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PII: S2405-8440(22)02857-2

DOI: https://doi.org/10.1016/j.heliyon.2022.e11569

Reference: HLY 11569

To appear in: HELIYON

Received Date: 2 May 2022

Revised Date: 1 August 2022

Accepted Date: 7 November 2022

Please cite this article as: A. Asghari, F. Jafari, M. Jameshorani, H. Chiti, M. Naseri, A. Ghafourirankouhi, O. Kooshkaki, A. Abdshah, N. Parsamanesh, Vitamin D role in hepatitis B: focus on immune system and genetics mechanism, *HELIYON*, https://doi.org/10.1016/j.heliyon.2022.e11569.

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Vitamin D role in hepatitis B: Focus on immune system and genetics mechanism

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Running title: Role of vitamin D in hepatitis B

Abstract

According to the World Health Organization (WHO) report, viral hepatitis has been a problem in

human society. Vitamins play a significant role in preventing the hepatocarcinoma and liver

cirrhosis. In this report, we will first focus on the vitamin D function in the immune system

reactions, and then investigate its role in the viral infections and the signaling pathway of hepatitis

B virus.

The existence of the cytochrome P450 (CYP) 27B1 enzyme, which is involved in vitamin D

synthesis in immune system cells, has drawn researchers 'attention to the field of immune system.

Toll like receptor (TLR) play a significant role in the immune system, and are one of the primary

receptors of the innate immune system. In addition, the synthesis of inflammatory cytokines, such

as Interferon γ (IFN γ) and Interleukin-2 (IL-2) is one of the key roles of T helper type 1 (Th1) cells;

these cells can suppress two cited cytokines via vitamin D. In the chronic phase of hepatitis B,

Cytotoxic T lymphocytes (CTLs) cells have weaker performance than the acute phase of the

disease. The association between vitamin D physiologies with viral infections is also confirmed by

genetic studies, carried out on genetic variations of vitamin D receptor (VDR) R-encoding disease

susceptibility gene. Vitamin D affects different phases of the disease. Therefore, further

experiments in this area are proposed.

Key words: Vitamin D, Hepatitis B, Immune system, Genetics

Introduction

Viral hepatitis has become a problem in human society. According to the World Health Organization (WHO) report, one third of people suffer from hepatitis B (HBV) and hepatitis C viruses (HCV) (1, 2). Hepatitis B has infected one-third of the world's population, 5% of infected individuals are known as carriers; among the infected subjects, 25% have chronic liver inflammation, which progresses to liver cirrhosis and finally hepatocarcinoma (3). Diet plays an important role in counteracting the effects of hepatocarcinoma and liver cirrhosis; one of the reasons could be the presence of vitamins in the diet (4, 5). Vitamins affect the development of hepatocarcinoma and liver cirrhosis (6).

1, 25-Dihydroxvitamin D3 (1, 25 (OH) D3) or vitamin D is one of the fat-soluble vitamins. It is mainly produced through UV radiation and conversion of 7-dehydrocholesterol to 1,25(OH)D3; although this vitamin is also obtained through diet such as: Oily fish, meat, milk products and fortified food products (7, 8). Generally has been agreed, that the least serum/plasma concentration of 25-hydroxyvitamin D (25(OH)D) is approximately 30 nmol/L to prevent vitamin D deficiency-related bone disease; hence this threshold is appropriate for defining vitamin D deficiency in population - based studies. Vitamin D deficiency has become a global issue in the 21st century. According to the existing studies, vitamin D deficiency has been seen in one billion people worldwide (9). Vitamin D status differs in various populations. Multiple studies have reported the rate of vitamin D deficiency with an evaluated prevalence of 30 -93 % in population with strong sun exposure, such as China, Turkey, India, Iran and Saudi Arabia over the past twenty years (10). Recently, vitamin D have been introduced as an effective factor in immune system regulation, autoimmune diseases, cardiovascular and respiratory health, pregnancy, obesity, erythropoiesis, diabetes, muscle function, and aging (11).

Vitamin D can show its effect on the immune system directly and indirectly. It directly modulates and regulated the immune system (12, 13); but indirectly has special effects on the progression and treatment of infectious disease. For example, it affects the absorption of minerals, such as calcium. It also has synergic effect with other vitamins and mineral, like zinc and vitamin C; each specifically affects the immune system (14, 15). They makes it difficult to understand the main function of vitamin D in antiviral drugs (16, 17). Studies have shown that vitamin D deficiency can reduce the effectiveness of antiviral drugs (17, 18).

In this article, we will first focus on the vitamin D biosynthesis and signaling in the immune system responses (innate and adaptive), and then discuss its role and effects on HBV immune-pathogenicity, due to viral infections and involved polymorphisms.

1. Vitamin D biosynthesis and signaling

A large body of evidence has suggested, that vitamin D could be obtained either from the diet of individuals or through synthesis in skin following exposure to sunlight. Because of cutaneous production of vitamin D due to Ultraviolet B (UVB) exposure, its synthesis might be related to different individualized factors, such as: latitude, season, use of sunblock, and skin pigmentation. Melanin has a key role in the synthesis of vitamin D; it absorbs the UVB radiation and inhibits Vitamin D synthesis from 7-dihydrocholestrol. As this form of vitamin D is inactive, in the next step it is hydroxylated in the liver to form 25(OH)D3 (19, 20).

It should be noted that this form of vitamin D (25 D) is still inactive, and mostly is used as a reliable form to measure and evaluate Vitamin D levels in individuals. This form 25(OH)D3 could be activated in kidneys with the effect of 1-alpha-hydroxylase, an enzyme which could be simulated

by parathyroid hormone (PTH); it converts to active compound of vitamin D, named 1, 25 dihydroxy vitamin D (1, 25 D) or calcitriol (21, 22).

This activated form (1, 25 D) could be further metabolized in the liver to produce the inactive form of 1, 24, 25 vitamin D, by mediating 24-hydroxylase enzyme (*CYP24A1*). In the physiological condition, a negative feedback loop regulates the levels of 1, 25 D in blood circulation. In this regard, the circulating level of the active form of vitamin D is maintained within limited boundaries, through negative feedback loop; 1, 25 D could inhibit renal 1-alpha-hydroxylase and stimulate the 24-hydroxylase enzyme. In addition, 1, 25 D has a couple of roles; it induces calcium absorption in the intestine, and osteoblast differentiation and matrix calcification in the bones. The active form implicates in the normal function of these organs by binding to *VDR* (7).

Vitamin D insufficiency in chronic liver disease is thought to be multiple (23). A decreased liver function might be one explanation for the low vitamin D levels seen in chronic liver diseases. Hepatic damages result in lower synthesis of vitamin D carrier proteins such vitamin D binding protein and albumin, as well as reduced vit D hepatic hydroxylation to 25(OH)D3 or calcidiol (24). In a review published in 2012, Stokes et al. defined the role of vit D in liver disease extremely effectively (25). Decreased exogenous exposure of patients to vit D sources, dietary vitD3 intestinal malabsorption, poor hepatic hydroxylation of 1, 25(OH)D3 to 25(OH)D3, and enhanced catabolic elimination of 25(OH)D3 are reported as key factors for vit D insufficiency in liver abnormalities (25). Trepo et al (26), Wong GL et al (23), and Finkelmeier et al (27) have all demonstrated a link between liver-related complications and low Vit-D levels (28).

Mousa et al demonstrated that despite prescription vitamin D supplementation in high-dose and comprehensive endpoint assessments, vitamin D supplementation could not increase insulin secretion or sensitivity in vitamin D-insufficient, obese, or overweight patients. As a result, even

in vitamin D-deficient communities, vitamin D supplementation is unlikely to be an effective approach for lowering diabetes risk (clinicaltrials.gov: NCT02112721) (29). Vitamin D supplementation in childhood decreases the incidence of T1DM later in life, according to observation data obtained from case-control and some few cohort research(30, 31). Clinicaltrials.gov recognized two RCTs (not yet published) on the prevention of asthma exacerbations in children (identifiers: NCT02687815 and NCT03365687) that may bring extra data on the effect of vitamin D in asthma cases (32).

VDR complex with the retinoid X receptor (*RXR*). The 1,25D-*VDR-RXR* heterodimer translocate to the nucleus, where it binds to the vitamin D responsive elements in the promoter regions of vitamin D responsive genes; it induces expression of these vitamin D-responsive genes. In addition, it has been shown that *VDR* could present on other tissues, including breast, bone marrow, brain, colon malignant cells and immune cells. For this reason, Vitamin D may have other important functions except its role in calcium and bone homeostasis. Furthermore, there are tissues other than kidney, which could express 1-alpha-hydroxylase; 25 D could be converted to 1, 25 D in these organs (33, 34). Recently, Vitamin D is considered to have endocrine functions, as it could act in paracrine or autocrine manner. In this regard, Vitamin D has other different actions, including stimulation of cell proliferation and promotion of cell differentiation. It may have a substantial role in promoting the protective immunity by its immunologic effects (Figure 1).

Vitamin D Supplementation in Different Conditions

Various factors such as age, weight, concomitant diseases, latitude, breed and nutritional culture can affect the daily requirement of vitamin D .Vitamin D supplementation should have no side effects, in addition to providing adequate serum levels of 25 (OH) D (35). Most early studies examining the relationship between serum vitamin D level and individuals' health, considered 20

ng / ml as the minimal level of serum 25 (OH) D for human well-being; but it has been contested by recent researchers. Most recent studies have found that maintaining the serum level of 25 (OH) D in the range of 30-50 ng / ml or 40-60 ng / ml is more appropriate (35-37).

According to the Australian research, the minimum serum level of this vitamin to reduce the risk of skeletal diseases, such as rickets (10 ng/m) and osteoporotic fractures (20 ng/ml) appears lower, in comparison to the prevention of premature death (30 ng/ml) and non-skeletal diseases, such as depression (30 ng/ml), diabetes and cardiovascular disease (32 ng/ml), falls and respiratory tract infections (38 ng/ml) and cancer (40 ng/ml) (38).

When there is vitamin D deficiency, the European guidelines have recommend vitamin D supplementation as follows: 1000 IU / Day for neonates under one month, 2000-3000 IU/Day for infants over one month and toddlers, 3000-5000 IU/ Day for children and adolescents aged 1- 18 years, 7000-10000 IU/Day or 50000 IU/ Week for adults and the elderly (39, 40).

In patients with vitamin D deficiency, who suffer from a concomitant disease, the need for vitamin D supplements may vary (41). For example, in patients with intestinal malabsorption, either intramuscular injection of vitamin D or larger oral doses up to 50,000 IUS should be administered every 2-3 days; even UVB light can be used as an alternative treatment (42). In severe hepatic and renal insufficiency, vitamin D deficiency should be treated with the active form of vitamin D (calcitriol). However, in patients with chronic renal failure, vitamin D usage is important to maintain serum 25 (OH) D levels above 30 ng/ml (43). In granulomatous diseases such as sarcoidosis and lymphoma, excessive vitamin D replacement will lead to hypercalcemia and osteomalacia. Serum 25 (OH) D levels should be maintained 20- 30 ng/ml, and should not exceed 30 ng/ml (44). Vitamin D deficiency should be treated in patients with hypercalcemic disorders, such as primary

or tertiary hyperparathyroidism; in these patients vitamin D replacement to achieve at least 30 ng/ml of concentration, will not exacerbate hypercalcemia (44).

