$-ATPase of sarcoplasmic reticulum (SERCA), which mediates the return of Ca$^{2+}$ to the SR after contraction. Intracellular Mg$^{2+}$ and Ca$^{2+}$ concentrations determine how many ATP molecules bind to the RyR, which determines the opening of these channel receptors and the release of Ca$^{2+}$ (117). SERCA also depends on Mg$^{2+}$ as cofactor of ATP. Additionally, Mg$^{2+}$ can bind to the Ca$^{2+}$ binding pocket of SERCA (584). The role of Mg$^{2+}$ in the regulation of the Na$^{+}$-Ca$^{2+}$ exchanger (NCX) has been poorly studied so far.

1. Muscle cramps

Muscle cramps are a recurrent and prominent symptom in patients with severe/chronic hypomagnesemia (49, 215). Although the role of Mg$^{2+}$ in the pathogenesis of muscle cramps is not completely understood, it is hypothesized that Mg$^{2+}$ directly influences muscular contractions by antagonizing Ca$^{2+}$-binding proteins. Moreover, Mg$^{2+}$-deficient patients may suffer from neuronal hyperexcitability that can contribute to muscular contraction. Mg$^{2+}$ has consequently been considered as treatment for muscle cramps in several studies. However, there is little evidence that Mg$^{2+}$ may relieve muscle cramps in the general population. A recent Cochrane systematic review and meta-analysis of all published studies shows no significant reduction in the number of cramps after Mg$^{2+}$ treatment (–3.93%, 95% CI: –21.12 to 13.26%) (168). In contrast, a Cochrane review from 2002 demonstrated that Mg$^{2+}$ might be beneficial for muscle cramps in pregnancy (OR 0.18, 95% CI 0.05–0.60) (583). Both meta-analyses are limited by a relatively small patient population. Large-scale studies are necessary to ascertain the utility of Mg$^{2+}$ for specific subpopulations or disease-related muscle cramps.

E. Magnesium in Pancreas

Mg$^{2+}$ has been implicated in both endocrine and exocrine functioning of the pancreas. In the pancreatic acini, intracellular Mg$^{2+}$ antagonizes Ca$^{2+}$-activated signaling events and enzyme secretion (573). It is known that ACh and cholecystokinin 8 (CCK8) evoke an increase in intracellular Ca$^{2+}$, which in turn activates calmodulin, resulting in the phosphorylation of proteins on the enzyme-containing ves-
icles. These vesicles will migrate towards the plasma membrane for exocytosis and secretion. Interestingly, ACh and CCK8 activation cause a marked Mg\textsuperscript{2+} efflux, allowing Ca\textsuperscript{2+} signaling to occur (483, 364). Its antagonizing role may be explained by inhibition of Ca\textsuperscript{2+} transporting proteins such as SERCA, PMCA, and Ca\textsuperscript{2+}-ATPases (120, 584).

In the islets of Langerhans, Mg\textsuperscript{2+} may influence the secretion of insulin, although experimental results are conflicting as to whether insulin secretion is increased or decreased. Milner and Hales (344) reported that Mg\textsuperscript{2+} reduced insulin secretion in an ex vivo model using rabbit pancreas. These results were confirmed in rat pancreas and rat insulinoma cells (99, 357). However, in rats, one study found increased insulin secretion in Mg\textsuperscript{2+}-deficient rats, while other studies in Mg\textsuperscript{2+}-deficient rats did not report alterations in plasma insulin concentrations (202, 338, 418). In addition, patients with low serum Mg\textsuperscript{2+} levels show decreased insulin secretion (422). The discrepancies among different experiments may be explained by the Ca\textsuperscript{2+} availability in each of the models. Since the effect of Mg\textsuperscript{2+} on insulin can be explained by its Ca\textsuperscript{2+} antagonizing role, it may not depend on the Mg\textsuperscript{2+} concentration itself, but on the cytosolic Ca\textsuperscript{2+}/Mg\textsuperscript{2+} ratio.

1. Diabetes

Patients with diabetes mellitus type 2 often have low serum Mg\textsuperscript{2+} levels (36, 491, 541). These low serum levels are associated with poor disease outcome and may even increase mortality (98). Hypomagnesemia may contribute to the development of diabetes mellitus type 2 by increasing insulin resistance. Insulin receptors (IR) are part of the family of tyrosine kinase receptors, and the kinase function is dependent on the binding of two Mg\textsuperscript{2+} ions (249). Upon activation of the IR, a complex intracellular signaling cascade is activated and mediated via insulin receptor substrate proteins (505). In low Mg\textsuperscript{2+} conditions, activation of the IR may result in diminished signal transduction, contributing to insulin resistance. Studies with hypomagnesemic rats bear this out, as lower IR phosphorylation was detected, although differences between individual organs were reported (419, 498). It has also been proposed that increased expression of other effectors such as IL-1, IL-6, IL-8, TNF-α, norepinephrine, epinephrine, and ROS may contribute to insulin resistance in Mg\textsuperscript{2+} deficiency (206). Interestingly, common SNPs in the TRPM6 gene are associated with an increased risk of developing diabetes mellitus type 2 (489). TRPM6 cannot be activated by insulin when these SNPs are present (361). These results suggest that Mg\textsuperscript{2+} levels may influence the onset and development of diabetes mellitus type 2. Several studies have examined the clinical effects of oral Mg\textsuperscript{2+} supplementation on glycemic control in diabetes mellitus type 2 patients. Some of these studies demonstrate impressive effects in reducing glycated hemoglobin (HbA\textsubscript{1c}) levels and fasting glucose concentrations (385, 423), but other studies show no improvement of glycemic control (107, 204). A meta-analysis of 8 studies, including a total of 370 patients, evidenced a reduction of fasting glucose levels (–0.56 mM; 95% CI, –1.10 to –0.01), reflected in a nonsignificant (P = 0.1) reduction of HbA\textsubscript{1c} levels (–0.31%, 95% CI, –0.81 to 0.19) (488). All together, these results indicate that Mg\textsuperscript{2+} supplementation may be a promising avenue for achieving glycemic control in diabetes patients.

F. Magnesium in Liver

The role of Mg\textsuperscript{2+} in the liver is poorly studied. However, given that many of the enzymatic reactions that take place in the hepatocytes are dependent on Mg\textsuperscript{2+}, particularly in fat metabolism, the importance of Mg\textsuperscript{2+} should not be underestimated. Mg\textsuperscript{2+} supplementation has been reported to reduce alanine aminotransferase (ALT) levels in obese women with hypomagnesemia (421). However, this result could not be reproduced in normomagnesemic patients, although a lower dose of Mg\textsuperscript{2+} was used in this study (271). The first reports of hypomagnesemia in liver diseases such as cirrhosis and nonalcoholic fatty liver disease suggest that liver function contributes to proper intestinal Mg\textsuperscript{2+} absorption (286, 526). Currently, the first clinical trials are being initiated to test the effects of Mg\textsuperscript{2+} supplementation in patients with liver cirrhosis.

