

This Provisional PDF corresponds to the article as it appeared upon acceptance. Fully formatted PDF and full text (HTML) versions will be made available soon.

## **Changes in circulating 25-hydroxyvitamin d according to vitamin D binding protein genotypes after vitamin D3 or D2 supplementation**

*Nutrition Journal* 2013, **12**:39 doi:10.1186/1475-2891-12-39

Hataikarn Nimitphong (hataikarnn@hotmail.com)  
Sunee Saetung (sunee.sae@mahidol.ac.th)  
Suwannee Chanprasertyotin (suwannee.cha@mahidol.ac.th)  
La-or Chailurkit (laor.cha@mahidol.ac.th)  
Boonsong Ongphiphadhanakul (boonsong.ong@mahidol.ac.th)

**ISSN** 1475-2891

**Article type** Research

**Submission date** 18 September 2012

**Acceptance date** 18 March 2013

**Publication date** 4 April 2013

**Article URL** <http://www.nutritionj.com/content/12/1/39>

This peer-reviewed article can be downloaded, printed and distributed freely for any purposes (see copyright notice below).

Articles in *Nutrition Journal* are listed in PubMed and archived at PubMed Central.

For information about publishing your research in *Nutrition Journal* or any BioMed Central journal, go to

<http://www.nutritionj.com/authors/instructions/>

For information about other BioMed Central publications go to

<http://www.biomedcentral.com/>

© 2013 Nimitphong *et al.*

This is an open access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

# Changes in circulating 25-hydroxyvitamin d according to vitamin D binding protein genotypes after vitamin D<sub>3</sub> or D<sub>2</sub> supplementation

Hataikarn Nimitphong<sup>1\*</sup>

\* Corresponding author

Email: hataikarnn@hotmail.com

Sunee Saetung<sup>1</sup>

Email: sunee.sae@mahidol.ac.th

Suwannee Chanprasertyotin<sup>1</sup>

Email: suwannee.cha@mahidol.ac.th

La-or Chailurkit<sup>1</sup>

Email: laor.cha@mahidol.ac.th

Boonsong Ongphiphadhanakul<sup>1</sup>

Email: boonsong.ong@mahidol.ac.th

<sup>1</sup> Division of Endocrinology and Metabolism, Department of Medicine, Faculty of Medicine, Ramathibodi Hospital, Mahidol University, Rama 6 Rd, Rajthevi, Bangkok 10400, Thailand

## Abstract

### Background

It is not known whether genetic variation in the vitamin D binding protein (DBP) influences 25-hydroxyvitamin D levels [25(OH)D] after vitamin D supplementation. We aimed to investigate the changes of total 25(OH)D, 25(OH)D<sub>3</sub> and 25(OH)D<sub>2</sub> in a Thai cohort, according to type of vitamin D supplement (vitamin D<sub>3</sub> or D<sub>2</sub>) and DBP genotype, after receiving vitamin D<sub>3</sub> or D<sub>2</sub> for 3 months.

### Methods

Thirty-nine healthy subjects completed the study. All subjects received 400 IU of either vitamin D<sub>3</sub> or D<sub>2</sub>, plus a calcium supplement, every day for 3 months. Total serum 25(OH)D, 25(OH)D<sub>3</sub> and 25(OH)D<sub>2</sub> were measured by LC-MS/MS. Individual genotyping of rs4588 in the *DBP* gene was performed using real-time PCR.

### Results

Vitamin D<sub>3</sub> supplementation of 400 IU/d increased 25(OH)D<sub>3</sub> significantly ( $+16.2 \pm 4.2$  nmol/L,  $p < 0.001$ ). Vitamin D<sub>2</sub> (400 IU/d) caused increased 25(OH)D<sub>2</sub> levels ( $+22.0 \pm 2.11$  nmol/L,  $p < 0.001$ ), together with a decrease of 25(OH)D<sub>3</sub> ( $-14.2 \pm 2.0$  nmol/L,  $p < 0.001$ ). At 3 month, subjects in vitamin D<sub>3</sub> group tended to have higher total 25(OH)D levels than those

in vitamin D<sub>2</sub> ( $67.8 \pm 3.9$  vs.  $61.0 \pm 3.0$  nmol/L;  $p = 0.08$ ). Subjects were then classified into two subgroups: homozygous for the *DBP* rs4588 C allele (CC), and the rest (CA or AA). With D<sub>3</sub> supplementation, subjects with CA or AA alleles had significantly less increase in 25(OH)D<sub>3</sub> and total 25(OH)D when compared with those with the CC allele. However, no difference was found when the supplement was vitamin D<sub>2</sub>.

## Conclusion

Genetic variation in *DBP* (rs4588 SNP) influences responsiveness to vitamin D<sub>3</sub> but not vitamin D<sub>2</sub>.

## Keywords

25-hydroxyvitamin D, vitamin D<sub>3</sub>, vitamin D<sub>2</sub>, vitamin D binding protein(DBP)

## Background

Vitamin D insufficiency has been increasingly recognized as a common health problem worldwide. Measures to augment vitamin D levels include increased sun exposure, higher consumption of vitamin D-rich foods, and taking vitamin D supplements. In order to achieve a 25-hydroxyvitamin D level [25(OH)D; a marker of vitamin D status] higher than the current recommended threshold of 50–75 nmol/L, higher doses of vitamin D than previously suggested are required in Caucasians ([1,2]). However, it is unclear if the suggestion holds across ethnic groups, particularly in populations with lower body fat as compared to Caucasians. Moreover, it has never been investigated in Asians if vitamin D<sub>2</sub> or D<sub>3</sub> supplementation would have a different effect on circulating vitamin D.