The maximum tolerable amounts of vitamin D intake, which do not lead to side effects are as follows: 1000IU/Day for neonates under one month, 2000IU/Day for infants and children between one month and 10 years, 4000IU/ Day for children and adolescents aged 11- 18 years, 10000IU/ Day for adults and the elderly (45, 46). The serum 25 (OH) D concentration up to 100ng/ml is usually safe; the concentrations more than 150 ng/ml are considered toxic (47).

Glutathione (GSH), a tripeptide found mainly in the liver, is the most essential thiol decreasing agent essential in redox process regulation (48, 49). Glutathione therapy has been shown quickly increase the anti-oxidative stress and improve the liver membrane structure in the treatment of chronic hepatitis B (50). Moreover, GSH inhibits cytokine production, reduces active effector cell aggregation, prevents effector cell activation, and reduces cytokine induced damage, all of which contribute to cytokines' pro-inflammatory action on liver cells (51). Glutathione is essential for the biosynthesis and hydroxylation of 1, 25-dihydroxy-vitamin D and 25-hydroxy-vitamin D from diary vitamin D in the body (52). Also, in the livers of obese mice given a high-fat diet, glutathione deficiency causes epigenetic changes in vitamin D metabolism genes. Several reports have demonstrated that combining vitamin D with L-cysteine (a glutathione precursor) results in a more effective rise in circulating 25-hydroxyvitamin D and a reduction in inflammatory biomarkers(53). In human renal proximal tubule epithelial cells, the increase in GSH level mediated by L-cysteine supplementation was associated with upregulation of VD-regulatory genes including VDR, RXR and CYP27B1 and also downregulation of CYP24A1. Other studies confirmed that, supplementing with a combination of L-cysteine and VD or glutathione precursor, rather than just VD, is helpful and aids in the effective implementation of VD administration(54). Moreover, combining vitamin

D and LC supplementation may be more beneficial in reducing the risk of oxidative stress and adverse effect related with type 2 diabetes or COVID-19 infection (55-57). This attitude be able to reduce cellular damage in systemic inflammatory disease, such as hepatitis, obesity, diabetes, and hypertension caused by cytokine storm. It appears that people with hepatitis B who have low glutathione levels need a different way to increase their vitamin D levels, immune system, and function in hepatitis B (58).

2. Role of Vitamin D in immune system

Innate immunity and vitamin D

A complex network compromising many cells, tissues, organs, and the substances, which helps the body fight infections and diseases, while maintaining tolerance to self is named immune system. The immune system includes white blood cells and organs and tissues of the lymph system, such as the thymus, spleen, tonsils, lymph nodes, lymph vessels, and bone marrow (59). The importance of vitamin D in the immune system was discovered, when the researchers found the role of vitamin D receptor and *CYP27B1*enzyme, which is involved in the vitamin D metabolism in the immune cells (60).

There are a couple of immune cells presenting *VDR* on the surface, including: antigen-presenting cells (macrophages and dendritic cells), T cells and B cells; on the other hand, the mentioned cells could synthase vitamin D. Additionally, it has been demonstrated that local levels of 1, 25 D may differ with circulating levels, as the mediating enzymes in local synthesis of vitamin D are different from the controls originating from kidney (61, 62).

Its regulatory role in the immune system was first considered by researchers, which is done by activating macrophages and monocytes (63). The presence of *CYP27B1* enzyme, has attracted the

attention of researchers to the field of immune system (64). It should be noted that the extra renal 1-alpha-hydroxylase enzyme in macrophages is not the same as enzyme renal-hydroxylase, as it not regulated by PTH (65). Extra renal hydroxylase function depends on circulating levels of vitamin D, rather it could be stimulated by cytokines, such as IFN γ , IL-1, or tumor necrosis factor (TNF- α) (66). Moreover, the macrophage 24- hydroxylase enzyme is a nonfunctional splice variant, so there is no negative feedback to regulate the local levels of 1, 25 D (67, 68).

Researches have shown, that vitamin D can play a significant role in both innate and acquired immune systems (69, 70). TLRs are stimulated by PAMPs (pathogen associated molecular patterns) and DAMPs (damage associated molecular patterns) detection; they induce immune responses against infectious agents including fungal, bacteria and virus (71). This receptor initiates anti - viral reactions by detecting the nucleic acids and envelope glycoproteins; antiviral response involves the production of cytokines, interferon and chemokines (72).

Vitamin D can reduce TNF-α production by affecting TLR 2 and 4 expression (73, 74). A study of vitamin D levels and TLR3 receptor gene expression found, that vitamin D reduces receptor expression in the epithelial cells (75). In another study by Alvarez-Rodriguez et al., 25OHD form of vitamin D reduced the TLR7 receptor expression in acquired immune cells and monocytes (76). Djukic et al. showed, that vitamin D deficiency reduced the levels of proinflammatory cytokines by TLR1/2, TLR3, TLR4 and TLR9 (77). Although vitamin D appears to have a decreasing effect on the TLR genes expression, some studies have reported conflicting findings. For example in a study by Samar Ojaimi and colleagues in 2013, it was shown that after taking vitamin D and reache a sufficient level in the vitamin D deficient individuals, TLR 2 gene expression has greatly increased (78).

Stimulation of innate immune receptors by DAMP and PAMP causes antimicrobial peptides expression. Antimicrobial peptides with at least 100 amino acids in their structure are considered as antimicrobial agents in the innate immune system. In general, antimicrobial peptides are divided into two categories: cathelicidin and defensins (79). Although we are more likely to recognize this infectious antimicrobial peptide as a defense barrier against bacteria, their role in the viral infections has also been observed, recently (80, 81).

LL-37 is the only member of the human cathelicidin family, overall LL-37 is effective in regulating immune system, and has spectrum antimicrobial activity role. Studies have shown that LL-37 binds to dsRNA and has antiviral activity (82, 83). LL-37 binding to RNA, causes LL-37 detection by scavenger receptors (SRs), and enhances TLR3 signaling, then generates an inflammatory response (84, 85). A study found that the presence of vitamin D induced LL-37, reduces respiratory system infection (86). Studies have shown, that vitamin D induces cathelicidin production by affecting signaling TLR2/1 (87).

Vitamin D also increases antiviral responses by inducing the expression of β -defensins (88). Vitamin D levels can increase the expression of V β -defensin-2 and β -defensin-3 by stimulating the TLR2/6 receptors (89). Some studies have shown that β -defensin-2 can cause antiviral responses by acting on virus receptors and replicating viruses (90) (Figure 2).

Adaptive Immune system and vitamin D

Acquired immune system can be divided into two main parts: humoral immunity and cellular immunity. B lymphocytes are the main members of the humoral immune system. Cellular immune organs include T lymphocytes. In the following, we will examine the effect of vitamin D on these two systems. Despite the stimulant effect of vitamin D on innate immunity, it modulates the

acquired immune system (91). In the following, we will examine the effect of vitamin D on the humoral and cellular immunity.

Cellular immunity is a defensive immune mechanism involving the recruitment of phagocytes, antigen-sensitized cytotoxic T cells and the release of antigen-responding cytokines and chemokines. Cellular immunity is most powerful against the virus-infected cells, intracellular bacteria, fungi and tumor cells (92). Important members of this immune system are T-cells, which themselves are generally divided into four basic sections, including T-helper 1, T-helper 2, T regulator, and cytotoxic T-cells; although these divisions are currently undergoing wide variations. One of the main functions of T helper (Th)1 cells is to produce inflammatory cytokines, such as IFNγ and IL-2, which can be reduced by vitamin D (92). This property can be exerted by affecting the NFAT / AP-1 and Runx1 transcription factors (93). But VitD3's direct impacts on Th2 cells are still unknown (94). However, its effect on T-helper 2 cell is different from T-helper 1. It activates the cited cells and increases IL-4 production (91). Vitamin D increases the function of regulatory T cells. This action is performed by increasing the transcription factor Foxp3, which has a special role in differentiating this cell from regulatory T cell (95). Vitamin D also acts as a subset of other Tcells, T-helper 17 inhibits these cells. Inhibition of T-helper 17 function occurs in two ways: inhibition of IL-17 expression and IL-17 function suppression to differentiate T-helper 17 (93). There are some studies, showed mDC treatment with vitamin D and taking it to a culture medium containing T cells, will increase the population of regulatory T cells (96). In another study, it was found that vitamin D could directly affect regulatory T cells. A seasonal study of lymphocyte counts showed that the number of CD8 + and CD 4 + Tcells increases in summer (97). Based on this, they hypothesized that vitamin D may affect T cell proliferation. One study showed that CTL activation affects the vitamin D receptors expression (98). Some studies have shown, that vitamin D induces

an inhibitory response to CTL and modulates the immune reactions in some infectious diseases (99, 100). This is the case if it has a different effect on tuberculosis disease, and the presence of vitamin D causes activation of CTL and production of cytokines such as IFN γ and TNF- α (101). Many studies have not been done on the effect of vitamin D on B cells. A study by Chen et al. showed, that B cells rarely act in the inactive state of *VDR*; activation of these cells are associated with this receptor expression increment (102). Vitamin D appears to have an effect on antibody production and antibody class switching in the in vitro studies (102, 103). But it does not show these effects in the in vivo studies (104). On the other hand, vitamin D can indirectly inactivate B cells by affecting Tcells activation (figure2).

The role of vitamin D on HBV

According to existing studies, there are conflicting data on the effect of vitamin D on hepatitis B disease. Some studies have declared vitamin D deficiency reduces the therapeutic effects in infectious patients. Vitamin D inhibits virus replication (105-108); it has an effective role in platelet and albumin levels and reducing ALT enzyme levels in patients with acute hepatitis (109-112). In a research study, severe vitamin D deficiency in patients with autoimmune hepatitis has led to disease progression and lack of response to treatment. Severe vitamin D deficiency has been identified as a prognostic marker (113). Another study of people with hepatitis B, which included patients with hepatocarcinoma and liver cirrhosis and hepatitis B infected individuals, evaluating the effect of vitamin D on disease progression, showed that the vitamin inhibits the adverse effects of disease and can prevent disease progression (105). A prospective cohort study found that vitamin D deficiency was common in individuals with chronic HBV infection, and it was associated with adverse effects in the patients (114). A study of HBV-transfected cells showed lowering the binding level of vitamin D increases the expression of viral genes and virus replication (115).

Other available studies point to lack of link between vitamin D levels and prevention of disease progression. Some studies found lack of association between vitamin D levels with HBV viral infection (116, 117). In another study, it was shown that, vitamin D levels has no effect on hepatic fibrosis and viral loaded virus in antigen e positive cases (118). A study also showed that hepatitis patients, who received tenofovir disoproxil fumarate (TDF) plus peginterferon alfa-2a (PegIFN) treatment complained about no effect of vitamin D normal level on the treatment effectiveness (109).

The discrepancies between studies on vitamin D deficiency and its effects on the hepatic infection of patients can be due to differences in the viral genotypes. For example, a study by Huijuan Zhu et al. showed that genotype B virus is more affected by vitamin D (119). This statement has been confirmed by other studies (120).

The HBV virus is a noncytopathic virus; liver damages are due to the host immune reaction against its own infected cells (121, 122). Although it has been a long time since the discovery of HBV, the immune-pathogenicity is still unknown. The main cause of liver damage is due to immune system responses to virus replication (123, 124). CTLs play a major role in immune-pathogenicity. However, due to the inefficiency of these cells in completely eliminating viruses, other immune cells such as TH, Natural killer (NK) and Neu contribute to generate non-specific inflammatory responses (125).