G. Magnesium in Immunity

Mg\textsuperscript{2+} is considered as an anti-inflammatory agent that reduces the expression and release of substance P and other proinflammatory molecules. Mg\textsuperscript{2+} also influences acquired immunity by regulating the proliferation and development of lymphocytes (156). Deletion of the TRPM7 Mg\textsuperscript{2+} channel caused cell death in the chicken B cell line DT40, which could be partially rescued by culturing the cells in high Mg\textsuperscript{2+} containing medium (455). In a mouse model with a specific T-cell deletion of TRPM7, T lymphocyte development was blocked at the CD4\textsuperscript{+}CD8\textsuperscript{−} stage, resulting in decreased CD4\textsuperscript{+} and CD4\textsuperscript{+}CD8\textsuperscript{+} cells in the thymus (266). Moreover, mutations in the MagT1 Mg\textsuperscript{2+} channel are causative for immunodeficiency and have been associated with decreased CD4\textsuperscript{+} T lymphocyte levels (311). These results suggest that Mg\textsuperscript{2+} is essential for T lymphocyte development and proliferation.

1. X-linked T-cell immunodeficiency

Patients with X-linked immunodeficiency with Mg\textsuperscript{2+} defect, Epstein-Barr virus infection and neoplasia (XMEN) have mutations in MagT1 (79, 311). They present with chronic Epstein-Barr virus infections, low CD4\textsuperscript{+} T-cell counts, and defective T-lymphocyte activation. These effects are hypothesized to result from a loss of PLCγ1 activation due to reduced Mg\textsuperscript{2+} influx via MagT1 (311). Recent studies in asthma pa-
Patients confirm the importance of Mg\(^{2+}\) availability for CD4\(^+\) function (315). An increased risk in T-cell lymphoblastic leukemia has been associated with Mg\(^{2+}\) deficiency (441). However, the role of Mg\(^{2+}\) in T-cell signaling needs to be investigated before further conclusions can be drawn. Special attention should be given to the involvement of other Mg\(^{2+}\) carriers, since TRPM7-deficient T cells seem protected from Fas-receptor-induced apoptosis (115).

H. Magnesium in Bone

Bone hydroxyapatite structures mainly consist of P\(_i\) and Ca\(^{2+}\) and are bound by Mg\(^{2+}\) ions at the surface of the hydroxyapatite crystals. Mg\(^{2+}\) increases the solubility of the minerals and thereby acts on the crystal size and formation (443). Crystals in Mg\(^{2+}\)-deficient bone are larger, and the bone may therefore be brittle and more susceptible to fractures (89). Moreover, Mg\(^{2+}\) stimulates osteoblast proliferation, suggesting that Mg\(^{2+}\) deficiency results in decreased bone formation (320) (FIGURE 4). The role of bone in Mg\(^{2+}\) homeostasis is described in more detail in section III B.

1. Osteoporosis

Several studies have associated low serum Mg\(^{2+}\) values with osteoporosis (61, 213). Although most studies have been performed in postmenopausal women, there is some evidence of low bone Mg\(^{2+}\) content in elderly subjects. In this study the patients had normal serum Mg\(^{2+}\) levels, but displayed significantly increased retention in a loading/tolerance test (91). A few small-scale studies have examined the effects of oral Mg\(^{2+}\) supplementation (200–750 mg/day) on bone mineral density (BMD) in patients with osteoporosis. In a pioneering study in 1991, daily administration of 600 mg Mg resulted in an 11% increased BMD after 12 mo (10). However, many other supplements including 500 mg/day Ca were simultaneously used, making it difficult to distinguish the effects of Mg\(^{2+}\). Subsequently, multiple studies have examined the effect of Mg\(^{2+}\) supplementation in different populations (138, 436, 494). Mg\(^{2+}\) seems to increase BMD in all of the studies, although the effects are relatively small (1–3%) and the small study sizes limit the conclusions that can be made from them. Testing the effect of Mg\(^{2+}\) supplementation in large cohorts of osteoporosis patients may further establish Mg\(^{2+}\) supplements to treat osteoporosis.

V. DISTURBANCES OF MAGNESIUM HOMEOSTASIS

Over the last decade, clinical interest in Mg\(^{2+}\) has been growing. Mg\(^{2+}\) deficiency has been associated with a wide range of diseases including diabetes mellitus type 2, hypertension, migraine, and depression. Therefore, the Mg\(^{2+}\) balance in patients is clinically significant; however, Mg\(^{2+}\) status in patients is not routinely determined. When Mg\(^{2+}\) is assessed in patients, total serum Mg\(^{2+}\) is the most commonly measured parameter. However, total serum Mg\(^{2+}\) levels do not necessarily represent body Mg\(^{2+}\) availability, since ionized Mg\(^{2+}\) determines the bioactive fraction. Only 2% of the clinical laboratories in the United States offer ionized Mg\(^{2+}\) tests, thus leaving total serum Mg\(^{2+}\) as the most used clinical representative of the patient’s Mg\(^{2+}\) status (139).

However, serum Mg\(^{2+}\) levels represent <1% of the body Mg\(^{2+}\) content, and therefore, it is a poor predictor of the body Mg\(^{2+}\) status. Additionally, the Mg\(^{2+}\) concentration in red blood cells is higher than in the serum (1.65–2.65 mM), and extra care should be taken to prevent hemolysis, which can result in a misrepresentation of total serum Mg\(^{2+}\) (520). In a recent systematic review, red blood cell (RBC) Mg\(^{2+}\) content has been proposed as an alternative measure to determine Mg\(^{2+}\) status, since its value is strongly affected by alterations in dietary Mg\(^{2+}\) intake. However, the rationale that alterations in dietary Mg\(^{2+}\) intake necessarily result in a physiologically relevant effect in Mg\(^{2+}\) availability is questionable, since the kidney and bone have a large capacity to compensate for reduced Mg\(^{2+}\) absorption. Only after a long-term depletion, patients may develop a clinically relevant hypomagnesemia.

However, the opposite also holds true; patients may be severely Mg\(^{2+}\) deficient, although serum or RBC Mg\(^{2+}\) levels are normal. In this case, the Mg\(^{2+}\) concentrations in bones and soft tissues are severely decreased, after long-term compensatory Mg\(^{2+}\) release to keep serum Mg\(^{2+}\) levels within normal range. To identify such patients, a Mg\(^{2+}\) loading test has been proposed in which acute oral Mg\(^{2+}\) administration is used to assess Mg\(^{2+}\) retention (363). In this test, the serum Mg\(^{2+}\) concentration and fecal Mg\(^{2+}\) excretion are calculated to determine intestinal Mg\(^{2+}\) absorption. Mg\(^{2+}\)-deficient patients will have lower bone Mg\(^{2+}\) content and therefore show high Mg\(^{2+}\) retention (90). Thus the loading test allows quantification of the exchangeable pool of Mg\(^{2+}\), which is more sensitive than serum Mg\(^{2+}\) concentrations. However, this test is rarely used in clinic, and as a result, standardization is lacking. Within a research setting, a urine excretion of <60% of the Mg\(^{2+}\) load is generally considered normal.