Vitamin D and its metabolites circulate in the plasma, bound to vitamin D binding protein (DBP) ([3]). Genetic variations in the *DBP* gene have consistently been found to be associated with 25(OH)D levels ([4-7]). However, it is currently unclear how the *DBP* genetic variation would affect the increase in 25(OH)D levels after taking vitamin D supplements.

In the present study, we investigated the changes of total 25(OH)D, 25(OH)D<sub>3</sub> and 25(OH)D<sub>2</sub> according to type of vitamin D supplement (400 IU of vitamin D<sub>3</sub> or D<sub>2</sub> for 3 months). Toward the end, we investigated the change in serum 25(OH)D levels according to *DBP* genotypes.

## Methods

### Subjects

A total of 41 subjects (34 females/7 males) aged 15–70 years were enrolled in an unblinded randomized control trial that began in August 2008. All subjects were healthy and did not have any underlying diseases. Subjects were excluded if they were taking vitamin D  $\geq 400$  IU/d before being included in the study. After inclusion, subjects were excluded from the final analysis if they were intolerant to the supplement or were lost to follow-up. Subjects were then randomly assigned into two groups, using a computer-generated randomized code,

to receive 400 IU/day of either vitamin D<sub>2</sub> or vitamin D<sub>3</sub> for 3 months. Two subjects (both of them were females) in the vitamin D<sub>2</sub> group were excluded from the study after enrollment; one subject had gastrointestinal side effects from the calcium supplement, and the second was lost to follow-up. Final analysis was based on data from 39 subjects (32 females/7 males): 20 and 19 subjects in the vitamin D<sub>3</sub> and D<sub>2</sub> groups, respectively. All subjects were supplemented during the rainy or winter season and we did not allow taking any additional vitamin supplement. The treatment period was only 3 months and all subjects stayed in the same environment. In addition, there is less natural vitamin D containing food and vitamin D fortified food in Thailand. Therefore we did not assess the duration of sun exposure and the amount of daily vitamin D intake.

All study participants arrived at the research unit at 0800 h after at least a 12 h overnight fast. Baseline characteristics – which included age, all medications currently in use, waist circumference (WC) and body mass index (BMI) – were recorded. Blood was collected at baseline. Subjects received either vitamin D<sub>3</sub> (Centrum®) or vitamin D<sub>2</sub> (MTV), as well as a calcium supplement (CaCO<sub>3</sub>) every day, and were asked to return to the clinic at 1 month and 3 months after the first visit. Fasting plasma glucose (FPG) was measured at baseline, and total serum 25(OH)D, 25(OH)D<sub>2</sub> and 25(OH)D<sub>3</sub> were measured at baseline and at every follow-up period. Intact plasma parathyroid hormone (PTH) was measured at baseline and 3 month. Subjects with total 25(OH)D levels <50 nmol/L were classified as having vitamin D deficiency. 25(OH)D, 25(OH)D<sub>2</sub> and 25(OH)D<sub>3</sub> were reported as levels (nmol/L) and as changes from baseline over the follow-up period (1 and 3 month).

## **Vitamin D and calcium supplementation**

All subjects received either vitamin D<sub>3</sub> or vitamin D<sub>2</sub> and a calcium supplement daily for 3 months. Subjects in the vitamin D<sub>3</sub> group received 400 IU/d vitamin D<sub>3</sub> and 675 mg/d elemental calcium [1 tablet of Centrum® (contained 400 IU of vitamin D<sub>3</sub> and 175 mg of elemental calcium) and 1 tablet of 1250 mg CaCO<sub>3</sub> (contained 500 mg of elemental calcium)]; subjects in the vitamin D<sub>2</sub> group received 400 IU/d vitamin D<sub>2</sub> and 500 mg/d elemental calcium [1 multivitamin tablet (contained 400 IU of vitamin D<sub>2</sub>) and 1 tablet of 1250 mg CaCO<sub>3</sub> (contained 500 mg of elemental calcium)]. Compliance was assessed by tablet-counting at every return visit, and was reported as % of medicine taken. All subjects had over 90% compliance for calcium, vitamin D<sub>2</sub> and vitamin D<sub>3</sub>.

## **Biochemical measurement**

Serum and plasma samples were kept frozen at –80 °C until analysis. Plasma intact parathyroid hormone (PTH) was determined by electrochemiluminescence immunoassay with an Elecsys 2010 analyzer (Roche Diagnostics, Mannheim, Germany). Serum 25(OH)D<sub>2</sub> and 25(OH)D<sub>3</sub> were analyzed by LC-MS/MS with an Agilent 1200 Infinity liquid chromatograph (Agilent Technologies, Waldbronn, Germany) coupled to a QTRAP® 5500 tandem mass spectrometer (AB SCIEX, Foster City, CA, USA) using a MassChrom® 25-OH-Vitamin D<sub>3</sub>/D<sub>2</sub> diagnostics kit (ChromSystems, Munich, Germany). The summation of serum 25(OH)D<sub>2</sub> and 25(OH)D<sub>3</sub> was used to reflect vitamin D status. The inter-assay and intra-assay coefficients of variation of serum total 25(OH)D level were 6.3% and 5.0%, respectively.

## SNP genotyping

DNA was extracted from a 200  $\mu$ L serum sample using a QIAamp® DNA Blood Mini Kit (Qiagen, Hilden, Germany) according to the manufacturer's protocol. Individual genotyping of rs4588 in the *DBP* gene was performed using real-time PCR (TaqMan® MGB probes): 20 ng of DNA was added into the PCR reaction, consisting of TaqMan Universal Master Mix (1x) and TaqMan MGB probes for intronic C/A SNP rs4588 (1x) in a total volume of 20  $\mu$ L. The real-time PCR reaction protocol was 10 min at 95 °C, 40 cycles of 15 s at 92 °C, and 1 min at 60 °C using a 7500 Real-Time PCR System (Applied Biosystems, Foster City, CA, USA).