HBV infection like other diseases, has two main phases, the acute and the chronic phases. Almost 95% of immune-competent persons with acute hepatitis successfully eradicate the HBV. But at this stage, we see disease complications, such as inflammation and necrosis of hepatocytes. The mortality rate is estimated to be 0.5-1% in the acute phase (126, 127). During acute HBV infection,

the serum level of ALT and T cells infiltration increase. It also reduces the level of HBV serum antigens and viral load.

The acute phase is associated with the onset of the innate immune response. Innate immunity plays an important role in disease pathogenesis. TLR receptors are important member of this system; they play a dual role, by producing inflammatory cytokines and preventing the proliferation of viruses and causing damage to the liver. The side effects are very mild and usually have no symptoms in the patient (128). Acute inflammatory responses usually lead to virus clearance. Therefore, it is preferable to create strong immune responses at this stage, to prevent the disease from becoming chronic (129).

In a study shown on HBV transgenic mice, all receptors were TLR3, TLR4, TLR5, TLR7, and TLR9. They are involved in inhibiting HBV replication by producing antiviral cytokines, especially INF type I and TNF- α (Isogawa, Robek et al. 2005). Although it is not clear which pattern recognition receptors detect the HBV; studies have shown that the core protein of the virus is detected by TLR 2 on macrophages, due to the arginine-rich domain (Cooper, Tal et al. 2005). There is much evidence that TLR2 plays a special role in initiating an immune response against HBV. In a study by Cooper et al., the receptor was activated by the HBc antigen to activate macrophages and produce pro-inflammatory cytokines. As a result, the disease remains in the acute phase. Vitamin D reduces the pattern recognition receptors expression, and as a result it reduces the inflammatory cytokines production (130). Some studies have shown that vitamin D by affecting TLR 2 expression can increase receptor expression, which in turn can inhibit the HBV virus in the early stages of the disease. It has been reported that HBV can interfere with TNF- α signaling and cathelicidin by reducing the expression of vitamin D receptor gene, thereby inhibiting the immune system response (131).

A study conducted by Monika Merkle et al. in 2015 showed, that IL37 secreted by the activation of innate immune receptors may have protective effect against HBV (132) According to other studies on the effect of vitamin D on IL37, it is expected that vitamin D inhibits the disease by increasing the expression of this factor, but a study showed that vitamin D in hepatitis patients has no effects on IL37 (133).

CD14 is a glycoprotein that binds to some ligands such as LPS; it activates TLR 4. HBsAg binding to the CD14 receptor causes the innate immune response. Although studies have shown that CD14 is a LPS receptor, and responds to TLR 4. Due to its similar lipid structure to HBsAg, it also binds to CD14 (Vanlandschoot, Van Houtte et al. 2002). Studies have shown, that vitamin D increases the expression of CD-14 in epidermal keratinocytes and monocytes (Oberg, Botling et al. 1993, Schauber, Dorschner et al. 2007).

Many studies have been done on the effect of vitamin D on NK and T cells. But in general, it can be said that vitamin D has a direct effect on the evolution and proliferation of NKT cells; in the situation of vitamin D deficiency, the number of NKT cells decreases (134-136). Analyzes of Peripheral blood mononuclear cells (PBMCs) in individuals in the acute phase of hepatitis B disease showed an increase in NKT count, and its activity before HBV-specific T cells activation (137). These cells play an important role in controlling the acute phase of HBV by producing cytokines, such as IFNγ and TNF-α. These cytokines cause cccDNA (covalently closed circular DNA) instability and decrease serum HBV DNA levels (138).

The role of specific immune systems in the acute phase of hepatitis B disease can be attributed to T cells, especially CTLs. In the acute phase of hepatitis B disease, these cells generate polyclonal T-cell response, while in the chronic phase of the disease the monoclonal or even an undetectable T-cell response will be generated (139, 140).

In the acute phase of the disease, CTLs contribute immunopathological effects of hepatitis B by producing cytokines, such as IFN γ and cytotoxic activity (141, 142). In general, it can be said that in patients with AHB, there is a disruption in the production of cytokines, such as type1 IFN I, IL15, and IFN γ . On the other hand, anti-inflammatory cytokines production, such as IL10 has been reported, which ultimately inhibits the immune system in the acute phase (138). Vitamin D seems to strengthen this pathway by creating its anti-inflammatory responses and immunomodulatory effect in this phase.

The chronic phase of hepatitis B disease can be divided into four categories, including: immune-tolerant, immune clearance, inactive HBs antigen carrier and reactivation phase. The specifications of each phase can be seen in the table 1 (143, 144).

In the chronic phase of hepatitis B, CTL cells have weaker performance than the acute phase of the disease (145). One of the causes of decreased CTL function in the immune-tolerant phase of chronic hepatitis B CHB may be an increase in the expression of inhibitory receptors, such as Cytotoxic T-lymphocyte antigen 4 (CTLA-4) and programmed cell death protein 1 (PD-1); they eventually cause the appearance of exhausted phenotype in these cells. (146-148). Studies have shown that vitamin D increases the expression of these two inhibitory receptors (149, 150). Other function of vitamin D against CTL cells, in the chronic phase of hepatitis B disease, studies have shown that CTL cell depletion occurs (151). Vitamin D can also inhibit the proliferation of CD8 cells (152). In the immune-tolerant phase, the number of CD4 + CD25 + FoxP3 + cells increases, which ultimately suppresses the antiviral responses. Many studies have shown the special effect of vitamin D on these cells. It increases the number and the immune-inhibitory function of these cells (153). One of the characteristics of the immune clearance phase in patients with chronic hepatitis is the increase of Th17 in the patients' liver and blood (154, 155). In patients with CHB, Th17 cells may

be related to immune activation and disease aggravation (156). Vitamin D has extensive inhibitory effects on Th17, which can be summarized as an inhibitory role of vitamin D in the differentiation, maintenance, transcription, and bioactivity processes (157).

In patients with CHB, the ratio of CD4+/CD8+ decreases (158). However, a study by Zhang et al. showed that vitamin D can affect this ratio. In the study of HBV infected mice, IFN and vitamin D treated mice had significantly lower levels of liver enzymes (ALT &AST) after treatment compared to mice, which did not receive vitamin D. The level of CD4 + and CD4 + / CD8 cells increases; it causes a balance between the two populations of CD + 4 and CD+ 8 cells. The levels of cytokines, such as IFN- γ , TNF- α , and IL-2 decrease, while IL- 4 did not show any differences between the two groups (159).

Innate immunity has a special role in the immune-pathogenesis of CHB; it is considered as a target for treatment (160). Many studies have shown that increased C-reactive protein (CRP) can lead to complications such as liver cirrhosis and fibrosis in the chronic phase of hepatitis B (161).

if the increase of vitamin D has a decreasing effect on CRP expression (162). In CHB patients, NK cells have inhibitory phenotypes (163), but vitamin D has a dual effect on them. It increases the expression of stimulant receptors, such as NKp44, NKp46, and NKp30, and also KLR (CD158) expression (164). Table2 showed clinical trial studies of vitamin D and hepatitis B infection.

Genetic studies polymorphisms

The association between vitamin D with viral hepatitis

Genetic studies polymorphisms

It is true that vitamin D levels can be an effective factor in disease pathogenesis, but various genetic factors can affect this process; they include the polymorphism of the involved enzymes and

receptors in the signaling pathway. The relationship between physiology of vitamin D and viral infections is also supported by genetic researches; they evaluated effective variants in the disease susceptibility, which encoding in the *VDR* gene.

Earlier studies revealed that the human VDR is a 75 kb nuclear receptor gene, found in the long arm of chromosome 12; it is composed of 11 introns and 11 exons (165). Vitamin D polymorphisms are different interindividual, and are dependent on the disease. Taiwan was the first country, which indicated the major effect of host genetic history on infection result (166). Furthermore, genetic variations in TNF and IFN, vitamin D receptor, estrogen receptor-1, and multiple HLA loci have been related to CHB in the subsequent studies (167, 168). Similarly, in various populations, HLA-DR 13 has been identified to have a protective role against chronic HBV infections. Recently, Kamatabi et al. showed that genetic variants of HLA-DP locus are substantially related to the risk of chronic HBV infection in Asian patients by genome-wide association strategy (169). The HLA-DP complexes are implicated in the presentation of antigen, and notably have been identified as predictive factors after HBV vaccination for antibody development (169). BsmI, TaqI, FokI and ApaI described by the endonuclease, are the most common studied SNP (170). Studies have shown that VDR gene polymorphism in rs731236 (Taq-1) (tt genotype) is significantly lower in patients with hepatitis B(171). Other studies have shown, that polymorphism rs2228570 (FokI) genotype ff makes people more susceptible to hepatitis B and also hepatocarcinoma (172, 173). Other studies have shown that rs7975232 (ApaI) polymorphism increases the liver damage risk caused by HBV (174); this polymorphism is also a prognostic factor in the effectiveness of treatment of hepatitis patients with Peg-IFN monotherapy (175, 176). In a study about the rs222020 variant, which is linked to the vitamin D-binding protein gene, Hbe-negative individuals who received Peg-IFN treatment were found to have normal levels of ALT liver enzymes, as well as lower Hbs antigen

levels (177). Gao and coworker demonstrated, that rs1540339, rs11168268, rs2239182, rs3819545, rs2239184, rs2239186 and rs7041 SNP sites were related to the susceptibility of healthy volunteers to the HBV infection (178); also, rs1800871, rs11168268, rs1544410, rs1800872, rs731236, rs3733359, rs1800896, and rs7041 polymorphisms were associated with hepatocellular carcinoma. In addition, four SNPs (rs2239184, rs2239186, rs2239181 and rs11168268) were meaningfully related to decompensation in cirrhotic cases. Two SNPs of rs2239184 and rs2239186 were significantly observed in all phases, except HBV infection to hepatocellular carcinoma (178). In chronic hepatitis B infection the frequency of polymorphism in TaqI, ApaI and BsmI alleles is related to HBeAg, directly(179). The polymorphism involved in the CYP2R1 (rs12794714) gene be involved in interferon-based therapies, known as PegIFN therapy Variation in ApaI, is often linked to increase HBV viral load and greater fibrosis and necroinflam mation (181). A study showed that TaqI VDR variation is often linked with both chronic HBV infection and occult HBV disease. In the cited study, negative HBsAg participants had lower probability of HBV load (182). Variability in the BsmI SNPs allele frequency is significantly related to primary biliary cirrhosis, while variability in FokI variants is linked with autoimmune hepatitis (183). TNF-α and TNF-β are formed by stimulated lymphocytes. They activate Nf-kB, and then induce hepatic fibrosis-related proinflammatory and hepatic inflammation genes (184, 185). The A/A genotype of TNF- β (sited in intron 2 of the gene) is more frequent in patients with severe liver problem due to chronic hepatitis B, compared to moderate liver diseases. In patients with chronic HCV, TNF-β A/A mutations have also been known to be associated with more serious liver fibrosis and problems (186, 187). According to the numerous reports, TaqI, FokI, ApaI, and BsmI VDR polymorphisms have association with HBV infection risk and HBV-related liver diseases progression. Several findings have focused on HBV pathogenesis and development of new

therapies. The recent discovery of genetic variants in the IL-28B locus, may predict hepatitis C

therapy for viral clearance (188, 189). These current discoveries could translate into genotype-based

therapy choices for HBV infected individual.

