RESEARCH ARTICLE

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Modulating effect of vitamin D status on serum anti-adenovirus 36 antibody amount in children with obesity: National Food and Nutrition Surveillance



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Abstract

Background: The association of ADV-36 infection and obesity has been reported in children. The objective of this study was to examine the hypothesis that the association between ADV-36 infection and adiposity may be mediated by sub-optimal vitamin D status of the host.

Methods: Ninety one apparently healthy children in different weight categories (normal weight: 33, overweight: 33, obesity: 25) aged 5–18 years were randomly selected from the registered population at National Food and Nutrition Surveillance Program (NFNS). The groups were matched based on age and sex. Anthropometric, biochemical and serological assessments were performed.

Results: The amount of anti-ADV36-Ab increased whereas circulating concentrations of 25(OH) D decreased across BMI categories with higher amounts in children with normal weight than in children with overweight and obesity $(31.0 \pm 16.4, 22.5 \pm 10.5 \text{ and } 21.9 \pm 9.8 \text{ nmol/L}$, respectively, p = 0.004). Logistic regression analysis revealed that for each unit increment of anti-ADV36-Ab, the chance of increase in weight was 8.5 times (OR: 8.5, p = 0.029). Interestingly, when 25(OH) D was introduced into the model, anti-ADV36-Ab was no longer the predictor of weight increment and the chance of increase in weight reduced 5% for each unit increase in 25(OH) D concentration (OR: 0.95, p = 0.012).

Conclusion: It is suggested that ADV36-induced lipogenesis may be mediated by vitamin D deficiency in children with obesity.

Keywords: Vitamin D, Adenovirus 36, Obesity, Children

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Background

Contributors to childhood obesity are in the "obesogenic environment" of the children including all those aspects in the child's environment that encourages him or her to eat more (usually unhealthy) foods and to have less physical activity [1]. One of the proposed potential contributors to obesity is suboptimal vitamin D status. Detection of vitamin D receptor (VDR) and its signaling pathways in adipose tissue indicated that vitamin D could potentially affect development, metabolism and functions of body fat mass [2]. Laboratory in vitro studies demonstrated the effect of calcitriol (1,25(OH)₂D₃), the active form of the vitamin, and its inactive metabolites on adipogenesis in murine 3 T3-L1 cell line. While VDR knock out (VDR-/-) and 1- α -hydroxylase knock out (CYP27B1 -/-) mice were highly resistant to weight gain due to dietary intake, the over-expression of human VDR in adipose tissue resulted in increased adipogenesis [3]. Interestingly, a cohort study revealed that circulating concentrations of 25(OH) D, the main biomarker of vitamin D status, below 50 nmol/L was related to new onset obesity in adults [4]. Nevertheless, despite some clinical trials showing the fat mass-decreasing and antiinflammatory effects of supplementary vitamin D in adults [5–7], actually the relationship between vitamin D deficiency and obesity is still like a "chicken and egg story" [8].

Among the causative factors of obesity, a rather newly proposed theory is viral infections [9]. Human adenovirus-36 (ADV-36) through the effect of viral E4orf1 gene on lipogenic enzymes may induce host adipogenesis [10, 11]. The association of ADV-36 infection and obesity has been reported by some research groups in children [12–14] and in adults, as well [15–18]. However, some other studies did not confirm this finding [19, 20] including a study conducted in Iran [21]. One meta-analytical study documented the association between ADV-36 infection and obesity only in adults but not in children [22].

Though the in vitro and in vivo animal studies have reported the association between ADV-36 infection and adipogenesis [23], this association is controversial in humans. In human studies despite statistically significant difference between proportion of seropositive subjects with obesity and seronegative subjects with normal weight, there are usually considerable number of seronegative subjects with obesity and seropositive subjects who are normal weight [24, 25]. Furthermore, the prevalence of overweight/obesity between 1980 and 2013 increased 27.5% for adults and 47.1% for children, with a more remarkable increase in developed countries [26]. This rise can hardly be explained just by ADV-36 infection, which is acquired through intranasal route and respiratory tract [27] and it is expected to be more

prevalent in economically poor countries, wherein hunger and underweight are, and probably continue to be, a problem [28].

The other important very noticeable issue is obesity comorbidities. Dysglycemia and diabetes is among the most common obesity complications [29]. It has been estimated that mortality rates due to cardiovascular problems and diabetes would increase by 41 and 21%, respectively, for every 5 unit increase in body mass index above 25 kg/m² [30]. Controversially, ADV-36 infection has been associated with lower dyslipidemia risk [18], increased insulin sensitivity [31] and lower occurrence of diabetes [31]. Even ADV-36 and certain gut microbes have been examined for their ability to ameliorate animal and human blood lipids and glucose [32]. The main question could be "what has protected ADV-36 seropositive normal weight children from adiposity?" One answer could be that seropositve children may be more prone to weight gain in future. At least one prospective study did not confirm this notion [33]. The other answer may be other contributing factors interacting with this association.

We hypothesized that the association between ADV-36 infection and adiposity may be mediated by sub-optimal vitamin D status of the host. To examine this hypothesis, we conducted a case control study on children and adolescents with normal weight, overweight and obesity.