## Statistical analysis

Data were expressed as mean  $\pm$  SEM unless stated otherwise. We performed non-parametric test to compare the difference between parameters. We used the Mann–Whitney test and the Chi-Square test to compare the difference of baseline characteristics and changes in vitamin D metabolites [total 25(OH)D, 25(OH)D<sub>2</sub> and 25(OH)D<sub>3</sub>] between groups of subjects stratified by vitamin D preparations and *DBP* genotypes. The changes of vitamin D metabolites at the 1<sup>st</sup> and 3<sup>rd</sup> month from baseline in each group of subjects were assessed by the Wilcoxon test. All statistical analyses were performed with SPSS version 16 (SPSS, Chicago, IL). A *P* value of <0.05 was considered statistically significant.

## Results

Thirty-nine subjects (82.05% female) with a mean age of  $36.2 \pm 1.3$  were included in the final analysis. All baseline characteristics of subjects in both vitamin D<sub>3</sub> and vitamin D<sub>2</sub> groups were similar (Table 1). Twenty-four out of 39 subjects (61.5%) had vitamin D deficiency [25(OH)D less than 50 nmol/L]: 14 and 10 subjects in the vitamin D<sub>3</sub> and vitamin D<sub>2</sub> groups, respectively (*p* = NS). As shown in Figures 1 and 2, after receiving 400 IU/d vitamin D<sub>3</sub>, 25(OH)D<sub>3</sub> levels increased significantly ( $7.8 \pm 1.5$  and  $16.2 \pm 4.2$  nmol/L at the 1<sup>st</sup> and the 3<sup>rd</sup> month, respectively; *p* < 0.001), with a small concurrent decrease in 25(OH)D<sub>2</sub>. Among subjects who received 400 IU/d vitamin D<sub>2</sub>, serum 25(OH)D<sub>2</sub> levels increased by  $15.6 \pm 1.3$  and  $22.0 \pm 2.1$  nmol/L at the 1<sup>st</sup> and 3<sup>rd</sup> month, respectively; there was, however, a significant decrease of serum 25(OH)D<sub>3</sub> ( $-14.2 \pm 2.0$  nmol/L at the 3<sup>rd</sup> month; *p* < 0.001). This resulted in a reduction in the increase of total 25(OH)D levels after vitamin D<sub>2</sub> supplementation. Nevertheless, when compared between the vitamin D<sub>2</sub> and D<sub>3</sub> groups, there were no significant difference in total 25(OH)D levels at 3 months from baseline [total 25(OH)D =  $61 \pm 3.1$  vs.  $67.6 \pm 3.9$  nmol/L and the increased of total 25(OH)D from baseline =  $7.8 \pm 2.2$  vs.  $15.9 \pm 4.3$  nmol/L (*p* = 0.08) for vitamin D<sub>2</sub> and vitamin D<sub>3</sub> group, respectively; Figure 1A). At the 3<sup>rd</sup> month, plasma intact PTH was decreased significantly from baseline in subjects of both vitamin D<sub>2</sub> [ $-0.6 \pm 0.3$  pmol/L, *p* < 0.05] and D<sub>3</sub> group [ $-0.7 \pm 0.2$  pmol/L, *p* < 0.01]. There was no difference in the decrement in intact PTH between 2 groups (*p* = 0.7).

**Table 1 Baseline characteristics according to vitamin D supplementation**

Clinical characteristics	Vitamin D <sub>3</sub> (n = 20)	Vitamin D <sub>2</sub> (n = 19)	P value
Age (years)	36.0 ± 1.9	36.7 ± 1.7	NS
Sex (M/F)	3/17	4/15	NS
Body weight (kg)	56.9 ± 2.0	59.5 ± 2.7	NS
Waist circumference (cm)	75.9 ± 1.5	78.4 ± 2.1	NS
BMI (kg/m <sup>2</sup> )	22.4 ± 0.5	23.2 ± 0.8	NS
FPG (mmol/L)	4.6 ± 0.1	4.5 ± 0.1	NS
Total 25(OH)D (nmol/L)	51.8 ± 3.8	53.2 ± 3.6	NS
25(OH)D <sub>3</sub> (nmol/L)	50.3 ± 3.8	51.7 ± 3.6	NS
25(OH)D <sub>2</sub> (nmol/L)	1.5 ± 0.1	1.5 ± 0.2	NS
PTH (pmol/L)	3.9 ± 0.3	4.0 ± 0.4	NS

Data is presented as mean ± SEM.