Conclusion

The relation between vitamin D physiologies with viral infections is also confirmed by genetic

research, carried out on genetic variations of VDR R-encoding disease susceptibility gene. Vitamin

D inhibits virus replication; it has an effective role in platelet and albumin levels and reducing ALT

enzyme levels in patients with acute hepatitis. In most HBV patients, vitamin D deficiency has been

identified especially in advanced liver diseases associated with adverse clinical outcomes.

However, Vitamin D can play different roles in diverse phases of disease. Therefore, it is suggested

to perform more researches in this field.

Funding: The authors did not receive support from any organization for the submitted work.

Financial interests: The authors declare they have no financial interests

Conflicts of interests: The authors declare no conflicts of interests

Ethics approval: This article does not contain any studies with animals performed by any of the

authors

Data, Material and/or Code availability: The article is a review and has no material.

Authors' Contribution Statements:

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results and contributed to the final manuscript

Fatemeh Jafari: Drafted or provided critical revision of the article, all authors discussed the

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Omid Kooshkaki: All authors discussed the results and contributed to the final manuscript

Alireza Abdshah: All authors discussed the results and contributed to the final manuscript

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Reference:

- 1. Cheng C-W, Feng C-M, Chua CS, editors. Help-Seeking Experiences of Hepatitis B Patients in Transnational Medical Care: The Solution to Health Inequality Is Social Mobility. Healthcare; 2019: Multidisciplinary Digital Publishing Institute.
- 2. Javanmard D, Alavian SM, Abedi F, Namaei MH, Asghari A, Ziaee M. High prevalence of hepatitis B virus infection in the Village of Esfandiar in South Khorasan Province, Iran. Hepatitis Monthly. 2018;18(8).
- 3. Magalhães MJ, Pedroto I. Hepatitis B virus inactive carriers: which follow-up strategy? GE Portuguese journal of gastroenterology. 2015;22(2):47-51.
- 4. Zhang D-m, Luo Y, Yishake D, Liu Z-y, He T-t, Luo Y, et al. Prediagnostic dietary intakes of vitamin A and β -carotene are associated with hepatocellular-carcinoma survival. Food & function. 2020;11(1):759-67.
- 5. Cui L-H, Quan Z-Y, Piao J-M, Zhang T-T, Jiang M-H, Shin M-H, et al. Plasma folate and vitamin B12 levels in patients with hepatocellular carcinoma. International journal of molecular sciences. 2016;17(7):1032.
- 6. Elkhateeb WA, Daba GM, El-Dein AN, Sheir DH, Fayad W, Shaheen MN, et al. Insights into the invitro hypocholesterolemic, antioxidant, antirotavirus, and anticolon cancer activities of the methanolic extracts of a Japanese lichen, Candelariella vitellina, and a Japanese mushroom, Ganoderma applanatum. Egyptian Pharmaceutical Journal. 2020;19(1):67.
- 7. Christakos S, Ajibade DV, Dhawan P, Fechner AJ, Mady LJ. Vitamin D: metabolism. Rheumatic Disease Clinics. 2012;38(1):1-11.
- 8. Naghizadeh M, Bahrami A, Mahavar N, Asghari A, Fereidouni M. Vitamin D and its association with allergic status and serum IgE. Revue Française d'Allergologie. 2019;59(6):427-33.
- Holick MF. Vitamin D deficiency. New England Journal of Medicine. 2007;357(3):266-81.
- 10. Heshmat R, Mohammad K, Majdzadeh S, Forouzanfar M, Bahrami A, Ranjbar Omrani G, et al. Vitamin D deficiency in Iran: A multi-center study among different urban areas. Iran J Public Health. 2008;37(1):72-8.
- 11. Christakos S, Hewison M, Gardner DG, Wagner CL, Sergeev IN, Rutten E, et al. Vitamin D: beyond bone. Annals of the New York Academy of Sciences. 2013;1287(1):45.
- 12. Medrano M, Carrillo-Cruz E, Montero I, Perez-Simon JA. Vitamin D: effect on haematopoiesis and immune system and clinical applications. International journal of molecular sciences. 2018;19(9):2663.
- 13. Chirumbolo S, Bjørklund G, Sboarina A, Vella A. The role of vitamin D in the immune system as a pro-survival molecule. Clinical therapeutics. 2017;39(5):894-916.
- 14. Zhao J-G, Zeng X-T, Wang J, Liu L. Association between calcium or vitamin D supplementation and fracture incidence in community-dwelling older adults: a systematic review and meta-analysis. Jama. 2017;318(24):2466-82.
- 15. Khammissa R, Fourie J, Motswaledi M, Ballyram R, Lemmer J, Feller L. The biological activities of vitamin D and its receptor in relation to calcium and bone homeostasis, cancer, immune and cardiovascular systems, skin biology, and oral health. BioMed research international. 2018;2018.
- 16. Wimalawansa SJ, Razzaque MS, Al-Daghri NM. Calcium and vitamin D in human health: Hype or real? The Journal of steroid biochemistry and molecular biology. 2018;180:4-14.
- 17. Telcian AG, Zdrenghea MT, Edwards MR, Laza-Stanca V, Mallia P, Johnston SL, et al. Vitamin D increases the antiviral activity of bronchial epithelial cells in vitro. Antiviral research. 2017;137:93-101.

- 18. Maggini S, Maldonado P, Cardim P, Fernandez Newball C, Sota Latino E. Vitamins C, D and zinc: synergistic roles in immune function and infections. Vitam Miner. 2017;6(167):2376-1318.1000167.
- 19. Ward LM, Gaboury I, Ladhani M, Zlotkin S. Vitamin D-deficiency rickets among children in Canada. Cmaj. 2007;177(2):161-6.
- 20. Parsamanesh N, Moossavi M, Tavakkoli T, Javdani H, Fakharian T, Moossavi SZ, et al. Positive correlation between vitamin D receptor gene Taql variant and gastric cancer predisposition in a sample of Iranian population. Journal of cellular physiology. 2019;234(9):15044-7.
- 21. Tuohimaa P. Vitamin D and aging. The Journal of steroid biochemistry and molecular biology. 2009;114(1-2):78-84.
- 22. Moossavi M, Parsamanesh N, Mohammadoo-Khorasani M, Moosavi M, Tavakkoli T, Fakharian T, et al. Positive correlation between vitamin D receptor gene Fokl polymorphism and colorectal cancer susceptibility in South-Khorasan of Iran. Journal of cellular biochemistry. 2018;119(10):8190-4.
- 23. Wong GL-H, Chan HL-Y, Chan H-Y, Tse C-H, Chim AM-L, Lo AO-S, et al. Adverse effects of vitamin D deficiency on outcomes of patients with chronic hepatitis B. Clinical Gastroenterology and Hepatology. 2015;13(4):783-90. e1.
- 24. Said E, El Agawy W, Ahmed R, Hassany M, Ahmed A, Fouad H, et al. Serum vitamin D levels in treatment-naïve chronic hepatitis B patients. Journal of translational internal medicine. 2017;5(4):230-4.
- 25. Stokes CS, Volmer DA, Grünhage F, Lammert F. Vitamin D in chronic liver disease. Liver International. 2013;33(3):338-52.
- 26. Trépo E, Ouziel R, Pradat P, Momozawa Y, Quertinmont E, Gervy C, et al. Marked 25-hydroxyvitamin D deficiency is associated with poor prognosis in patients with alcoholic liver disease. Journal of hepatology. 2013;59(2):344-50.
- 27. Finkelmeier F, Kronenberger B, Zeuzem S, Piiper A, Waidmann O. Low 25-hydroxyvitamin D levels are associated with infections and mortality in patients with cirrhosis. PloS one. 2015;10(6):e0132119.
- 28. Paternostro R, Wagner D, Reiberger T, Mandorfer M, Schwarzer R, Ferlitsch M, et al. Low 25-OH-vitamin D levels reflect hepatic dysfunction and are associated with mortality in patients with liver cirrhosis. Wiener Klinische Wochenschrift. 2017;129(1-2):8-15.
- 29. Mousa A, Naderpoor N, de Courten MP, Teede H, Kellow N, Walker K, et al. Vitamin D supplementation has no effect on insulin sensitivity or secretion in vitamin D-deficient, overweight or obese adults: a randomized placebo-controlled trial. Am J Clin Nutr. 2017;105(6):1372-81.
- 30. Antico A, Tampoia M, Tozzoli R, Bizzaro N. Can supplementation with vitamin D reduce the risk or modify the course of autoimmune diseases? A systematic review of the literature. Autoimmunity reviews. 2012;12(2):127-36.
- 31. Atkinson MA, Eisenbarth GS, Michels AW. Type 1 diabetes. The Lancet. 2014;383(9911):69-82.
- 32. Maretzke F, Bechthold A, Egert S, Ernst JB, Melo van Lent D, Pilz S, et al. Role of vitamin D in preventing and treating selected extraskeletal diseases—an umbrella review. Nutrients. 2020;12(4):969.
- 33. Townsend K, Evans KN, Campbell MJ, Colston KW, Adams JS, Hewison M. Biological actions of extra-renal 25-hydroxyvitamin D- 1α -hydroxylase and implications for chemoprevention and treatment. The Journal of steroid biochemistry and molecular biology. 2005;97(1-2):103-9.
- 34. Christakos S, Dhawan P, Verstuyf A, Verlinden L, Carmeliet G. Vitamin D: metabolism, molecular mechanism of action, and pleiotropic effects. Physiological reviews. 2016;96(1):365-408.
- 35. Garland CF, Kim JJ, Mohr SB, Gorham ED, Grant WB, Giovannucci EL, et al. Meta-analysis of all-cause mortality according to serum 25-hydroxyvitamin D. American journal of public health. 2014;104(8):e43-e50.
- 36. Wimalawansa SJ. Non-musculoskeletal benefits of vitamin D. The Journal of steroid biochemistry and molecular biology. 2018;175:60-81.