If a Mg\(^{2+}\) disturbance is suspected, urinary Mg\(^{2+}\) concentrations are regularly determined. It should be noted that reliable urine Mg\(^{2+}\) determinations require at least a complete 24-h sampling, since the circadian rhythm influences renal Mg\(^{2+}\) excretion (158). The results of urine Mg\(^{2+}\) tests may provide information about the cause of Mg\(^{2+}\) deficiencies; normal or high urinary excretion indicates renal Mg\(^{2+}\) wasting, whereas low Mg\(^{2+}\) excretion suggests reduced intestinal absorption. This information can then be used to guide treatment plans.
Importantly, physicians must be aware that urinary Mg\textsuperscript{2+} concentrations are poor indicators of a patient’s Mg\textsuperscript{2+} status when kidney function is reduced. In several pathophysiological circumstances such as diabetes and chronic kidney disease, filtration may be altered. Additionally, the use of certain drugs including diuretics may cause bias in determining a patient’s Mg\textsuperscript{2+} status.

A. Hypermagnesemia

Serum Mg\textsuperscript{2+} levels above 1.1 mM are generally considered hypermagnesemic. Hypermagnesemia may be clinically observed in patients suffering from nausea, vomiting, lethargy, headaches, and/or flushing. When Mg\textsuperscript{2+} levels rise above 3.0 mM it may even cause severe cardiac defects that are characterized by brachycardia, hypotension, and prolongation of the QRS, PR, and QT intervals (TABLE 3). Extreme hypermagnesemia can therefore result in coma, asystole, and death by cardiac arrest. However, hypermagnesemia is rare and until now no genetic causes for it have been identified. Hypermagnesemic patients are often treated by infusion of Ca\textsuperscript{2+} salts to antagonize the cardiac effects of Mg\textsuperscript{2+} (433). Also the efficacy of hemodialysis in the treatment of acute hypermagnesemia has been illustrated (228, 379).

**Table 3. Symptoms of Mg\textsuperscript{2+} disturbances**

<table>
<thead>
<tr>
<th>Concentration</th>
<th>Clinical Manifestation</th>
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<tbody>
<tr>
<td>&lt;0.7 mM</td>
<td>Neuromuscular irritability</td>
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<tr>
<td></td>
<td>Hypocalcemia</td>
</tr>
<tr>
<td>&lt;0.4 mM</td>
<td>Tetany</td>
</tr>
<tr>
<td></td>
<td>Seizures</td>
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<tr>
<td>≥0.4 mM</td>
<td>Arrhythmias</td>
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<tr>
<td>&gt;1.2 mM</td>
<td>Lethargy</td>
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<tr>
<td>&gt;2 mM</td>
<td>Drowsiness</td>
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<tr>
<td></td>
<td>Flushing</td>
</tr>
<tr>
<td></td>
<td>Nausea and vomiting</td>
</tr>
<tr>
<td>≥3 mM</td>
<td>Diminished deep tendon reflex</td>
</tr>
<tr>
<td></td>
<td>Somnolence</td>
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<tr>
<td>≥5 mM</td>
<td>Loss of deep tendon reflexes</td>
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<tr>
<td></td>
<td>Hypotension</td>
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<tr>
<td>≥10 mM</td>
<td>ECG changes</td>
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<tr>
<td></td>
<td>Complete heart block</td>
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<td></td>
<td>Cardiac arrest</td>
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<td>Paralysis</td>
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<td></td>
<td>Coma</td>
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<td>Death</td>
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</table>

1. Drug-induced hypermagnesemia

Only a few drug-induced cases of hypermagnesemia have been reported, and most of them are the direct consequence of the administration of Mg\textsuperscript{2+} itself or Mg\textsuperscript{2+}-containing drugs (TABLE 4).

a) EPSOM SALTS. Epsom salts consist of MgSO\textsubscript{4} and are generally used as bath salts or a home remedy against abdominal pain, constipation, and muscle strains. Excessive ingestion of Epsom salts may result in Mg\textsuperscript{2+} overdose and hypermagnesemia. As a result, several fatal cases have been reported in literature (52, 231).

b) CATHARTICS, LAXATIVES, AND ENEMA. Several Mg\textsuperscript{2+} derivatives have been used as cathartics, laxatives, and enemas with the similar goals of softening the stool and easing defecation by increasing their water content. For a long time magnesium citrate [Mg\textsubscript{3}(C\textsubscript{6}H\textsubscript{5}O\textsubscript{7})\textsubscript{2}] was the most commonly used cathartic. Due to the risk of developing hypermagnesemia and other electrolyte disturbances, polyethylene glycol and electrolyte lavage solutions are currently the first drugs of choice (378, 451). Several Mg\textsuperscript{2+}-containing substances [Mg\textsubscript{3}(C\textsubscript{6}H\textsubscript{5}O\textsubscript{7})\textsubscript{2}, Mg(OH)\textsubscript{2}, MgSO\textsubscript{4}] have been used as laxatives. Mg\textsuperscript{2+} substances elevate the intestinal osmotic pressure, but also act on aquaporin-3 expression and thus increase water permeability (373). Mg\textsuperscript{2+} has been used as a component of enemas to treat constipation. However, both use of Mg\textsuperscript{2+} as a laxative and as an enema may result in fatal hypermagnesemia (413, 521, 586). Therefore, Mg\textsuperscript{2+} administration should be avoided in patients with reduced kidney function, and serum Mg\textsuperscript{2+} should be closely monitored during treatment.

B. Hypomagnesemia

Hypomagnesemia is generally defined as serum Mg\textsuperscript{2+} levels below 0.7 mM. Patients suffer from nonspecific symptoms such as depression, tiredness, muscle spasms, and muscle weakness, and diagnosis therefore may take years (TABLE 3) (177, 523). Only severe Mg\textsuperscript{2+} depletion (<0.4 mM) may lead to cardiac arrhythmias, tetany, and seizures. Secondary to hypomagnesemia, disturbances in K\textsuperscript{+} and Ca\textsuperscript{2+} handling are often detected. Hypokalemia can be attributed to increased renal K\textsuperscript{+} secretion via ROMK in the connecting tubule (CNT) and collecting duct (CD) (248). Low intracellular Mg\textsuperscript{2+} levels release the Mg\textsuperscript{2+}-dependent inhibition of ROMK channels, resulting in increased renal K\textsuperscript{+} secretion. Hypokalemia can be explained by low PTH levels due to altered activation of the CaSR (574).

Hypomagnesemia is generally treated by oral Mg\textsuperscript{2+} supplementation (± 360 mg/day), although oral Mg\textsuperscript{2+} intake may cause diarrhea at high doses. Intravenous Mg\textsuperscript{2+} supplementation may be more effective, but this treatment has the disadvantage that it requires regular hospital visits. The
treatment regimen of intravenous Mg²⁺ supplementation normally consists of 8–12 g of magnesium sulfate in the first 24 h followed by 4–6 g/day for 3 or 4 days (523). When serum Mg²⁺ levels are extremely low or are accompanied by hypokalemia, Mg²⁺ supplementation may not be sufficient to restore normal Mg²⁺ levels. In that case, patients are often cosupplemented with K⁺ or receive amiloride to prevent K⁺ secretion.

The following section of this review will focus on the causes of hypomagnesemia. Drug-induced and genetic hypomagnesemia will be distinguished from more general origins of Mg²⁺ deficiency (TABLES 4 AND 5).

1. General causes of hypomagnesemia

A) DIETARY Mg²⁺ INTAKE. Estimations state that up to 60% of American do not meet daily Mg²⁺ requirements (163, 281). Chronic inadequate intake of Mg²⁺ leads to hypomagnesemia. Consumption of Mg²⁺-rich foods such as kelp, nuts, green vegetables, and whole grains may prevent this. However, insufficient Mg²⁺ intake may also be caused by pathological conditions such as anorexia nervosa (51).