**Figure 1 Total 25(OH)D, 25(OH)D<sub>2</sub> and 25(OH)D<sub>3</sub> levels at baseline, and at 1 and 3 months after vitamin D<sub>3</sub> or vitamin D<sub>2</sub> supplementation.****Figure 2 The differences of total 25(OH)D, 25(OH)D<sub>2</sub> and 25(OH)D<sub>3</sub> levels from baseline at 1 and 3 months after vitamin D<sub>3</sub> or vitamin D<sub>2</sub> supplementation.**

Table 2 shows the genotype distribution of the *DBP* rs4588 SNP. The genotype distributions of subjects given vitamin D<sub>3</sub> or vitamin D<sub>2</sub> supplements were not significantly different. Subjects in each vitamin D supplementation group were then divided into two subgroups based on the presence of the A (minor) allele: those homozygous for the C allele (group 1), and the rest (CA or AA: group 2). When we compared baseline 25(OH)D and 25(OH)D<sub>3</sub> levels in subjects between group 1 (n = 22) and group 2 (n = 17), there was no difference in either baseline 25(OH)D (52.9 ± 4 vs. 52.1 ± 3 nmol/L, *p* = 0.5) or baseline 25(OH)D<sub>3</sub> levels (51.4 ± 4.1 vs. 50.6 ± 3 nmol/L, *p* = 0.6). We further classified subjects into 4 groups according to *DBP* genotype and type of vitamin D supplement (Table 3). While there was no difference in age, gender, BMI, and baseline 25(OH)D<sub>2</sub>, 25(OH)D<sub>3</sub> and total 25(OH)D between the genotypes among subjects in the vitamin D<sub>2</sub> group, some differences were found in gender, baseline 25(OH)D<sub>3</sub> and total 25(OH)D between the genotypes among subjects in the vitamin D<sub>3</sub> group (Table 3). When comparing changes in vitamin D metabolites at 3 months with baseline values (Table 4), it was found that subjects in group 2 (CA or AA alleles) had significantly less increase in 25(OH)D<sub>3</sub> levels after taking vitamin D<sub>3</sub>. Likewise, the increment in total 25(OH)D was lower. On the other hand, no difference in the increments of 25(OH)D<sub>3</sub>, 25(OH)D<sub>2</sub> or total 25(OH)D was detected between the two genotype groups when the supplement was vitamin D<sub>2</sub>. There was no difference in changes in serum intact PTH between two *DBP* genotype subgroups after 3 months of vitamin D supplementation (Table 4).

**Table 2 Genotype distribution of the *DBP* rs4588 SNP in the study population**

<b>DBP genotype</b>	<b>D<sub>3</sub> supplement (n = 20)</b>	<b>D<sub>2</sub> supplement (n = 19)</b>	<b>Total</b>
CC (%)	12 (60%)	10 (52.6%)	22 (56.4%)
CA (%)	6 (30%)	5 (26.3%)	11 (28.2%)
AA (%)	2 (10%)	4 (21.1%)	6 (15.4%)

**Table 3 Baseline characteristics according to *DBP* genotype and type of vitamin D supplement**

	<b>D<sub>3</sub> supplement</b>			<b>D<sub>2</sub> supplement</b>		
	<b>CC (n = 12)</b>	<b>CA/AA (n = 8)</b>	<b><i>P</i> value</b>	<b>CC (n = 10)</b>	<b>CA/AA (n = 9)</b>	<b><i>P</i> value</b>
Age (years)	37.6 ± 2.6	33.5 ± 2.8	NS	36.1 ± 2.1	37.4 ± 2.8	NS
% female	100	62.5	<0.05	72.7	88.9	NS
BMI (kg/m <sup>2</sup> )	22.0 ± 0.7	22.9 ± 0.9	NS	23.6 ± 1.1	22.7 ± 1.2	NS
25(OH)D <sub>3</sub> (nmol/L)	47.4 ± 5.7	55.0 ± 4.2	<0.05	56.3 ± 5.7	46.8 ± 3.9	NS
25(OH)D <sub>2</sub> (nmol/L)	1.6 ± 0.3	1.3 ± 0.1	NS	1.4 ± 0.2	1.6 ± 0.3	NS
total 25(OH)D (nmol/L)	49.0 ± 5.8	56.3 ± 4.2	<0.05	57.7 ± 5.7	48.3 ± 3.9	NS
PTH (pmol/L)	4.0 ± 0.5	3.7 ± 0.5	NS	4.0 ± 0.6	3.9 ± 0.6	NS

Data is presented as mean ± SEM.

**Table 4 Changes in vitamin D metabolites and PTH at 3 months compared to baseline according to *DBP* genotypes and vitamin D preparations**

	<b>D<sub>3</sub> supplement</b>			<b>D<sub>2</sub> supplement</b>		
	<b>CC (n = 12)</b>	<b>CA/AA (n = 8)</b>	<b><i>P</i> value</b>	<b>CC (n = 10)</b>	<b>CA/AA (n = 9)</b>	<b><i>P</i> value</b>
Δ 25(OH)D <sub>3</sub> (nmol/L)	22.98 ± 6.00	6.09 ± 3.03	<0.01	-15.38 ± 2.60	-12.85 ± 3.21	NS
Δ 25(OH)D <sub>2</sub> (nmol/L)	-0.39 ± 0.36	-0.28 ± 0.10	NS	23.31 ± 3.59	20.47 ± 2.11	NS
Δ 25(OH)D (nmol/L)	22.58 ± 6.18	5.84 ± 3.07	<0.01	7.93 ± 3.10	7.61 ± 3.35	NS
Δ PTH (pmol/L)	-0.8 ± 0.3	-0.7 ± 0.4	NS	-0.6 ± 0.5	-0.7 ± 0.4	NS

Data is presented as mean ± SEM.