- 37. McDonnell SL, Baggerly C, French CB, Baggerly LL, Garland CF, Gorham ED, et al. Serum 25-hydroxyvitamin D concentrations≥ 40 ng/ml are associated with> 65% lower cancer risk: pooled analysis of randomized trial and prospective cohort study. PloS one. 2016;11(4):e0152441.
- 38. Spedding S, Vanlint S, Morris H, Scragg R. Does vitamin D sufficiency equate to a single serum 25-hydroxyvitamin D level or are different levels required for non-skeletal diseases? Nutrients. 2013;5(12):5127-39.
- 39. Płudowski P, Karczmarewicz E, Bayer M, Carter G, Chlebna-Sokół D, Czech-Kowalska J, et al. Practical guidelines for the supplementation of vitamin D and the treatment of deficits in Central Europe—recommended vitamin D intakes in the general population and groups at risk of vitamin D deficiency. Endokrynologia Polska. 2013;64(4):319-27.
- 40. Mellati AA, Sharifi F, Faghihzade S, Mousaviviri SA, Chiti H, Kazemi SAN. Vitamin D status and its associations with components of metabolic syndrome in healthy children. Journal of Pediatric Endocrinology and Metabolism. 2015;28(5-6):641-8.
- 41. Soltani Z, Khamse M, Chiti H, Valizadeh M, Mazloomzadeh S. Plasma 25 (OH) vitamin-D level and metabolic syndrome risk factors among physicians of Zanjan. J Adv Med Biomed Res. 2015;23(99):64-73.
- 42. Dabai NS, Pramyothin P, Holick MF. The effect of ultraviolet radiation from a novel portable fluorescent lamp on serum 25-hydroxyvitamin D3 levels in healthy adults with F itzpatrick skin types II and III. Photodermatology, photoimmunology & photomedicine. 2012;28(6):307-11.
- 43. Manson J, Brannon PM, Rosen CJ, Taylor CL. Vitamin D deficiency-is there really a pandemic. N Engl J Med. 2016;375(19):1817-20.
- 44. Wacker M, Holick MF. Vitamin D—effects on skeletal and extraskeletal health and the need for supplementation. Nutrients. 2013;5(1):111-48.
- 45. Holick MF, Binkley NC, Bischoff-Ferrari HA, Gordon CM, Hanley DA, Heaney RP, et al. Evaluation, treatment, and prevention of vitamin D deficiency: an Endocrine Society clinical practice guideline. The Journal of Clinical Endocrinology & Metabolism. 2011;96(7):1911-30.
- 46. Ross AC, Manson JE, Abrams SA, Aloia JF, Brannon PM, Clinton SK, et al. The 2011 report on dietary reference intakes for calcium and vitamin D from the Institute of Medicine: what clinicians need to know. The Journal of Clinical Endocrinology & Metabolism. 2011;96(1):53-8.
- 47. Holick M. Vitamin D update 2015: What we need to know about its health benefits and potential for toxicity. Standardy medyczne pediatria. 2015;12(5):759-63.
- 48. Vairetti M, Di Pasqua LG, Cagna M, Richelmi P, Ferrigno A, Berardo C. Changes in Glutathione Content in Liver Diseases: An Update. Antioxidants. 2021;10(3):364.
- 49. Cazanave S, Berson A, Haouzi D, Vadrot N, Fau D, Grodet A, et al. High hepatic glutathione stores alleviate Fas-induced apoptosis in mice. Journal of hepatology. 2007;46(5):858-68.
- 50. Qian L, Wang W, Zhou Y, Ma J. Effects of reduced glutathione therapy on chronic hepatitis B. Central-European journal of immunology. 2017;42(1):97.
- 51. Parsanathan R, Jain SK. Glutathione deficiency alters the vitamin D-metabolizing enzymes CYP27B1 and CYP24A1 in human renal proximal tubule epithelial cells and kidney of HFD-fed mice. Free Radical Biology and Medicine. 2019;131:376-81.
- 52. Jain SK, Parsanathan R, Achari AE, Kanikarla-Marie P, Bocchini Jr JA. Glutathione stimulates vitamin D regulatory and glucose-metabolism genes, lowers oxidative stress and inflammation, and increases 25-hydroxy-vitamin D levels in blood: a novel approach to treat 25-hydroxyvitamin D deficiency. Antioxidants & redox signaling. 2018;29(17):1792-807.
- 53. Jain SK, Kanikarla-Marie P, Warden C, Micinski D. L-cysteine supplementation upregulates glutathione (GSH) and vitamin D binding protein (VDBP) in hepatocytes cultured in high glucose and in vivo in liver, and increases blood levels of GSH, VDBP, and 25-hydroxy-vitamin D in Zucker diabetic fatty rats. Molecular nutrition & food research. 2016;60(5):1090-8.

- 54. Jain SK, Micinski D, Parsanathan R. I-Cysteine Stimulates the Effect of Vitamin D on Inhibition of Oxidative Stress, IL-8, and MCP-1 Secretion in High Glucose Treated Monocytes. Journal of the American College of Nutrition. 2021;40(4):327-32.
- 55. Jain SK, Parsanathan R, Levine SN, Bocchini JA, Holick MF, Vanchiere JA. The potential link between inherited G6PD deficiency, oxidative stress, and vitamin D deficiency and the racial inequities in mortality associated with COVID-19. Free Radical Biology and Medicine. 2020;161:84-91.
- 56. Parsamanesh N, Pezeshgi A, Hemmati M, Jameshorani M, Saboory E. Neurological manifestations of coronavirus infections: role of angiotensin-converting enzyme 2 in COVID-19. International Journal of Neuroscience. 2020:1-8.
- 57. Asghari A, Naseri M, Safari H, Saboory E, Parsamanesh N. The novel insight of SARS-CoV-2 molecular biology and pathogenesis and therapeutic options. DNA and Cell Biology. 2020;39(10):1741-53.
- 58. Amiri-Dashatan N, Koushki M, Parsamanesh N, Chiti H. Serum cortisol concentration and COVID-19 severity: a systematic review and meta-analysis. Journal of Investigative Medicine. 2022;70(3):766-72.
- 59. Yıldız B. Investigation of the role of cGAMP in differentiation of T lymphocytes: Bilkent University; 2016.
- 60. Omdahl JL, Morris HA, May BK. Hydroxylase enzymes of the vitamin D pathway: expression, function, and regulation. Annual review of nutrition. 2002;22(1):139-66.
- 61. Morán-Auth Y, Penna-Martinez M, Shoghi F, Ramos-Lopez E, Badenhoop K. Vitamin D status and gene transcription in immune cells. The Journal of steroid biochemistry and molecular biology. 2013;136:83-5.
- 62. Smolders J, Thewissen M, Theunissen R, Peelen E, Knippenberg S, Menheere P, et al. Vitamin D-related gene expression profiles in immune cells of patients with relapsing remitting multiple sclerosis. Journal of neuroimmunology. 2011;235(1-2):91-7.
- 63. Adams JS, Sharma OP, Gacad MA, Singer FR. Metabolism of 25-hydroxyvitamin D3 by cultured pulmonary alveolar macrophages in sarcoidosis. The Journal of clinical investigation. 1983;72(5):1856-60.
- 64. Omdahl JL, Morris HA, May BK. Hydroxylase enzymes of the vitamin D pathway: expression, function, and regulation. Annu Rev Nutr. 2002;22:139-66.
- 65. Wu S, Ren S, Nguyen L, Adams JS, Hewison M. Splice variants of the CYP27b1 gene and the regulation of 1,25-dihydroxyvitamin D3 production. Endocrinology. 2007;148(7):3410-8.
- 66. van Etten E, Stoffels K, Gysemans C, Mathieu C, Overbergh L. Regulation of vitamin D homeostasis: implications for the immune system. Nutrition reviews. 2008;66(10 Suppl 2):S125-34.
- 67. Parsamanesh N, Karami-Zarandi M, Banach M, Penson PE, Sahebkar A. Effects of statins on myocarditis: a review of underlying molecular mechanisms. Progress in cardiovascular diseases. 2021;67:53-64.
- 68. Ghorbani-Abdi-Saedabad A, Hanafi-Bojd MY, Parsamanesh N, Tayarani-Najaran Z, Mollaei H, Hoshyar R. Anticancer and apoptotic activities of parthenolide in combination with epirubicin in mdamb-468 breast cancer cells. Molecular Biology Reports. 2020;47(8):5807-15.
- 69. Aranow C. Vitamin D and the immune system. Journal of investigative medicine: the official publication of the American Federation for Clinical Research. 2011;59(6):881-6.
- 70. Azrielant S SY. Vitamin D and the immune system.2017;19(8):510-1.
- 71. Moen SH, Ehrnström B, Kojen JF, Yurchenko M, Beckwith KS, Afset JE, et al. Human Toll-like Receptor 8 (TLR8) Is an Important Sensor of Pyogenic Bacteria, and Is Attenuated by Cell Surface TLR Signaling. Frontiers in immunology. 2019;10:1209.
- 72. Kawai T, Akira S. Innate immune recognition of viral infection. Nature Immunology. 2006;7(2):131-7.

- 73. Do JE, Kwon SY, Park S, Lee ES. Effects of vitamin D on expression of Toll-like receptors of monocytes from patients with Behcet's disease. Rheumatology (Oxford, England). 2008;47(6):840-8.
- 74. Berzsenyi MD, Roberts SK, Preiss S, Woollard DJ, Beard MR, Skinner NA, et al. Hepatic TLR2 & TLR4 expression correlates with hepatic inflammation and TNF- α in HCV & HCV/HIV infection. Journal of viral hepatitis. 2011;18(12):852-60.
- 75. Adamczak DM. The Role of Toll-Like Receptors and Vitamin D in Cardiovascular Diseases-A Review. Int J Mol Sci. 2017;18(11).
- 76. Alvarez-Rodriguez L, Lopez-Hoyos M, Garcia-Unzueta M, Amado JA, Cacho PM, Martinez-Taboada VM. Age and low levels of circulating vitamin D are associated with impaired innate immune function. Journal of leukocyte biology. 2012;91(5):829-38.
- 77. Djukic M, Onken ML, Schütze S, Redlich S, Götz A, Hanisch UK, et al. Vitamin d deficiency reduces the immune response, phagocytosis rate, and intracellular killing rate of microglial cells. Infection and immunity. 2014;82(6):2585-94.
- 78. Ojaimi S, Skinner NA, Strauss BJ, Sundararajan V, Woolley I, Visvanathan K. Vitamin D deficiency impacts on expression of toll-like receptor-2 and cytokine profile: a pilot study. Journal of translational medicine. 2013;11:176.
- 79. Gwyer Findlay E, Currie SM, Davidson DJ. Cationic host defence peptides: potential as antiviral therapeutics. BioDrugs: clinical immunotherapeutics, biopharmaceuticals and gene therapy. 2013;27(5):479-93.
- 80. Diamond G, Beckloff N, Weinberg A, Kisich KO. The roles of antimicrobial peptides in innate host defense. Current pharmaceutical design. 2009;15(21):2377-92.
- 81. Telcian AG, Zdrenghea MT, Edwards MR, Laza-Stanca V, Mallia P, Johnston SL, et al. Vitamin D increases the antiviral activity of bronchial epithelial cells in vitro. Antiviral Research. 2017;137:93-101.
- 82. Sousa FH, Casanova V, Findlay F, Stevens C, Svoboda P, Pohl J, et al. Cathelicidins display conserved direct antiviral activity towards rhinovirus. Peptides. 2017;95:76-83.
- 83. Currie SM, Gwyer Findlay E, McFarlane AJ, Fitch PM, Böttcher B, Colegrave N, et al. Cathelicidins Have Direct Antiviral Activity against Respiratory Syncytial Virus In Vitro and Protective Function In Vivo in Mice and Humans. Journal of immunology (Baltimore, Md: 1950). 2016;196(6):2699-710.
- 84. Takahashi T, Kulkarni NN, Lee EY, Zhang L-j, Wong GCL, Gallo RL. Cathelicidin promotes inflammation by enabling binding of self-RNA to cell surface scavenger receptors. Scientific Reports. 2018;8(1):4032.
- 85. Lai Y, Adhikarakunnathu S, Bhardwaj K, Ranjith-Kumar CT, Wen Y, Jordan JL, et al. LL37 and cationic peptides enhance TLR3 signaling by viral double-stranded RNAs. PLoS One. 2011;6(10):e26632.
- 86. Ramos-Martínez E, López-Vancell MR, Fernández de Córdova-Aguirre JC, Rojas-Serrano J, Chavarría A, Velasco-Medina A, et al. Reduction of respiratory infections in asthma patients supplemented with vitamin D is related to increased serum IL-10 and IFNγ levels and cathelicidin expression. Cytokine. 2018;108:239-46.
- 87. Liu PT, Stenger S, Tang DH, Modlin RL. Cutting edge: vitamin D-mediated human antimicrobial activity against Mycobacterium tuberculosis is dependent on the induction of cathelicidin. Journal of immunology (Baltimore, Md: 1950). 2007;179(4):2060-3.
- 88. Wang TT, Nestel FP, Bourdeau V, Nagai Y, Wang Q, Liao J, et al. Cutting edge: 1,25-dihydroxyvitamin D3 is a direct inducer of antimicrobial peptide gene expression. Journal of immunology (Baltimore, Md: 1950). 2004;173(5):2909-12.
- 89. Büchau AS, Schauber J, Hultsch T, Stuetz A, Gallo RL. Pimecrolimus enhances TLR2/6-induced expression of antimicrobial peptides in keratinocytes. The Journal of investigative dermatology. 2008;128(11):2646-54.