B) VOMITING AND DIARRHEA. Vomiting and diarrhea may further exacerbate the effects of inadequate Mg²⁺ uptake. Specifically, diarrhea is the consequence of inadequate water reabsorption along the intestine. Since water reabsorption is a prerequisite for Mg²⁺ reabsorption to set the concentration gradient, diarrhea may result in Mg²⁺ deficiency.

C) ALCOHOLISM. Since the early 1960s it has been recognized that alcoholism may cause severe hypomagnesemia (224, 336). Patients suffer from unexplained renal Mg²⁺ wasting, but may also have reduced intestinal Mg²⁺ absorption due to vomiting or diarrhea. In hepatocytes, ethanol completely blocked Mg²⁺ uptake (75). A similar mechanism may take place in the kidney, explaining reduced Mg²⁺ reabsorption. Alcoholics often have reduced PTH levels, which may further contribute to low serum Mg²⁺ levels (7).

<table>
<thead>
<tr>
<th>Class</th>
<th>Drug</th>
<th>Mechanism</th>
<th>Prevention/Treatment</th>
<th>Reference Nos.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Drug-induced hypomagnesemia</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Diuretics</td>
<td>Furosemide</td>
<td>TAL: reduced paracellular Mg²⁺ reabsorption</td>
<td>Combined K⁺ and Mg²⁺ supplementation, Switch to K⁺ and Mg²⁺-sparing diuretics such as amiloride</td>
<td>92, 101, 127, 368</td>
</tr>
<tr>
<td></td>
<td>Thiazide</td>
<td>DCT: reduced TRPM6 expression</td>
<td>Mg²⁺ supplementation</td>
<td></td>
</tr>
<tr>
<td>EGFR inhibitors</td>
<td>Cetuximab</td>
<td>DCT: reduced TRPM6 activity</td>
<td>Mg²⁺ supplementation</td>
<td>341, 459, 510</td>
</tr>
<tr>
<td>Proton pump inhibitors</td>
<td>Omeprozole, lansoprazole, pantoprazole, rabeprazole, etc.</td>
<td>Intestine: reduced Mg²⁺ absorption</td>
<td>Mg²⁺ supplementation Switch to histamine2 receptor antagonists</td>
<td>144, 226</td>
</tr>
<tr>
<td>Calcinurin inhibitors</td>
<td>Cyclosporin A, tacrolimus</td>
<td>DCT: reduced TRPM6 expression</td>
<td>Mg²⁺ supplementation</td>
<td>39, 517</td>
</tr>
<tr>
<td>Platinum derivates</td>
<td>Cyclosporin A, tacrolimus</td>
<td>DCT: cell death? Reduced TRPM6 expression?</td>
<td>Mg²⁺ supplementation</td>
<td>142, 453, 495, 593</td>
</tr>
<tr>
<td>Antimicrobials</td>
<td>AGAs</td>
<td>TAL: reduced paracellular Mg²⁺ reabsorption</td>
<td>Mg²⁺ supplementation</td>
<td>23, 38, 169, 188, 535, 582, 585</td>
</tr>
<tr>
<td></td>
<td>Pentamidine</td>
<td>DCT: cell death?</td>
<td>Mg²⁺ supplementation</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Rapamycin</td>
<td>TAL: reduced paracellular Mg²⁺ reabsorption</td>
<td>Mg²⁺ supplementation</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Amphoteracin B</td>
<td>?</td>
<td>Mg²⁺ chelating</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Foscarinet</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Drug-induced hypermagnesemia</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Epsom salt poisoning</td>
<td>MgSO₄</td>
<td>Intestinal Mg²⁺ overload</td>
<td>Hemodialysis</td>
<td>52, 231</td>
</tr>
<tr>
<td>Cathartics</td>
<td>Mg₃(C₂H₃O₇)₄</td>
<td>Intestinal Mg²⁺ overload</td>
<td>Switch to polyethylene glycol or electrolyte lavage solutions</td>
<td>378, 451</td>
</tr>
<tr>
<td>Laxatives</td>
<td>MgSO₄, Mg(OH)₂, Mg₃(C₂H₃O₇)₄</td>
<td>Intestinal Mg²⁺ overload</td>
<td>Switch to bulk (fiber-based) laxatives</td>
<td>413, 521, 586</td>
</tr>
<tr>
<td>Enema</td>
<td>MgSO₄</td>
<td>Intestinal Mg²⁺ overload</td>
<td>Switch to fleet (sodium phosphate) enema</td>
<td></td>
</tr>
</tbody>
</table>

EGFR, epidermal growth factor receptor; TAL, thick ascending limb of Henle’s loop; DCT, distal convoluted tubule; TRPM6, transient receptor potential melastatin type 6; AGAs, aminoglycoside antibiotics.
2. Genetic hypomagnesemia

A) CLDN16. Familial hypomagnesemia with hypercalciuria and nephrocalcinosis type I (FHHNC type I; OMIM 248250) is caused by mutations in claudin 16, previously known as paracellin-1 (480). Patients suffer from renal Mg\(^{2+}\) wasting, hypomagnesemia, renal Ca\(^{2+}\) wasting, renal parenchymal calcification (nephrocalcinosis), and renal failure. Serum and urinary Na\(^{+}\), K\(^{+}\), Cl\(^{-}\), and HCO\(_3^{-}\) values are initially normal, but may be indirectly affected after progression of renal failure. Sometimes these symptoms are extended to urinary tract infections, kidney stones, and hyperuricemia; Mg\(^{2+}\) supplementation is not capable of restoring normal serum Mg\(^{2+}\) levels or slowing disease progression (567). A few dozen different mutations have been reported, all characterized by a recessive mode of inheri-

### Table 5. Genetic causes of hypomagnesemia

<table>
<thead>
<tr>
<th>Gene</th>
<th>Protein</th>
<th>Disease</th>
<th>OMIM</th>
<th>Inh.</th>
<th>Segment</th>
<th>Blood Mg(^{2+})</th>
<th>Urine Mg(^{2+})</th>
<th>Blood Ca(^{2+})</th>
<th>Urine Ca(^{2+})</th>
<th>Other Symptoms</th>
<th>Reference Nos.</th>
</tr>
</thead>
<tbody>
<tr>
<td>CLDN16</td>
<td>Claudin 16</td>
<td>FHHNC type 1</td>
<td>248250</td>
<td>R</td>
<td>TAL</td>
<td>↓</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>Nephrocalcinosis, renal failure</td>
<td>480</td>
</tr>
<tr>
<td>CLDN19</td>
<td>Claudin 19</td>
<td>FHHNC type 2</td>
<td>248190</td>
<td>R</td>
<td>TAL</td>
<td>↓</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>Nephrocalcinosis, renal failure, visual impairment</td>
<td>293</td>
</tr>
<tr>
<td>SLC12A1</td>
<td>NKCC2</td>
<td>Bartter type 1</td>
<td>601678</td>
<td>R</td>
<td>TAL</td>
<td>↓</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>Na(^{+}) wasting, hypokalemic alkalosis, high</td>
<td>478</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>renal failure</td>
<td></td>
</tr>
<tr>
<td>KCNJ1</td>
<td>ROMK</td>
<td>Bartter type 2</td>
<td>241200</td>
<td>R</td>
<td>TAL</td>
<td>↓</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>Na(^{+}) wasting, hypokalemic alkalosis, high</td>
<td>479</td>
</tr>
<tr>
<td>CLCNKB</td>
<td>CLC-Kb</td>
<td>Bartter type 3</td>
<td>607364</td>
<td>R</td>
<td>TAL</td>
<td>↓</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>Na(^{+}) wasting, hypokalemic alkalosis, high</td>
<td>477</td>
</tr>
<tr>
<td>BSND</td>
<td>Barttin</td>
<td>Bartter type 4</td>
<td>602522</td>
<td>R</td>
<td>TAL</td>
<td>↓</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>Na(^{+}) wasting, hypokalemic alkalosis, high</td>
<td>50</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>renal failure</td>
<td></td>
</tr>
</tbody>
</table>