## Discussion

In the present study, 400 IU/day of vitamin D<sub>3</sub> tended to increase total 25(OH)D levels more when compared with the same dosage of vitamin D<sub>2</sub> ( $p = 0.08$ ). The underlying basis for this finding appears to be a concurrent decrease in 25(OH)D<sub>3</sub> after supplementation with vitamin D<sub>2</sub>. This finding is in keeping with a number of studies which demonstrated a decrease in 25(OH)D<sub>3</sub> after vitamin D<sub>2</sub> supplementation either daily or weekly ([8-10]). However, there was also one other study which could not demonstrate a concurrent decrease in 25(OH)D<sub>3</sub>, and found that vitamin D<sub>2</sub> and D<sub>3</sub> were equipotent in improving vitamin D status ([11]). It is noteworthy that nearly all of the studies that demonstrated less efficacy of vitamin D<sub>2</sub> in increasing 25(OH)D levels used relatively higher doses of vitamin D, indicating that the less efficacy of vitamin D<sub>2</sub> may be dose-dependent ([9,12]). The recent meta-analysis indicated that vitamin D<sub>3</sub> is more efficacious at raising serum 25(OH)D concentration when given as a bolus dose [50,000 IU single dose (oral), 300,000 IU single dose (oral and intramuscular) and 50,000 IU/month (oral);  $p = 0.0002$ ] compared with administration of vitamin D<sub>2</sub>, but the effect was lost with daily supplement [1,000-4,000 IU/d;  $p = 0.10$ ] [13]. However, our findings suggest that this is not the case, since we were able to demonstrate the trend of the difference between D<sub>3</sub> and D<sub>2</sub> supplementation even at a lower dose, i.e. 400 IU daily. All of the previous studies were performed in Caucasians, and it is unclear if ethnic differences in

vitamin D metabolism could be the basis of our findings. The decrease in 25(OH)D<sub>3</sub> after vitamin D<sub>2</sub> supplementation, same as reported in the study of Demetriou ET, et al. [14], likely due to competition for the 25-hydroxylase enzyme by vitamin D<sub>3</sub> and vitamin D<sub>2</sub>. However, it is probable that enzymatic catalyzation by other enzymes with relatively minor roles, such as CYP24A1 ([8]) and CYP3A4, may be different for vitamin D<sub>3</sub> and D<sub>2</sub>, and thus be partially accountable for the observation.

The ability of either vitamin D<sub>2</sub> or vitamin D<sub>3</sub> to increase circulating 25(OH)D varies considerably among individuals. The explanations for the large between-individual difference include differences in adiposity ([15]), enzymatic degradation of vitamin D metabolites ([8]), and dietary composition ([16]), as well as fat malabsorption ([17]). In the present study, vitamin D<sub>2</sub> or vitamin D<sub>3</sub> supplementation resulted in varying increases in 25(OH)D. No association with adiposity as assessed by BMI, however, was demonstrated. Since the number of study subjects was small, the lack of power to detect association could possibly be responsible. Dietary composition and enzymatic degradation of vitamin D metabolites were not assessed in the present study.

The present study demonstrated that DBP genetic variation is another factor which can influence the responsiveness to vitamin D supplementation. The major function of DBP is the binding, solubilization and transport of vitamin D and its metabolites ([18]). A previous study found that both serum 25(OH)D<sub>3</sub> and 1,25(OH)<sub>2</sub>D<sub>3</sub> concentrations were significantly lower in mice lacking DBP, compared to wild-type mice ([19]). In another recent study, Lauridsen *et al.* showed that DBP phenotype determines the median plasma concentration of 25(OH)D<sub>3</sub> and 1,25(OH)<sub>2</sub>D<sub>3</sub> ([5]). With regard to genetic variation, single nucleotide polymorphisms in the *DBP* gene have been demonstrated to be related to 25(OH)D levels in Caucasians and Africans ([4,6]). A difference in the response of serum 25(OH)D after vitamin D<sub>3</sub> supplementation according to DBP genetic variants has also been reported ([7]). It is of note that the DBP genetic variants affected the change in vitamin D status only for vitamin D<sub>3</sub> but not vitamin D<sub>2</sub> supplementation in our study. One study reported that vitamin D<sub>3</sub> had greater affinity for the DBP than vitamin D<sub>2</sub> and 25(OH)D<sub>3</sub> also had greater affinity for the DBP than 25(OH)D<sub>2</sub> [20]. However, the binding capacity of DBP cannot totally explain the difference in responsiveness to vitamin D<sub>3</sub> supplementation as demonstrated in Armas, LA.'s study [9]. After receiving single dose of 50,000vitamin D<sub>3</sub> or vitamin D<sub>2</sub> orally, a similar rising of 25(OH)D levels in the first 3 days were observed in both group. However, much more rapid decline of serum 25(OH)D in the vitamin D<sub>2</sub>-treated subjects after 3 days seem to reflect substantially more rapid metabolism or clearance of the vitamin D<sub>2</sub> metabolite [9]. We speculated that the greater affinity of 25(OH)D<sub>3</sub> for the DBP and less clearance of 25(OH)D<sub>3</sub> would maintain its serum levels and accountable for the influence of DBP genetic variants in the case of vitamin D<sub>3</sub>,not vitamin D<sub>2</sub>. Further studies such as the differences in metabolism between vitamin D<sub>2</sub> and vitamin D<sub>3</sub> after binding to DBP are warranted.

There are a number of limitations in this study. The sample size is relatively small and may not have enough power to detect small changes, particularly those related to vitamin D<sub>2</sub> supplementation. A 50 nmol/L difference in baseline 25(OH)D<sub>2</sub> and 25(OH)D<sub>3</sub> was present. This may influence the increment of 25(OH)D<sub>2</sub> or 25(OH)D<sub>3</sub> levels after receiving vitamin D<sub>2</sub> or vitamin D<sub>3</sub> supplementation, and thus possibly affect the total 25(OH)D levels at the end of the study. The change in 25(OH)D at the end of the study may also be influenced by lifestyle factors influencing the degree of sun exposure during the course of the study. This potential confounding effect cannot be assessed since there was not a non-supplemented group in our study. It would be important to know if both the vitamin D<sub>2</sub> and vitamin D<sub>3</sub>

supplements had the same amount of vitamin D. However, vitamin D contents of both preparations were not measured. However, total 25(OH)D levels of subjects in both groups were similar at 3 months, suggesting that the variability in vitamin D content between vitamin D<sub>2</sub> and vitamin D<sub>3</sub> preparations, if any existed, was likely to be small. The other limitation is that multivitamin tablets containing vitamin D were used (in the vitamin D<sub>2</sub> group), and it is unknown if other constituents of the multivitamin would affect vitamin D absorption or metabolism. It has been demonstrated that statins, atorvastatin and rosuvastatin in particular, can influence the circulating levels of 25(OH)D ([21-23]). To our knowledge, no interference from common vitamins and minerals with vitamin D metabolism has been reported.