- 90. Aguilar-Jiménez W, Zapata W, Caruz A, Rugeles MT. High transcript levels of vitamin D receptor are correlated with higher mRNA expression of human beta defensins and IL-10 in mucosa of HIV-1-exposed seronegative individuals. PLoS One. 2013;8(12):e82717.
- 91. Cantorna MT, Snyder L, Lin YD, Yang L. Vitamin D and 1,25(OH)2D regulation of T cells. Nutrients. 2015;7(4):3011-21.
- 92. Lemire JM, Adams JS, Kermani-Arab V, Bakke AC, Sakai R, Jordan SC. 1,25-Dihydroxyvitamin D3 suppresses human T helper/inducer lymphocyte activity in vitro. Journal of immunology (Baltimore, Md: 1950). 1985;134(5):3032-5.
- 93. Joshi S, Pantalena LC, Liu XK, Gaffen SL, Liu H, Rohowsky-Kochan C, et al. 1,25-dihydroxyvitamin D(3) ameliorates Th17 autoimmunity via transcriptional modulation of interleukin-17A. Molecular and cellular biology. 2011;31(17):3653-69.
- 94. Boonstra A, Barrat FJ, Crain C, Heath VL, Savelkoul HF, O'Garra A. 1alpha,25-Dihydroxyvitamin d3 has a direct effect on naive CD4(+) T cells to enhance the development of Th2 cells. Journal of immunology (Baltimore, Md: 1950). 2001;167(9):4974-80.
- 95. Jeffery LE, Burke F, Mura M, Zheng Y, Qureshi OS, Hewison M, et al. 1,25-Dihydroxyvitamin D3 and IL-2 combine to inhibit T cell production of inflammatory cytokines and promote development of regulatory T cells expressing CTLA-4 and FoxP3. Journal of immunology (Baltimore, Md: 1950). 2009;183(9):5458-67.
- 96. Adorini L, Penna G. Induction of tolerogenic dendritic cells by vitamin D receptor agonists. Handbook of experimental pharmacology. 2009(188):251-73.
- 97. Khoo AL, Chai LY, Koenen HJ, Sweep FC, Joosten I, Netea MG, et al. Regulation of cytokine responses by seasonality of vitamin D status in healthy individuals. Clinical and experimental immunology. 2011;164(1):72-9.
- 98. von Essen MR, Kongsbak M, Schjerling P, Olgaard K, Odum N, Geisler C. Vitamin D controls T cell antigen receptor signaling and activation of human T cells. Nat Immunol. 2010;11(4):344-9.
- 99. Anand SP, Selvaraj P, Narayanan PJC. Effect of 1, 25 dihydroxyvitamin D3 on intracellular IFN- γ and TNF- α positive T cell subsets in pulmonary tuberculosis. 2009;45(2):105-10.
- 100. Lysandropoulos AP, Jaquiéry E, Jilek S, Pantaleo G, Schluep M, Du Pasquier RAJJon. Vitamin D has a direct immunomodulatory effect on CD8+ T cells of patients with early multiple sclerosis and healthy control subjects. 2011;233(1-2):240-4.
- 101. Prabhu Anand S, Selvaraj P, Narayanan PR. Effect of 1,25 dihydroxyvitamin D3 on intracellular IFN-gamma and TNF-alpha positive T cell subsets in pulmonary tuberculosis. Cytokine. 2009;45(2):105-10.
- 102. Chen S, Sims GP, Chen XX, Gu YY, Chen S, Lipsky PE. Modulatory effects of 1,25-dihydroxyvitamin D3 on human B cell differentiation. Journal of immunology (Baltimore, Md: 1950). 2007;179(3):1634-47.
- 103. Lemire JM, Adams JS, Sakai R, Jordan SC. 1 alpha,25-dihydroxyvitamin D3 suppresses proliferation and immunoglobulin production by normal human peripheral blood mononuclear cells. The Journal of clinical investigation. 1984;74(2):657-61.
- 104. Peelen E, Rijkers G, Meerveld-Eggink A, Meijvis S, Vogt M, Cohen Tervaert JW, et al. Relatively high serum vitamin D levels do not impair the antibody response to encapsulated bacteria. European journal of clinical microbiology & infectious diseases: official publication of the European Society of Clinical Microbiology. 2013;32(1):61-9.
- 105. Hoan NX, Khuyen N, Binh MT, Giang DP, Van Tong H, Hoan PQ, et al. Association of vitamin D deficiency with hepatitis B virus related liver diseases. BMC infectious diseases. 2016;16(1):507.
- 106. Chen EQ, Bai L, Zhou TY, Fe M, Zhang DM, Tang H. Sustained suppression of viral replication in improving vitamin D serum concentrations in patients with chronic hepatitis B. Sci Rep. 2015;5:15441.

- 107. Farnik H, Bojunga J, Berger A, Allwinn R, Waidmann O, Kronenberger B, et al. Low vitamin D serum concentration is associated with high levels of hepatitis B virus replication in chronically infected patients. Hepatology (Baltimore, Md). 2013;58(4):1270-6.
- 108. Mohamadkhani A, Bastani F, Khorrami S, Ghanbari R, Eghtesad S, Sharafkhah M, et al. Negative Association of Plasma Levels of Vitamin D and miR-378 With Viral Load in Patients With Chronic Hepatitis B Infection. Hepatitis monthly. 2015;15(6):e28315.
- 109. Chan HL, Elkhashab M, Trinh H, Tak WY, Ma X, Chuang WL, et al. Association of baseline vitamin D levels with clinical parameters and treatment outcomes in chronic hepatitis B. Journal of hepatology. 2015;63(5):1086-92.
- 110. Targher G, Bertolini L, Scala L, Cigolini M, Zenari L, Falezza G, et al. Associations between serum 25-hydroxyvitamin D3 concentrations and liver histology in patients with non-alcoholic fatty liver disease. Nutrition, metabolism, and cardiovascular diseases: NMCD. 2007;17(7):517-24.
- 111. Wong GL, Chan HL, Chan HY, Tse CH, Chim AM, Lo AO, et al. Adverse effects of vitamin D deficiency on outcomes of patients with chronic hepatitis B. Clinical gastroenterology and hepatology: the official clinical practice journal of the American Gastroenterological Association. 2015;13(4):783-90.e1.
- 112. Hormati A, Jameshorani M, Sarkeshikian SS, Molaei M, Jahangiri M, Ghadir MR. Effects of Atorvastatin Alongside Conventional Medical Treatment on Liver Fibrosis and Dysfunction in Patients with Chronic Hepatitis B: A Double-Blinded Clinical Trial. Hepatitis Monthly. 2019;19(2).
- 113. Ebadi M, Bhanji RA, Mazurak VC, Lytvyak E, Mason A, Czaja AJ, et al. Severe vitamin D deficiency is a prognostic biomarker in autoimmune hepatitis. Alimentary Pharmacology & Therapeutics. 2019;49(2):173-82.
- 114. Wong GL-H, Chan HL-Y, Chan H-Y, Tse C-H, Chim AM-L, Lo AO-S, et al. Adverse Effects of Vitamin D Deficiency on Outcomes of Patients With Chronic Hepatitis B. Clinical Gastroenterology and Hepatology. 2015;13(4):783-90.e1.
- 115. Gotlieb N, Tachlytski I, Lapidot Y, Sultan M, Safran M, Ben-Ari Z. Hepatitis B virus downregulates vitamin D receptor levels in hepatoma cell lines, thereby preventing vitamin D-dependent inhibition of viral transcription and production. Molecular medicine (Cambridge, Mass). 2018;24(1):53.
- 116. Berkan-Kawińska A, Koślińska-Berkan E, Piekarska A. Original article
br>The prevalence and severity of 25-(OH)-vitamin D insufficiency in HCV infected and in HBV infected patients: a prospective study. 2015;1(1):5-11.
- 117. Parfieniuk-Kowerda A, Świderska M, Rogalska M, Maciaszek M, Jaroszewicz J, Flisiak R. Chronic hepatitis B virus infection is associated with decreased serum 25(OH)D concentration in non-cirrhotic patients. 2019;5(1):75-80.
- 118. Zhao XY, Li J, Wang JH, Habib S, Wei W, Sun SJ, et al. Vitamin D serum level is associated with Child-Pugh score and metabolic enzyme imbalances, but not viral load in chronic hepatitis B patients. Medicine. 2016;95(27):e3926.
- 119. Zhu H, Liu X, Ding Y, Zhou H, Wang Y, Zhou Z, et al. Relationships between low serum vitamin D levels and HBV "a" determinant mutations in chronic hepatitis B patients. Journal of infection in developing countries. 2016;10(9):1025-30.
- 120. Yu R, Sun J, Zheng Z, Chen J, Fan R, Liang X, et al. Association between vitamin D level and viral load or fibrosis stage in chronic hepatitis B patients from Southern China. Journal of gastroenterology and hepatology. 2015;30(3):566-74.
- 121. Ghadir MR, Sarkeshikian SS, Hormati A, Alemi F, Alavinejad P, Jameshorani M, et al. Efficacy of Atorvastatin Plus Aspirin in Comparison with Atorvastatin Alone on Liver Function and Degree of Fibrosis in Patients with Cryptogenic Cirrhosis: A Randomized Double-Blind Clinical Trial. Hepatitis Monthly. 2019;19(11).