**Human genetic Mg\(^{2+}\) disorders in TAL**

<table>
<thead>
<tr>
<th>Gene</th>
<th>Protein</th>
<th>Disease</th>
<th>OMIM</th>
<th>Inh.</th>
<th>Segment</th>
<th>Blood Mg(^{2+})</th>
<th>Urine Mg(^{2+})</th>
<th>Blood Ca(^{2+})</th>
<th>Urine Ca(^{2+})</th>
<th>Other Symptoms</th>
<th>Reference Nos.</th>
</tr>
</thead>
<tbody>
<tr>
<td>TRPM6</td>
<td>TRPM6</td>
<td>HSH</td>
<td>602014</td>
<td>R</td>
<td>DCT</td>
<td>↓</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>Seizures, muscle spasms, mental retardation</td>
<td>454, 543</td>
</tr>
<tr>
<td>EGF</td>
<td>EGF</td>
<td>IRH</td>
<td>611718</td>
<td>R</td>
<td>DCT</td>
<td>↓</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>Seizures, mental retardation</td>
<td>197</td>
</tr>
<tr>
<td>CNNM2</td>
<td>CNNM2</td>
<td>HSMR</td>
<td>613882</td>
<td>D/R</td>
<td>DCT</td>
<td>↓</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>Seizures, mental retardation</td>
<td>497</td>
</tr>
<tr>
<td>KCNJ10</td>
<td>Kir4.1</td>
<td>SeSAME/EAST</td>
<td>612780</td>
<td>R</td>
<td>DCT</td>
<td>↓</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>Hypokalemic, metabolic alkalosis, sensorineural</td>
<td>54, 458</td>
</tr>
<tr>
<td>FXYD2</td>
<td>FXYD2</td>
<td>IDH</td>
<td>154020</td>
<td>D</td>
<td>DCT</td>
<td>↓</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>Convulsions</td>
<td>339</td>
</tr>
<tr>
<td>HNF1B</td>
<td>HNF1β</td>
<td>RCAD</td>
<td>137920</td>
<td>D</td>
<td>DCT</td>
<td>↓</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>Renal cysts, MODY5, renal malformations</td>
<td>11</td>
</tr>
<tr>
<td>PCBD1</td>
<td>PCBD1</td>
<td>RCAD-like</td>
<td>264070</td>
<td>R</td>
<td>DCT</td>
<td>↓</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>Transient hyperphenylalaninemia, MODY5-like</td>
<td>154</td>
</tr>
<tr>
<td>SLC12A3</td>
<td>NCC</td>
<td>Gitelman syndrome</td>
<td>263800</td>
<td>R</td>
<td>DCT</td>
<td>↓</td>
<td>–</td>
<td>–</td>
<td>–</td>
<td>Hypokalemic, metabolic alkalosis, tetany, chondrocalcinosis</td>
<td>481</td>
</tr>
</tbody>
</table>

SLC, solute carrier; NKCC2, Na\(^{+}\)-K\(^{+}\)-2Cl\(^{-}\) cotransporter; TRPM6, transient receptor potential melastatin type 6; EGF, epidermal growth factor; CNNM2, cyclin M2; FXYD2, FXYD domain containing ion transport regulator 2; HNF1B, hepatocyte nuclear factor 1B; PCBD1, pterin-4-alpha-carbinolamine dehydratase; NCC, Na\(^{+}\)-Cl\(^{-}\) cotransporter; FHHNC, familial hypomagnesemia with hypercalciuria and nephrocalcinosis; HSH, hypomagnesemia with secondary hypocalcemia; IRH, isolated recessive hypomagnesemia; HSMR, hypomagnesemia with seizures and mental retardation; ADH, autosomal dominant hypomagnesemia; SeSAME, sensorineural deafness, seizures, ataxia, mental retardation, and electrolyte imbalance; EAST, epilepsy, ataxia, sensorineural deafness, and tubulopathy; IDH, isolated dominant hypomagnesemia; RCAD, renal cysts and diabetes; OMIM: online Mendelian inheritance in man; D, dominant; R, recessive; TAL, thick ascending limb of Henle’s loop; DCT, distal convoluted tubule; MODY, maturity-onset diabetes of the young.
tance (177). All symptoms can be traced to the TAL, the main site for paracellular Ca\textsuperscript{2+} and Mg\textsuperscript{2+} reabsorption. Claudin 16 is part of the tight junction between the cells, and disruption of these tight junctions results in a lack of Ca\textsuperscript{2+} and Mg\textsuperscript{2+} reabsorption in TAL, which can only partially be compensated in the downstream DCT and CNT segments.

B) CLDN19. Similar to FHHNC type I, patients with FHHNC type 2 (OMIM 248190) suffer from severe hypomagnesemia accompanied by hypercalciuria and nephrocalcinosis. Additionally, patients have ocular defects consisting of macular colobomata, significant myopia, and horizontal nystagmus. FHHNC type 2 is caused by mutations in claudin 19, which is expressed in the TAL segment of the kidney, in parallel with claudin 16. In the initial publication from Konrad et al. (293), 12 patients from 10 families were genotyped and characterized. Remarkably, 9 of 12 patients developed chronic kidney disease or underwent kidney transplantation. Indeed, other studies confirmed that FHHNC type 2 patients are more prone to developing of CKD and develop the disease at an earlier age compared with type I patients (177). Over the years, several treatment regimens have been proposed, including oral magnesium supplementation, thiazide diuretics, and indomethacin. However, none of these treatments significantly increased serum Mg\textsuperscript{2+} levels (177).

C) TRPM6. Hypomagnesemia with secondary hypocalcemia (HSH; OMIM 602014) is characterized by extremely low serum Mg\textsuperscript{2+} levels (0.1–0.3 mM) accompanied by low serum Ca\textsuperscript{2+} levels, which result in severe muscular and neurological complications including seizures and mental retardation (454, 543). The disorder was first characterized by Paunier and colleagues in 1968 and later mapped to a region at chromosome 9q in 1997 (390, 544). In 2002, two independent groups identified mutations in TRPM6 to be causative for HSH (454, 543). TRPM6 forms the epithelial Mg\textsuperscript{2+} channel responsible for transcellular Mg\textsuperscript{2+} transport in the colon and DCT segment of the kidney (540). Therefore, mutations result in reduced intestinal absorption and renal Mg\textsuperscript{2+} wasting. HSH has an autosomal-recessive mode of inheritance, and currently a few dozen mutations have been found. Patients are generally treated with Mg\textsuperscript{2+} supplements and antiepileptic drugs against seizures. Serum Mg\textsuperscript{2+} levels improve after supplementation, but did not recover to normal levels (297).