## Conclusion

Daily supplementation with vitamin D<sub>2</sub> tended to result in lower total 25(OH)D levels than supplementation with vitamin D<sub>3</sub> due to a concurrent decrease in 25(OH)D<sub>3</sub> levels in the former case. Genetic variation in vitamin D binding protein (rs4588 SNP) influences responsiveness to vitamin D<sub>3</sub> but not vitamin D<sub>2</sub>.

## Competing interest

The authors have nothing to declare.

## Author's contribution

HN and BO conceived of the study, participated in its design and coordination, performed the statistical analysis and helped to draft the manuscript. SS carried out the genotyping of rs4588 in the *DBP* gene. SC carried out the biochemical measurement. LC carried out the vitamin D metabolites measurement (LC-MS/MS). All authors read and approved the final manuscript.

## Acknowledgments

Address all correspondence and requests for reprints to : Hataikarn Nimitphong, M.D., at Division of Endocrinology and Metabolism, Department of Medicine, Faculty of Medicine, Ramathibodi Hospital, Rama 6 Rd., Rajthevi, Bangkok 10400, Thailand; Email: hataikarnn@hotmail.com, This work was supported by the Faculty Research fund, Ramathibodi Hospital.

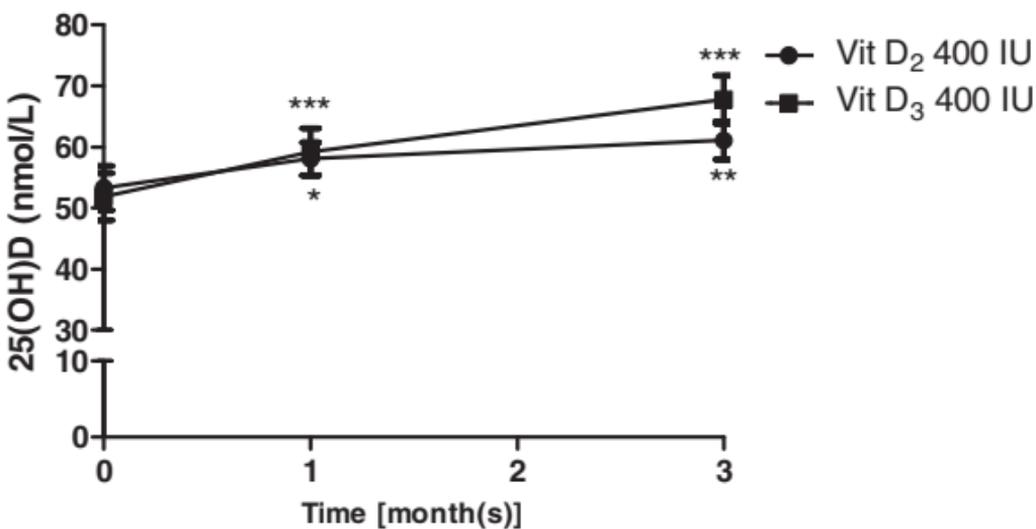
## References

1. Institute of Medicine: *Dietary Reference Intakes for Calcium and Vitamin D*. Washington, DC: National Academy of Sciences; 2011.
2. Holick MF, Binkley NC, Bischoff-Ferrari HA, Gordon CM, Hanley DA, Heaney RP, Murad MH, Weaver CM: **Evaluation, treatment, and prevention of vitamin D deficiency: an Endocrine Society clinical practice guideline.** *J Clin Endocrinol Metab*, **96**(7):1911–1930.