- 122. Osmani F, Ziaee M. Relationship between levels of vitamin D3 and hepatitis B: A case-control Study. International Journal of Infectious Diseases. 2020;101:333.
- 123. Tseng TC, Liu CJ, Chen CL, Wang CC, Su TH, Kuo SF, et al. Serum hepatitis B virus-DNA levels correlate with long-term adverse outcomes in spontaneous hepatitis B e antigen seroconverters. The Journal of infectious diseases. 2012;205(1):54-63.
- 124. Liu CJ, Chen PJ, Lai MY, Kao JH, Chang CF, Wu HL, et al. A prospective study characterizing full-length hepatitis B virus genomes during acute exacerbation. Gastroenterology. 2003;124(1):80-90.
- 125. Rehermann B. Pathogenesis of chronic viral hepatitis: differential roles of T cells and NK cells. Nature medicine. 2013;19(7):859-68.
- 126. McBride WJH. Mandell, Douglas and Bennett's Principles and Practice of Infectious Diseases 7th edition. CSIRO; 2010.
- 127. Kasper DL, Fauci AS. Harrison's Infectious Diseases. By.
- 128. Suresh M, Czerwinski S, Murreddu MG, Kallakury BV, Ramesh A, Gudima SO, et al. Innate and adaptive immunity associated with resolution of acute woodchuck hepatitis virus infection in adult woodchucks. PLoS pathogens. 2019;15(12):e1008248.
- 129. Suresh M, Czerwinski S, Murreddu MG, Kallakury BV, Ramesh A, Gudima SO, et al. Innate and adaptive immunity associated with resolution of acute woodchuck hepatitis virus infection in adult woodchucks. PLoS pathogens. 2019;15(12):e1008248-e.
- 130. Cooper A, Tal G, Lider O, Shaul Y. Cytokine induction by the hepatitis B virus capsid in macrophages is facilitated by membrane heparan sulfate and involves TLR2. Journal of immunology (Baltimore, Md: 1950). 2005;175(5):3165-76.
- 131. Gotlieb N, Tachlytski I, Lapidot Y, Sultan M, Safran M, Ben-Ari ZJMM. Hepatitis B virus downregulates vitamin D receptor levels in hepatoma cell lines, thereby preventing vitamin D-dependent inhibition of viral transcription and production. 2018;24(1):1-8.
- 132. Merkle M, Pircher J, Mannell H, Krötz F, Blüm P, Czermak T, et al. LL37 inhibits the inflammatory endothelial response induced by viral or endogenous DNA. Journal of Autoimmunity. 2015;65:19-29.
- 133. Iacob SA, Panaitescu E, Iacob DG, Cojocaru M. The human cathelicidin LL37 peptide has high plasma levels in B and C hepatitis related to viral activity but not to 25-hydroxyvitamin D plasma level. Romanian journal of internal medicine = Revue roumaine de medecine interne. 2012;50(3):217-23.
- 134. Yu S, Cantorna MT. The vitamin D receptor is required for iNKT cell development. Proceedings of the National Academy of Sciences of the United States of America. 2008;105(13):5207-12.
- 135. Yu S, Cantorna MT. Epigenetic reduction in invariant NKT cells following in utero vitamin D deficiency in mice. Journal of immunology (Baltimore, Md: 1950). 2011;186(3):1384-90.
- 136. Bendelac A, Savage PB, Teyton L. The biology of NKT cells. Annual review of immunology. 2007;25:297-336.
- 137. Li J, Han Y, Jin K, Wan Y, Wang S, Liu B, et al. Dynamic changes of cytotoxic T lymphocytes (CTLs), natural killer (NK) cells, and natural killer T (NKT) cells in patients with acute hepatitis B infection. Virol J [Internet]. 2011 2011/05//; 8:[199 p.]. Available from: http://europepmc.org/abstract/MED/21535873

https://www.ncbi.nlm.nih.gov/pmc/articles/pmid/21535873/?tool=EBI

https://www.ncbi.nlm.nih.gov/pmc/articles/pmid/21535873/pdf/?tool=EBI

https://doi.org/10.1186/1743-422X-8-199

https://europepmc.org/articles/PMC3096949

https://europepmc.org/articles/PMC3096949?pdf=render.

- 138. Dunn C, Peppa D, Khanna P, Nebbia G, Jones M, Brendish N, et al. Temporal analysis of early immune responses in patients with acute hepatitis B virus infection. Gastroenterology. 2009;137(4):1289-300.
- 139. Jung MC, Spengler U, Schraut W, Hoffmann R, Zachoval R, Eisenburg J, et al. Hepatitis B virus antigen-specific T-cell activation in patients with acute and chronic hepatitis B. Journal of hepatology. 1991;13(3):310-7.
- 140. Tseng T-C, Huang L-R. Immunopathogenesis of Hepatitis B Virus. The Journal of infectious diseases. 2017;216(suppl_8):S765-S70.
- 141. Tan A, Koh S, Bertoletti A. Immune Response in Hepatitis B Virus Infection. Cold Spring Harbor perspectives in medicine. 2015;5(8):a021428.
- 142. Shin E-C, Sung PS, Park S-H. Immune responses and immunopathology in acute and chronic viral hepatitis. Nature Reviews Immunology. 2016;16(8):509-23.
- 143. Tseng TC, Kao JH. Clinical utility of quantitative HBsAg in natural history and nucleos(t)ide analogue treatment of chronic hepatitis B: new trick of old dog. Journal of gastroenterology. 2013;48(1):13-21.
- 144. Liaw YF, Chu CM. Hepatitis B virus infection. Lancet (London, England). 2009;373(9663):582-92.
- 145. Kennedy PTF, Sandalova E, Jo J, Gill U, Ushiro-Lumb I, Tan AT, et al. Preserved T-cell function in children and young adults with immune-tolerant chronic hepatitis B. Gastroenterology. 2012;143(3):637-45.
- 146. Fisicaro P, Valdatta C, Massari M, Loggi E, Biasini E, Sacchelli L, et al. Antiviral intrahepatic T-cell responses can be restored by blocking programmed death-1 pathway in chronic hepatitis B. Gastroenterology. 2010;138(2):682-93, 93.e1-4.
- 147. Raziorrouh B, Schraut W, Gerlach T, Nowack D, Grüner NH, Ulsenheimer A, et al. The immunoregulatory role of CD244 in chronic hepatitis B infection and its inhibitory potential on virus-specific CD8+ T-cell function. Hepatology (Baltimore, Md). 2010;52(6):1934-47.
- 148. Schurich A, Khanna P, Lopes AR, Han KJ, Peppa D, Micco L, et al. Role of the coinhibitory receptor cytotoxic T lymphocyte antigen-4 on apoptosis-Prone CD8 T cells in persistent hepatitis B virus infection. Hepatology (Baltimore, Md). 2011;53(5):1494-503.
- 149. Sharifi A, Vahedi H, Honarvar MR, Alipoor B, Nikniaz Z, Rafiei H, et al. Vitamin D Increases CTLA-4 Gene Expression in Patients with Mild to Moderate Ulcerative Colitis. Middle East J Dig Dis. 2019;11(4):199-204.
- 150. Jeffery LE, Qureshi OS, Gardner D, Hou TZ, Briggs Z, Soskic B, et al. Vitamin D Antagonises the Suppressive Effect of Inflammatory Cytokines on CTLA-4 Expression and Regulatory Function. PloS one. 2015;10(7):e0131539-e.
- 151. Peppa D, Gill US, Reynolds G, Easom NJ, Pallett LJ, Schurich A, et al. Up-regulation of a death receptor renders antiviral T cells susceptible to NK cell-mediated deletion. The Journal of experimental medicine. 2013;210(1):99-114.
- 152. Karkeni E, Morin SO, Bou Tayeh B, Goubard A, Josselin E, Castellano R, et al. Vitamin D Controls Tumor Growth and CD8+ T Cell Infiltration in Breast Cancer. Frontiers in immunology. 2019;10:1307.
- 153. Fisher SA, Rahimzadeh M, Brierley C, Gration B, Doree C, Kimber CE, et al. The role of vitamin D in increasing circulating T regulatory cell numbers and modulating T regulatory cell phenotypes in patients with inflammatory disease or in healthy volunteers: A systematic review. PLoS One. 2019;14(9):e0222313.
- 154. Zhang JY, Zhang Z, Lin F, Zou ZS, Xu RN, Jin L, et al. Interleukin-17-producing CD4(+) T cells increase with severity of liver damage in patients with chronic hepatitis B. Hepatology (Baltimore, Md). 2010;51(1):81-91.

- 155. Zhang GL, Xie DY, Lin BL, Xie C, Ye YN, Peng L, et al. Imbalance of interleukin-17-producing CD4 T cells/regulatory T cells axis occurs in remission stage of patients with hepatitis B virus-related acute-on-chronic liver failure. Journal of gastroenterology and hepatology. 2013;28(3):513-21.
- 156. Yang B, Wang Y, Zhao C, Yan W, Che H, Shen C, et al. Increased Th17 cells and interleukin-17 contribute to immune activation and disease aggravation in patients with chronic hepatitis B virus infection. Immunology letters. 2013;149(1-2):41-9.
- 157. Dankers W, Davelaar N, van Hamburg JP, van de Peppel J, Colin EM, Lubberts E. Human Memory Th17 Cell Populations Change Into Anti-inflammatory Cells With Regulatory Capacity Upon Exposure to Active Vitamin D. Frontiers in immunology. 2019;10:1504.
- 158. You J, Sriplung H, Geater A, Chongsuvivatwong V, Zhuang L, Chen HY, et al. Effect of viral load on T-lymphocyte failure in patients with chronic hepatitis B. World journal of gastroenterology. 2008;14(7):1112-9.
- 159. Bi Y, Zhang W, Sun J, Gao RJIJol, Pharmacology. Therapeutic and immune function improvement of vitamin D combined with IFN- α on mouse with hepatitis B infection. 2018;32:2058738418775250.
- 160. Meng Z, Chen Y, Lu M. Advances in Targeting the Innate and Adaptive Immune Systems to Cure Chronic Hepatitis B Virus Infection. Frontiers in Immunology. 2020;10:3127.
- 161. Ma LN, Liu XY, Luo X, Hu YC, Liu SW, Tang YY, et al. Serum high-sensitivity C-reactive protein are associated with HBV replication, liver damage and fibrosis in patients with chronic hepatitis B. Hepatogastroenterology. 2015;62(138):368-72.
- 162. Kruit A, Zanen P. The association between vitamin D and C-reactive protein levels in patients with inflammatory and non-inflammatory diseases. Clinical Biochemistry. 2016;49(7):534-7.
- 163. Lunemann S, Malone DF, Hengst J, Port K, Grabowski J, Deterding K, et al. Compromised function of natural killer cells in acute and chronic viral hepatitis. The Journal of infectious diseases. 2014;209(9):1362-73.
- 164. Al-Jaderi Z, Maghazachi AA. Effects of Vitamin D3, Calcipotriol and FTY720 on the Expression of Surface Molecules and Cytolytic Activities of Human Natural Killer Cells and Dendritic Cells. Toxins. 2013;5(11).
- 165. Moudi B, Heidari Z, Mahmoudzadeh-Sagheb H. CCR5, MCP-1 and VDR Gene Polymorphisms Are Associated with the Susceptibility to HBV Infection. Indian journal of clinical biochemistry: IJCB. 2019;34(4):407-17.
- 166. Lin TM, Chen CJ, Wu MM, Yang CS, Chen JS, Lin CC, et al. Hepatitis B virus markers in Chinese twins. Anticancer research. 1989;9(3):737-41.
- 167. Welsh J. Vitamin D and breast cancer: Past and present. J Steroid Biochem Mol Biol. 2018;177:15-20.
- 168. Landel V, Millet P, Baranger K, Loriod B, Féron F. Vitamin D interacts with Esr1 and Igf1 to regulate molecular pathways relevant to Alzheimer's disease. Molecular Neurodegeneration. 2016;11(1):22.
- 169. Kamatani Y, Wattanapokayakit S, Ochi H, Kawaguchi T, Takahashi A, Hosono N, et al. A genome-wide association study identifies variants in the HLA-DP locus associated with chronic hepatitis B in Asians. Nature genetics. 2009;41(5):591-5.
- 170. Xu Y, He B, Pan Y, Deng Q, Sun H, Li R, et al. Systematic review and meta-analysis on vitamin D receptor polymorphisms and cancer risk. Tumor Biology. 2014;35(5):4153-69.
- 171. Moudi B, Heidari Z, Mahmoudzadeh-Sagheb H. CCR5, MCP-1 and VDR Gene Polymorphisms Are Associated with the Susceptibility to HBV Infection. Indian Journal of Clinical Biochemistry. 2019;34(4):407-17.
- 172. Peng Q, Yang S, Lao X, Li R, Chen Z, Wang J, et al. Association of Single Nucleotide Polymorphisms in VDR and DBP Genes with HBV-Related Hepatocellular Carcinoma Risk in a Chinese Population. PLOS ONE. 2014;9(12):e116026.