D) EGF. Isolated autosomal recessive hypomagnesemia (IRH; OMIM 611718) is caused by mutations in the EGF gene (197). In a consanguine family from Dutch origin, two sisters presented with serum Mg\textsuperscript{2+} levels of 0.53 and 0.56 mM and urinary Mg\textsuperscript{2+} values of 3.9 and 3.7 mmol/24 h, respectively (173). Serum Ca\textsuperscript{2+}, Na\textsuperscript{+}, K\textsuperscript{+}, Cl\textsuperscript{−}, HCO\textsubscript{3}−, and blood pH values were normal. The patients presented with epileptic seizures during their first year of life, which could be controlled by antiepileptic drugs. Moreover, psychomotor retardation was observed in these patients. Plasma renin activity, plasma aldosterone, and parathyroid hormone concentrations were in the normal range. Homozygosity mapping and subsequent Sanger sequencing of gene candidates led to the identification of a homozygous c.3209T mutation in exon 22 resulting a p.P1070L missense mutation at protein level (197). This residue is particularly important for plasma membrane targeting of the EGF molecule and the mutation results in impaired basolateral sorting of pro-EGF. Therefore, TRPM6 activity is not stimulated, resulting in renal Mg\textsuperscript{2+} wasting (197, 515). Until now, only a single family has been described with EGF mutations, but studies with EGFR antagonists further underline the clinical importance of EGF for renal Mg\textsuperscript{2+} handling (discussed in detail in sect. VB3).

E) KCNA1. In a Brazilian family, mutations in KCNA1 encoding voltage-gated K\textsuperscript{+} channel Kv1.1 cause autosomal dominant hypomagnesemia (OMIM 176260) (176). The patient presented in the clinic with muscle cramps, muscle, weakness, tetanic episodes, and tremor. Serum Mg\textsuperscript{2+} values were low (0.37 mM), whereas other electrolytes and metabolites including Na\textsuperscript{+}, K\textsuperscript{+}, Ca\textsuperscript{2+}, P\textsubscript{i}, uric acid, bicarbonate, urea, creatinine, glucose, bilirubin, aminotransferases, alkaline phosphate, and lactate dehydrogenase were all normal. Urinary creatinine clearance as well as Mg\textsuperscript{2+} and Ca\textsuperscript{2+} excretion were within normal range. KCNA1 mutations were previously linked to ataxia and myokymia (62, 146). Therefore, a cerebral MRI was performed in these patients, showing slight atrophy of the cerebral vermis. Family members suffer from myokymic discharge in electromyography analysis, which is in line with the previously observed mixed phenotype. Intravenous Mg\textsuperscript{2+} infusion improved the clinical symptoms. Kv1.1 has been proposed to cause apical hyperpolarization, which allows the uptake of Mg\textsuperscript{2+} via TRPM6. The p.N255D (c.A763G) mutation identified in the Brazilian family disrupts Kv1.1 activity and therefore associated with hypomagnesemia, even though Kv1.1 function is impaired (12, 263). Identification of additional hypomagnesemic families with Kv1.1 mutations may aid our understanding of Kv1.1 function in DCT. It has been suggested that other factors contribute to the apical membrane potential and may compensate for a loss of Kv1.1 function; ROMK may be one of the compensatory factors (104, 140).

F) CNNM2. Mutations in CNNM2 are causative for hypomagnesemia with seizures and mental retardation (HSMR; OMIM 613882). Two unrelated families with seizures and dominant hypomagnesemia were reported to carry CNNM2 mutations (497). In these patients, serum Mg\textsuperscript{2+} levels range between 0.3 and 0.5 mM, but no other electrolyte disturbances were detected. The patients’ symptoms
include seizures, loss of consciousness, loss of muscle tone, headaches, and staring (340). Recently, five additional families were reported (28), making CNNM2 the most common genetic cause of isolated hypomagnesemia after TRPM6 and CLDN16–19. Interestingly, in this new cohort CNNM2 mutations were linked to a phenotype of impaired brain development and mental retardation. This intellectual disability was most prominent in a family with a recessive pattern of inheritance, emphasizing the heterogeneous inheritance of CNNM2 depending on the location and severity of the mutations. HSMR patients are treated with antiepileptic drugs and Mg2+ supplements. Serum Mg2+ levels improved after supplementation, but did not reach normal levels. Although the exact function of CNNM2 remains to be elucidated, mutations can disrupt the MgATP binding domain and reduce CNNM2 membrane expression (28, 105). Recent data from patch-clamp and Mg2+ uptake experiments favor the hypothesis that CNNM2 does not transport Mg2+ itself, but rather regulates other Mg2+-transporting proteins (28, 105, 497).

G) KCNJ10. Mutations in KCNJ10 encoding the Kir4.1 K+ channel can cause seizures, sensorineural deafness, ataxia, mental retardation, and electrolyte imbalance/epilepsy, ataxia, sensorineural deafness, and renal tubulopathy syndrome (SeSAME/EAST; OMIM 612780) (54, 417, 458). Patients were reported to have marked electrolyte abnormalities, including hypokalemic metabolic alkalosis without hypertension, severe hypomagnesemia, and renal Na+, K+, and Mg2+ wasting. In some patients, high renin and aldosterone levels, salt craving, and polyuria were observed. Kir4.1 is hypothesized to be involved in K+ recycling at the basolateral membrane of DCT cells. When Kir4.1 is mutated, K+ availability becomes rate limiting for Na+-K+-ATPase activity. Thus the Na+-K+-ATPase will be inhibited, resulting in a reduced potential across the basolateral membrane. Consequently, Na+ and Mg2+ transport will be reduced in DCT. To compensate for this, ENaC activity in CNT will be increased at the expense of K+ excretion via ROMK. Therefore, SeSAME/EAST patients suffer from severe hypomagnesemia and hypokalemia. To treat the hypomagnesemia, patients are often given Mg2+ and K+ supplements, in combination with aldosterone antagonists or ENaC inhibitors (34). SeSAME/EAST patients suffer from a severe neurological phenotype consisting of tonic-clonic seizures in infancy, cerebellar ataxia, and hearing loss. Moreover, magnetic resonance imaging evidenced subtle symmetrical signal changes in the cerebellar dentate nuclei (96).

H) FXYD2. Gene linkage studies identified the FXYD domain containing ion transport regulator 2 (FXYD2) mutations in a family with dominant isolated renal Mg2+ wasting (IDH; OMIM 154020) (339). The patients in this family presented with low serum Mg2+ values (±0.4 mM), while other plasma electrolytes, including Na+, K+, Ca2+, Cl−, and HCO3−, were normal. Urinary Mg2+ excretion was increased, whereas Ca2+ excretion was slightly lowered (172). The c.G121A mutation results in a p.G41R missense mutation at the protein level, which causes misrouting of FXYD2 to the membrane (66). FXYD2 encodes for the γ-subunit of the Na+-K+–ATPase. Although the exact role of FXYD2 in the DCT is unknown, it has been hypothesized to stabilize the Na+-K+–ATPase, influencing the membrane potential necessary for Mg2+ transport (345). However, other reports suggest that it may function independently as an inward rectifier channel (464). Functional analysis of the patient’s proximal tubular cells showed no differences in Na+, K+, or ATP affinity of the Na+–K+–ATPase, but demonstrated a lower FXYD2 protein expression (67).