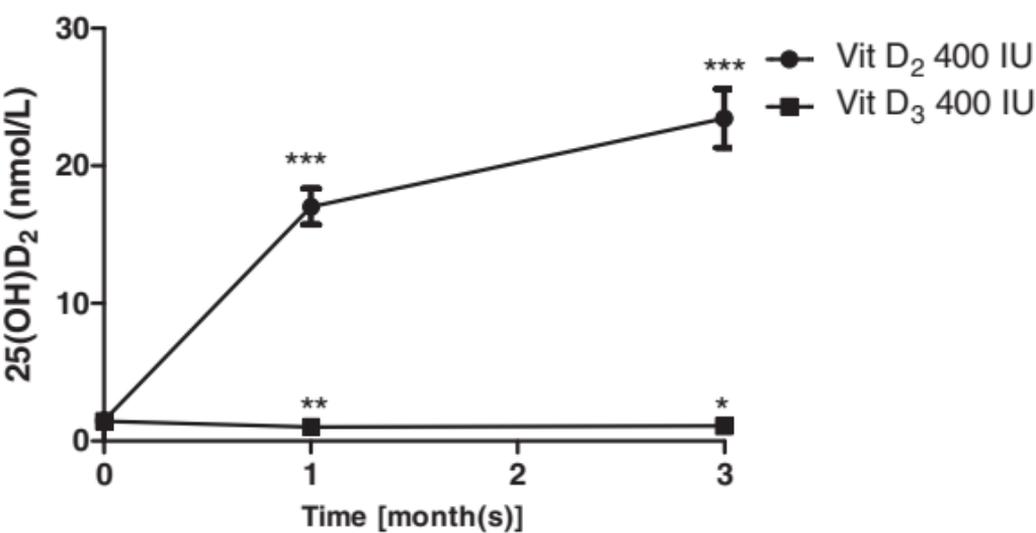
3. Bikle DD, Gee E, Halloran B, Kowalski MA, Ryzen E, Haddad JG: **Assessment of the free fraction of 25-hydroxyvitamin D in serum and its regulation by albumin and the vitamin D-binding protein.** *J Clin Endocrinol Metab* 1986, **63**(4):954–959.
4. Sinotte M, Diorio C, Berube S, Pollak M, Brisson J: **Genetic polymorphisms of the vitamin D binding protein and plasma concentrations of 25-hydroxyvitamin D in premenopausal women.** *Am J Clin Nutr* 2009, **89**(2):634–640.
5. Lauridsen AL, Vestergaard P, Hermann AP, Brot C, Heickendorff L, Mosekilde L, Nexø E: **Plasma concentrations of 25-hydroxy-vitamin D and 1,25-dihydroxy-vitamin D are related to the phenotype of Gc (vitamin D-binding protein): a cross-sectional study on 595 early postmenopausal women.** *Calcif Tissue Int* 2005, **77**(1):15–22.
6. Engelman CD, Fingerlin TE, Langefeld CD, Hicks PJ, Rich SS, Wagenknecht LE, Bowden DW, Norris JM: **Genetic and environmental determinants of 25-hydroxyvitamin D and 1,25-dihydroxyvitamin D levels in Hispanic and African Americans.** *J Clin Endocrinol Metab* 2008, **93**(9):3381–3388.
7. Fu L, Yun F, Oczak M, Wong BY, Vieth R, Cole DE: **Common genetic variants of the vitamin D binding protein (DBP) predict differences in response of serum 25-hydroxyvitamin D [25(OH)D] to vitamin D supplementation.** *Clin Biochem* 2009, **42**(10–11):1174–1177.
8. Thacher TD, Fischer PR, Obadofin MO, Levine MA, Singh RJ, Pettifor JM: **Comparison of metabolism of vitamins D2 and D3 in children with nutritional rickets.** *J Bone Miner Res*, **25**(9):1988–1995.
9. Armas LA, Hollis BW, Heaney RP: **Vitamin D2 is much less effective than vitamin D3 in humans.** *J Clin Endocrinol Metab* 2004, **89**(11):5387–5391.
10. Glendenning P, Chew GT, Seymour HM, Gillett MJ, Goldswain PR, Inderjeeth CA, Vasikaran SD, Taranto M, Musk AA, Fraser WD: **Serum 25-hydroxyvitamin D levels in vitamin D-insufficient hip fracture patients after supplementation with ergocalciferol and cholecalciferol.** *Bone* 2009, **45**(5):870–875.
11. Holick MF, Biancuzzo RM, Chen TC, Klein EK, Young A, Bibuld D, Reitz R, Salameh W, Ameri A, Tannenbaum AD: **Vitamin D2 is as effective as vitamin D3 in maintaining circulating concentrations of 25-hydroxyvitamin D.** *J Clin Endocrinol Metab* 2008, **93**(3):677–681.
12. Heaney RP, Recker RR, Grote J, Horst RL, Armas LA: **Vitamin D(3) is more potent than vitamin D(2) in humans.** *J Clin Endocrinol Metab*, **96**(3):E447–452.
13. Tripkovic L, Lambert H, Hart K, Smith CP, Bucca G, Penson S, Chope G, Hypponen E, Berry J, Vieth R, *et al*: **Comparison of vitamin D2 and vitamin D3 supplementation in raising serum 25-hydroxyvitamin D status: a systematic review and meta-analysis.** *Am J Clin Nutr*, **95**(6):1357–1364.

14. Demetriou ET, Travison TG, Holick MF: **Treatment with 50,000 IU vitamin D(2) every other week and effect on serum 25-hydroxyvitamin D(2), 25-hydroxyvitamin D(3), and total 25-hydroxyvitamin D in a clinical setting.** *Endocr Pract*, **18**(3):399–402.
15. Wortsman J, Matsuoka LY, Chen TC, Lu Z, Holick MF: **Decreased bioavailability of vitamin D in obesity.** *Am J Clin Nutr* 2000, **72**(3):690–693.
16. Niramitmahapanya S, Harris SS, Dawson-Hughes B: **Type of dietary fat is associated with the 25-hydroxyvitamin D3 increment in response to vitamin D supplementation.** *J Clin Endocrinol Metab*, **96**(10):3170–3174.
17. Farraye FA, Nimitphong H, Stucchi A, Dendrinis K, Boulanger AB, Vijjeswarapu A, Tanennbaum A, Biancuzzo R, Chen TC, Holick MF: **Use of a novel vitamin D bioavailability test demonstrates that vitamin D absorption is decreased in patients with quiescent Crohn's disease.** *Inflamm Bowel Dis*, **17**(10):2116–2121.
18. Speeckaert M, Huang G, Delanghe JR, Taes YE: **Biological and clinical aspects of the vitamin D binding protein (Gc-globulin) and its polymorphism.** *Clin Chim Acta* 2006, **372**(1–2):33–42.
19. Safadi FF, Thornton P, Magiera H, Hollis BW, Gentile M, Haddad JG, Liebhaber SA, Cooke NE: **Osteopathy and resistance to vitamin D toxicity in mice null for vitamin D binding protein.** *J Clin Invest* 1999, **103**(2):239–251.
20. Hollis BW: **Comparison of equilibrium and disequilibrium assay conditions for ergocalciferol, cholecalciferol and their major metabolites.** *J Steroid Biochem* 1984, **21**(1):81–86.
21. Sathyapalan T, Shepherd J, Arnett C, Coady AM, Kilpatrick ES, Atkin SL: **Atorvastatin increases 25-hydroxy vitamin D concentrations in patients with polycystic ovary syndrome.** *Clin Chem*, **56**(11):1696–1700.
22. Ertugrul DT, Yavuz B, Cil H, Ata N, Akin KO, Kucukazman M, Yalcin AA, Dal K, Yavuz BB, Tural E: **STATIN-D Study: Comparison of the Influences of Rosuvastatin and Fluvastatin Treatment on the Levels of 25 Hydroxyvitamin D.** *Cardiovasc Ther*.
23. Goldstein MR, Mascitelli L, Pezzetta F: **Rosuvastatin and vitamin D: might there be hypovitaminosis D on JUPITER.** *Int J Cardiol*, **145**(3):556–557.