- 173. He Q, Huang Y, Zhang L, Yan Y, Liu J, Song X, et al. Association between vitamin D receptor polymorphisms and hepatitis B virus infection susceptibility: a meta-analysis study. 2018;645:105-12.
- 174. Hoan NX, Khuyen N, Giang DP, Binh MT, Toan NL, Anh DT, et al. Vitamin D receptor Apal polymorphism associated with progression of liver disease in Vietnamese patients chronically infected with hepatitis B virus. BMC medical genetics. 2019;20(1):201.
- 175. Shan B, Wang JY, Wang X, Fu JJ, Li L, Pan XC, et al. VDR rs7975232/Apal genetic variation predicts sustained HBsAg loss in HBeAg-positive chronic hepatitis B patients treated with pegylated interferon. Journal of medical virology. 2019;91(5):765-74.
- 176. Cusato J, Boglione L, De Nicolò A, Imbornone R, Cardellino CS, Ghisetti V, et al. Association of vitamin D pathway SNPs and clinical response to interferon in a cohort of HBeAg-negative patients. Pharmacogenomics. 2017;18(7):651-61.
- 177. Thanapirom K, Suksawatamnuay S, Sukeepaisarnjaroen W, Treeprasertsuk S, Tanwandee T, Charatcharoenwitthaya P, et al. Vitamin D-Binding protein Gene Polymorphism Predicts Pegylated Interferon-Related HBsAg Seroclearance in HBeAg-Negative Thai Chronic Hepatitis B Patients: A Multicentre Study %J Asian Pacific Journal of Cancer Prevention. 2019;20(4):1257-64.
- 178. Gao W, Wang R, Wang X, Wu H, Wang Y, Lu X, et al. Vitamin D serum levels and receptor genetic polymorphisms are associated with hepatitis B virus and HIV infections and IFN- λ levels. Biomarkers in Medicine. 2017;11(9):733-40.
- 179. Huang Y, Liao Y, Chen W, Chen C, Hu J, Liu C, et al. Vitamin D receptor gene polymorphisms and distinct clinical phenotypes of hepatitis B carriers in Taiwan. Genes & Immunity. 2010;11(1):87-93.
- 180. Thanapirom K, Suksawatamnuay S, Sukeepaisarnjareon W, Tanwandee T, Charatcharoenwitthaya P, Thongsawat S, et al. Genetic variation in the vitamin D pathway CYP2R1 gene predicts sustained HBeAg seroconversion in chronic hepatitis B patients treated with pegylated interferon: A multicenter study. PLOS ONE. 2017;12(3):e0173263.
- 181. Xie Q, Hu X, Zhang Y, Jiang X, Li X, Li J. Decreasing hepatitis B viral load is associated with a risk of significant liver fibrosis in hepatitis B e antigen positive chronic hepatitis B. Journal of medical virology. 2014;86(11):1828-37.
- 182. Allain J-P. Global epidemiology of occult HBV infection. Annals of Blood. 2017;2(5).
- 183. Falleti E, Bitetto D, Fabris C, Cussigh A, Fontanini E, Fornasiere E, et al. Vitamin D receptor gene polymorphisms and hepatocellular carcinoma in alcoholic cirrhosis. World journal of gastroenterology. 2010;16(24):3016-24.
- 184. Yang YM, Seki E. TNFα in liver fibrosis. Current pathobiology reports. 2015;3(4):253-61.
- 185. Rex J, Lutz A, Faletti LE, Albrecht U, Thomas M, Bode JG, et al. IL-1β and TNFα Differentially Influence NF-κB Activity and FasL-Induced Apoptosis in Primary Murine Hepatocytes During LPS-Induced Inflammation. Frontiers in Physiology. 2019;10(117).
- 186. Lin J, Wu JF, Zhang Q, Zhang HW, Cao GW. Virus-related liver cirrhosis: molecular basis and therapeutic options. World journal of gastroenterology. 2014;20(21):6457-69.
- 187. Araujo OC, Barros JJ, do Ó KM, Nabuco LC, Luz CA, Perez RM, et al. Genetic variability of hepatitis B and C viruses in Brazilian patients with and without hepatocellular carcinoma. Journal of medical virology. 2014;86(2):217-23.
- 188. Asthana M, Sahu SK, Kumar A, Mohanty S, Chakrabarti S, Das P, et al. Role of Interleukin 28B Polymorphisms in Response to Interferon Based Therapy for Hepatitis C Virus Clearance. Current drug metabolism. 2018;19(3):215-23.
- 189. Thanapirom K, Suksawatamnuay S, Sukeepaisarnjaroen W, Tangkijvanich P, Thaimai P, Wasitthankasem R, et al. Genetic associations of vitamin D receptor polymorphisms with advanced liver fibrosis and response to pegylated interferon-based therapy in chronic hepatitis C. PeerJ. 2019;7:e7666.

Journal Pre-problem

Table 1. Chronic phase of hepatitis B disease categories								
Category	HBV DNA	HBe Ag	Anti- HBe	ALT				
Immune-tolerant	High	Positive	Negative	Normal/low	Replication of HBV			
Immune clearance	High (Decrease /Low)	Positive	Negative	high	Liver disease/inflammation			
Inactive HBs antigen carrier	Low/Undetectable	Negative	Positive	Normal	Remission of liver disease			
Reactivation phase	High	Negative	Positive	Elevated				

Table2. Clinical trial studies of vitamin D and hepatitis B infection						
Study Title	Interventions	Locations	Detailed Description	Status	Current Primary Outcome	
The Relationship Between Vitamin D and Hepatitis B Virus Replication	Dietary Supplement: Vitamin D	Taipei Tzu Chi Hospital, Buddhist Tzu Chi Medical Foundation New Taipei city, Taiwan	-Randomized case-control trial -Total of 149 HBV carriers with inadequate vitamin D (< 30 ng/mL) -Randomly divided to two groups: one group receiving vitamin D supplement (1600 IU/day) for 2 months and another group as controls	Completed	- Dynamic Change of HBV DNA - Change of Serum qHBsAg (IU/mL) [Time Frame: baseline, after 2-month vitamin D supplement] The serum qHBsAg levels were measured before and after 2- month vitamin D supplement	
Oral Vitamin D Treatment for the Prevention of HCC	Drug: Vitamin D3	Randomized	-Potential participants will be identified from the follow-up cohort of chronic hepatitis B in the third -The participant instructed to begin taking 2 tablets per day (800 IU total) of vitamin D3 besides their regular anti-virus treatment -The researcher will investigate general treatment benefits and the potential to reduce the development of Hepatocellular carcinoma (HCC), also known as liver cancer	Not yet recruiting	-Change in serum levels of 25-hydroxy vitamin D [Time Frame: at baseline, and at 6 and 12 months] -Change in serum levels of 25-hydroxyvitamin D at 6 months and 12 months compared to baseline	
The Beneficial Effect of Vitamin D Supplement to Peginterferon Alpha 2a or to Telbivudine Monotherapy in Patients With Chronic HBV Infection	Drug: Peginterferon + Vitamin D Drug: Peginterferon Drug: Sebivo Drug: entecavir+ vitamin D	Ziv medical center liver unit Safed, Israel, Israel Liver clinic Safed, Israel	Not Provided	Unknown	-Treatment efficacy [Time Frame: 120 weeks] - Primary end point will be sustained viral response which was defined as clearance of HBeAg from serum and HBV -DNA less than 10,000 copies/mL (2000 IU/mL) at 6 months after treatment -HBsAg titre during treatment and at 6 months follow up will be measured -Histologic response [Time Frame: 120 WEEKS]	
To Study the Effect of Adding on PEG- INF Therapy for Patients Diagnosed With Chronic Hepatitis B	Drug: PEG- IFN & Nucleos(t)tide analogues Drug: Nucleos(t)tide analogues	King Abdulaziz Medical City Jeddah, Saudi Arabia King Abdulaziz Hospital Jeddah, Saudi Arabia King Abdulaziz Gedah, Saudi Arabia King Abdulaziz Medical City Riyadh, Saudi Arabia	-IFN-α with its dual immunomodulatory and antiviral effects was the first drug for Chronic HBV treatment followed by introduction of nucleos(t)ide analogues(NA) - Directly inhibit HBV polymerase and provide an effective on treatment maintained viral suppression - PEG-IFN allows a convenient once a week dosing interval and of equal or superior treatment efficacy than conventional (IFN) Due to its predominant immunomodulatory effect PEG-IFN offers the advantage of higher	Unknown	The loss of HbsAg between groups (NA) group and NA +Peg_INF group assessed by HbsAg test [Time Frame: 48 weeks] The loss of HbsAg between groups (NA) group and NA +Peg_INF group assessed by HbsAg test	

-			
		sustained off treatment response	onse
		rate compared to NA thus al	llowing
		a finite duration of treatmen	t. The
		NA act by directly inhibiting	g HBV
		polymerase resulting in effe	ctive
		on treatment maintained vira	al
		suppression (HBV DNA PC	CR
		<200 for last 3-6 month)	

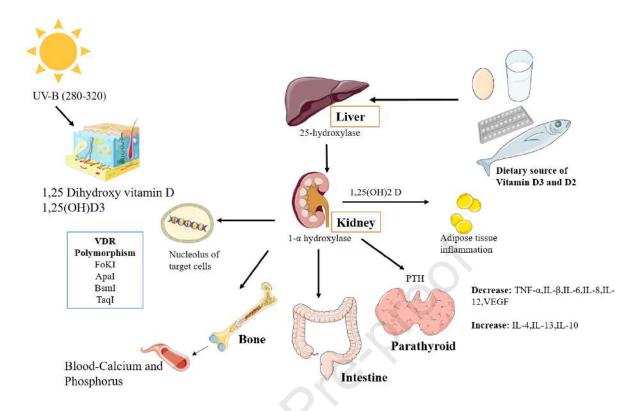


Figure 1. Vitamin D activation and metabolism

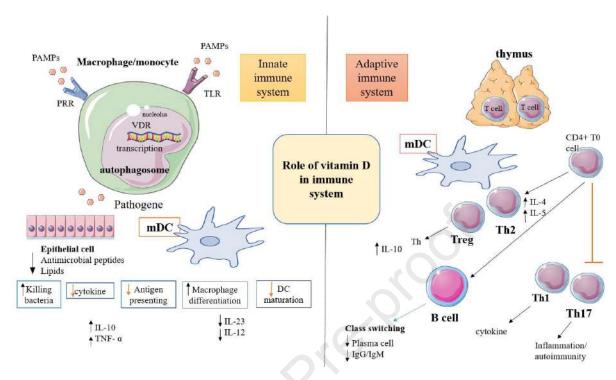


Figure 2. The Role of Vitamin D in the Immune System