I) HNF1B. Renal cysts and diabetes syndrome (RCAD; OMIM 137920) is caused by mutations in hepatocyte nuclear factor 1β (HNF1β) and consists of a heterogeneous group of symptoms including renal cysts (±70% of patients), maturity onset diabetes of the young subtype 5 (MODY5; ±50%), and hypomagnesemia (±45%) (11, 82). HNF1β is a transcription factor regulating gene expression in kidney development (331). FXYD2β expression is one of several genes that is regulated by HNF1β (155), which may explain its role in renal Mg2+ handling. However, the possibility that HNF1β regulates other DCT genes involved in renal Mg2+ transport cannot be excluded.

J) PCBD1. In a small cohort of three patients, mutations in perin-α-carbinolamine dehydratase 1 (PCBD1) have been linked to hypomagnesemia, renal Mg2+-wasting, and MODY5-like diabetes (154). PCBD1 mutations are known to cause transient neonatal hyperphenylalaninemia and high urinary levels of primapertin (HPABH4D; OMIM 264070) (518, 519). HPABH4D patients are diagnosed at birth by Guthrie testing and suffer from a transient, benign defect in impaired BH4 regeneration. A follow-up study of three patients at ±18 yr of age showed that the patients display a mild hypomagnesemia (±0.6 mM) and MODY5-like diabetes, but no renal cysts (154). Interestingly, serum and urinary Na+, K+, Ca2+, and Cl− levels were within the normal range. The phenotype of the HPABH4D patients resembles that of RCAD patients, and the treatment regime consists of sulfonylureas and Mg2+-supplements. The origin of renal cysts in RCAD patients may be traced to the CD region of PKHD1 (194). However, since PCBD1 is not expressed in CD, HPABH4D patients are protected from cyst formation (154).

K) SLC12A3. Hypomagnesemia and hypokalemia are the cardinal symptoms of a hereditary electrolyte disorder characterized by Dr. Gitelman in 1969 that has been known since as Gitelman’s syndrome (175). Patients present with tetany, paresthesias, and chondrocalcinosis (284). The severity of the symptoms depends on the degree of hypokalemia. Except hypokalemia and hypomagnesemia, laboratory inves-
tigations often show metabolic alkalosis and hypocalciuria, sometimes associated with a mild hypotension and prolonged QT interval. SLC12A3 encodes the thiazide-sensitive Na\(^+\)-Cl\(^-\) cotransporter (NCC), and mutations here cause Gitelman’s syndrome (163, 481). Patients with Gitelman’s syndrome are often treated with oral Mg\(^{2+}\) supplements (284). Interestingly, in some patients Mg\(^{2+}\) supplementation restores normal K\(^+\) levels, suggesting that hypokalemia is secondary to hypomagnesemia (205). This hypothesis is further substantiated by the NCC KO mouse, which is hypomagnesemic but does not display K\(^+\) disturbances under basal conditions (460). NCC KO mice have markedly reduced TRPM6 expression levels, possibly explaining the renal Mg\(^{2+}\) wasting observed in Gitelman’s syndrome (368). However, the mechanism by which a loss of NCC function results in reduced TRPM6 expression remains unresolved. It has been suggested that the atrophy of the DCT segment observed in KO mice may partially explain this phenomenon (321).

1) SLC12A1, BSN, CLCNKB, and KCNJ1. Bartter’s syndrome was originally described by Dr. Bartter in 1962 and is characterized by salt wasting, hypokalemic alkalosis, elevated plasma renin and aldosterone levels, and low blood pressure (40). Mutations in SLC12A1, encoding NKCC2, Barttin, CLC-Kb, KCNJ1, encoding ROMK, or CaSR form the genetic basis of Bartter’s syndrome (50, 477–479). Mild hypomagnesemia is sometimes observed in Bartter’s patients, which may be explained by a reduced driving force for paracellular Mg\(^{2+}\) reabsorption in the TAL. Compensation for reduced TAL Mg\(^{2+}\) reabsorption may take place in the DCT, which explains why Bartter’s patients often have normal Mg\(^{2+}\) levels. CLC-Kb and Barttin are also expressed in DCT, which justifies why patients with mutations in these genes more often show hypomagnesemia (261).

3. Drug-induced hypomagnesemia

A) Diuretics. Hypomagnesemia has been associated with diuretics targeting the TAL and DCT segments of the kidney. In 1968, Duarte (127) reported renal Ca\(^{2+}\) and Mg\(^{2+}\) wasting as a consequence of furosemide treatment. Furosemide inhibits the activity of NKCC2, reducing the positive transepithelial membrane potential that drives paracellular Mg\(^{2+}\) transport in TAL (408). Although the incidence of furosemide-induced hypomagnesemia is unclear, a significant number of patients who use it may suffer from Mg\(^{2+}\) wasting (92, 330). In a recent animal study, furosemide treatment did not result in hypomagnesemia since increased TRPM6 expression in the DCT was able to compensate for reduced Mg\(^{2+}\) reabsorption in TAL (530). The clinical effects of furosemide treatment on Mg\(^{2+}\) levels may, therefore, depend on the individual’s ability to compensate at DCT level.

The use of thiazide diuretics, which target NCC in DCT, frequently induces renal Mg\(^{2+}\) wasting (366). Although most studies do not report hypomagnesemia in patients being treated with thiazide diuretics, some patient groups may be at risk (101). Patients with low initial Mg\(^{2+}\) values, such as elderly patients or patients with chronic heart failure, may develop hypomagnesemia after chronic thiazide treatment (238, 285). In mice, thiazide treatment reduces the renal expression of TRPM6, explaining the high urinary Mg\(^{2+}\) excretion (368). However, in most patients these effects may be small and the clinical consequences may depend on the patients’ basal serum Mg\(^{2+}\) values.

V) EGFR inhibitors. In 2005, it was first reported that the use of the EGFR inhibitor cetuximab can result in severe hypomagnesemia (459). Cetuximab (erbitux) is a monoclonal antibody against the EGFR and is generally prescribed for the treatment of colorectal or head and neck cancer. Up to 50–60% of patients using cetuximab may develop, and 10–20% display serum levels below 0.4 mM (341, 510). A recent meta-analysis showed a RR of 3.87 (396). Immediately after the first reports of cetuximab-induced hypomagnesemia, questions were raised whether the EGFR inhibitor erlotinib (tarceva) may have similar effects (16). However, as of this writing, no clinical reports on the Mg\(^{2+}\) status after the use of erlotinib are available. Animal studies demonstrated a small reduction in serum Mg\(^{2+}\) values after erlotinib administration (122). Erlotinib is generally administered as a tablet that also contains Mg\(^{2+}\) stearate. The copresence of Mg\(^{2+}\) in these tablets may explain the absence of clinical consequences of erlotinib administration on serum Mg\(^{2+}\) levels.