### Total 25(OH)D



### 25(OH)D<sub>2</sub>



### 25(OH)D<sub>3</sub>

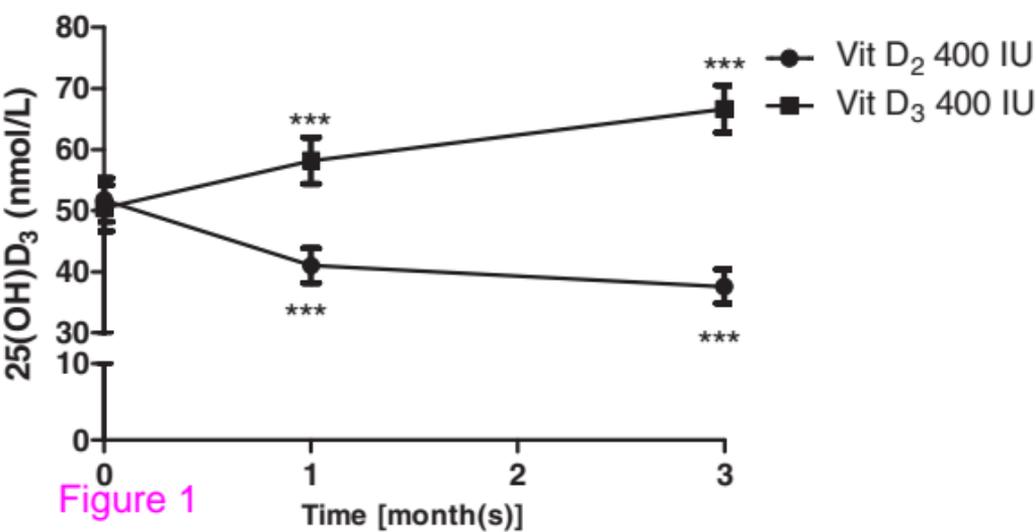
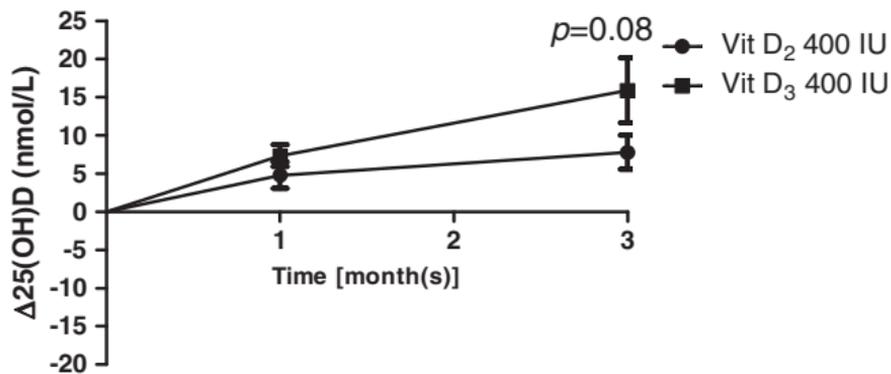
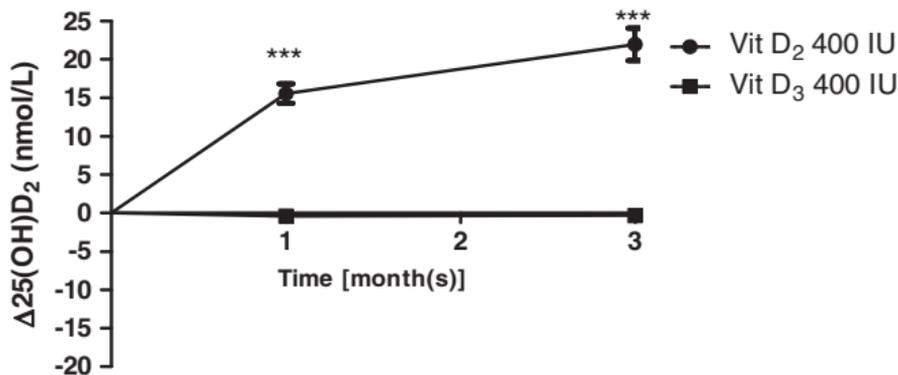


Figure 1

### $\Delta$ Total 25(OH)D



### $\Delta$ 25(OH)D<sub>2</sub>



### $\Delta$ 25(OH)D<sub>3</sub>