C) Proton pump inhibitors (PPI). In 2006, the use of protein pump inhibitors (PPI) was associated with hypomagnesemia for the first time in two separate patients receiving long-term omeprazole treatment (144). Since then, many new cases of PPI-induced hypomagnesemia have been reported (295). A recent systematic review of 36 cases demonstrated that discontinuation of PPIs resulted in recovery from hypomagnesemia within 4 days, and rechallenge led to reoccurrence within 4 days (226). Urinary Mg\(^{2+}\) excretion is low in these patients, suggesting normal kidney function and thus an effect of PPI use on intestinal Mg\(^{2+}\) absorption. In mice, administration of omeprazole increased the expression of TRPM6 in colon (300). Therefore, it was hypothesized that omeprazole may inhibit the activity of the colonic H\(^+\)-K\(^+\)-ATPase, resulting in reduced extraction of protons into the colon. Since TRPM6 activity increases at lower external pH, decreased proton secretion may reduce TRPM6 activity, for which increased TRPM6 expression may compensate (300, 313). However, increased TRPM6 expression may not be sufficient to prevent malabsorption of Mg\(^{2+}\) in colon in all patients. Individual variability of this compensatory mechanism may explain why only a subset of PPI users develop hypomagnesemia.
D) CYCLOSPORIN INHIBITORS. The calcineurin inhibitors (CNI) cyclosporin A (CsA) and tacrolimus (FK506) are currently the first immunosuppressant drugs of choice after transplantation. The use of CNIs has been associated with hypertension and renal Mg$^{2+}$ wasting (39, 517). Whereas the hypertension may be explained by an increased activity of NCC, renal Mg$^{2+}$ wasting is not fully understood (242). Up to 90% of all patients suffer from significantly reduced serum Mg$^{2+}$ levels after initiation of CsA treatment, and in a recent cohort even 35% of patients remained hypomagnesemic despite Mg$^{2+}$ supplementation (39, 439, 517). In rats, CsA and FK506 treatment increased renal Mg$^{2+}$ wasting and hypomagnesemia (29, 365). In FK506-treated rats TRPM6 mRNA is downregulated, resulting in dramatically increased Mg$^{2+}$ excretion (365). In a recent study with CsA treatment in rats, TRPM6, TRPM7, and EGF mRNA expression were reduced, although the fractional excretion of Mg$^{2+}$ did not significantly change (306). Interestingly, EGF treatment did not change Mg$^{2+}$ excretion and TRPM6 expression in CsA-treated rats, whereas it reduced Mg$^{2+}$ wasting and increased TRPM6 expression in control rats. These results suggest that CsA may interfere with the EGF signaling pathway in DCT cells. Patients receiving CNI treatment are generally supplemented with Mg$^{2+}$ to prevent hypomagnesemia.

E) CISPLATIN/CARBOPLATIN. Already from the introduction of cisplatin (cis-diamminedichloroplatinum) as anti-cancer therapeutic, hypomagnesemia has been reported in ~40%–80% of treated patients (453, 593). Nephrotoxicity is a common side effect of cisplatin treatment, mainly as a consequence of AGA use range from 20 to 80% (157, 562). AGAs may activate the CaSR, resulting in reduced paracellular Mg$^{2+}$ transport in the TAL and inhibition of Mg$^{2+}$ transport in the DCT. Studies in MDCT cells show reduced PTH-activated Mg$^{2+}$ transport (269). Moreover, animal studies have evidenced that use of AGAs cause hypomagnesemia, due to reduced expression of NKCC2 that provides the driving force for TAL Mg$^{2+}$ transport (167, 446). In a recent study with gentamycin-treated rats, TRPM6 expression was upregulated, suggesting that the DCT compensates for reduced TAL Mg$^{2+}$ reabsorption (307).

Pentamidine is an antimicrobial against Pneumocystis jirovecii infections that are often diagnosed in AIDS patients. The use of pentamidine has been associated with severe hypomagnesemia due to renal Mg$^{2+}$ wasting at the start of the 1990s (65, 188, 465). The exact mechanism of reduced Mg$^{2+}$ reabsorption remains unresolved. However, pentamidine reduces ENaC activity, resulting in hyperkalemia (283). Moreover, there have been reports of tubular necrosis after pentamidine treatment (546), which may cause atrophy of the DCT segment.

Rapamycin (sirolimus) is an antibiotic that is frequently used to prevent organ rejection after transplantation. Rapamycin inhibits mTOR activity, and its use has been associated with hypomagnesemia in 10–25% of patients (23, 535). Rapamycin-treated rats exhibit reduced expression of NKCC2 (100). Interestingly, TRPM6 expression was increased in the same study. This could be compensation for the reduced Mg$^{2+}$ uptake in TAL, but a direct effect of rapamycin on TRPM6 expression cannot be excluded. A recent in vitro study showed the opposite effect; rapamycin decreased TRPM6 expression in an EGF-dependent manner (255).

Amphotericin B is an antifungal agent that has been associated with hypomagnesemia and hypokalemia (38). The mechanism underlying urinary Mg$^{2+}$ wasting in these patients is unknown. Oral Mg$^{2+}$ supplementation together with amiloride treatment is generally used to restore Mg$^{2+}$ levels (178).

Foscarnet inhibits viral DNA polymerases by chelating divalent cations and, therefore, its use may cause hypomagnesemia (169). Patients also suffer from hypocalcemia and
VI. CONCLUDING REMARKS

Over the last decade, Mg$^{2+}$ has been considered as a treatment for several major diseases including preeclampsia, stroke, myocardial infarction, and asthma in several large-scale clinical trials. These findings have raised interest in Mg$^{2+}$ among neurologists, cardiologists, and pneumologists. Nevertheless, Mg$^{2+}$ levels are still not determined routinely in daily clinical practice, even though up to 60% of all critically ill patients are Mg$^{2+}$ deficient (84, 145). Serum Mg$^{2+}$ should be determined standardly alongside Na$^+$, K$^+$, and Ca$^{2+}$ measurements in patients. Mg$^{2+}$ disturbances can cause muscle cramps, arrhythmias, and seizures, and therefore, Mg$^{2+}$ should be considered when patients present in clinic with these symptoms. However, the molecular mechanisms underlying the effects of Mg$^{2+}$ in brain, heart, and lung are still largely unknown. Finding the molecular basis of the role of Mg$^{2+}$ in these diseases may further extend the significance of Mg$^{2+}$ therapies in clinic.

Genetic and drug-induced disorders of Mg$^{2+}$ homeostasis have enhanced the knowledge on Mg$^{2+}$ (re)absorption in the kidney and intestine. These studies form a perfect example of the powerful interaction of clinical and fundamental studies. For instance, the increased knowledge on the role of EGF in renal Mg$^{2+}$ handling has resulted in the standardization of Mg$^{2+}$ measurements in patients using EGFR blockers. This resulted in early detection of Mg$^{2+}$ disturbances and changes in treatment strategies. Through synergistic clinical and fundamental efforts in the fields of brain, heart, and lung Mg$^{2+}$ research, the unexplained role of Mg$^{2+}$ in, among others, migraine, depression, epilepsy, COPD, and hypertension, may be elucidated. Over the last decades Mg$^{2+}$ research has been centered around the kidney and the intestine. By extending the field to the heart, brain, and lungs and by involving both fundamental and clinical researchers, Mg$^{2+}$ will never be thought of as “a forgotten cation” anymore.

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DISCLOSURES

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