Methods

Using G*Power 3.1 software (Universität Düsseldorf, Düsseldorf, Germany), the needed total sample size to assure statistical power of 0.8 and effect size of 0.35 was calculated 84 participants. In total, 91apparently healthy children in different weight categories (normal weight: 33, overweight: 33, obesity: 25) aged 5-18 years were randomly selected from the registered population at National Food and Nutrition Surveillance Program (NFNS), a population-based survey conducted periodically in Iran by the National Nutrition and Food Technology Research Institute (NNFTRI) in collaboration with the Deputy of Health of Iran Ministry of Health and Medical Education and United Nations Children's Fund (UNICEF) as described elsewhere [34]. The groups were matched based on age and sex. The inclusion criteria were having no history of hepatic or renal disease and taking no vitamin D supplements or medications affecting vitamin D metabolism such as anticonvulsant or corticosteroids at least 2 months prior to the study. The study protocol and objectives were clarified for all participants and their parents or their legal guardians before they signed a written informed consent. The study protocol and procedures were approved by Ethics Committee of NNFTRI.

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Anthropometric measurements

Height was measured to the nearest of 0.1 cm using a wall-mounted stadiometer (Seca 206, Seca Company, Hamburg, Germany). Weight was measured while participants wearing light clothing and no shoes with a digital scale accurate to the nearest of 0.1 kg (Seca 840, Seca Company, Hamburg, Germany). Both instruments were calibrated daily. Body mass index (BMI), which has a strong correlation with adiposity in children [35], was defined as weight (kg)/height (m)². Overweight and obesity were categorized using the BMI for age z-score (1–2 and > 2, respectively).

Laboratory investigations

Blood samples drawn in the morning (08:00-10:00 h) after an overnight fast were centrifuged at room temperature for 10 min at 800 g to separate sera, which were then aliquoted and stored at $-80 \,^{\circ}\text{C}$ immediately until the day of analysis.

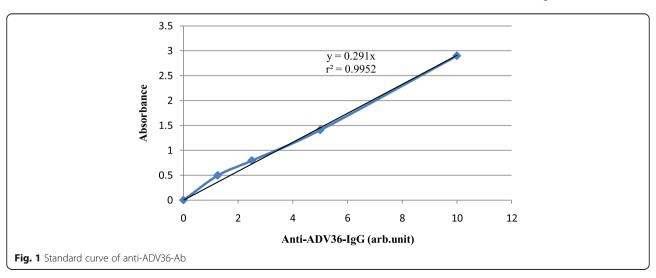
Total cholesterol, triglycerides, low-density lipoprotein-cholesterol (LDL-C) and high-density lipoprotein-cholesterol (HDL-C) concentrations were measured by using enzymatic methods (Pars-Azmoon, Tehran, Iran) and an auto-analyzer (Selecta E; Vitalab, Holliston, the Netherlands).

To determine serum concentrations of 25(OH) D, a commercial kit of direct enzyme immune-assay (EIA) was used (DIAsource, Louvain-la-Neuve, Belgium). The intra- and inter-assay variations were 2.5–7.8% (for values of 13.7–203 nmol/L) and 4.3–9.2% (for values 44.25–213.7 nmol/L), respectively, according to the manufacturer's manual. The accuracy of measurements of 25(OH) D concentrations were ensured using high-performance liquid chromatography [36] in the Laboratory of Nutrition Research, NNFTRI, that has been participating in the Vitamin D External Quality Assessment Scheme (DEQAS) since 2012.

Anti-adenovirus 36 antibody (anti-ADV-36-Ab) was assayed using EIA kit (Zellbio, Veltlinerweg, Ulm, Germany). Briefly, 50 µL pre-diluted serum samples (1: 5), control sera (ready to use negative and positive, both included in the kit) followed by enzyme conjugate were transferred to the antibody coated microwells of a plate. After incubation and washing, a color reaction was developed following adding a chromogen to the wells. The reaction was halted using a stop solution. In this assay, the amount of absorbance at 450 nm is proportional to the absolute amount of specific antibody. We generated a standard curve using serially diluted positive control serum. The concentration of anti-ADV-36-Ab was expressed using arbitrary (arb) unit (Fig. 1). The intraand inter-assay variations were < 10 and < 12%, according to the manufacturer.

Statistical analyses

Continuous characteristics were expressed as mean ± standard deviation (SD); categorical variables were displayed as frequencies. Normality of distribution was checked for all variables using Shapirio-Wilk test. Tests for differences of continuous variables among three BMI categories were performed using analysis of variance (ANOVA) or Kruskal-Wallis. Levene's test was used to evaluate homogeneity of between-group variances. For those variables with statistically unequal variances, Dunnett's T3 was applied for pair wise multiple comparisons. The analysis of covariate test was used to compare anti-ADV-36-Ab among BMI categories by adjusting serum 25(OH) D concentrations. Pearson's correlation coefficient and multiple linear and logistic regression analysis were used to assess relationships between variables. A 2-tailed p <was considered significant. Analyses were performed using Statistical Package for Social Sciences 21.0 for Windows (SPSS Inc., Chicago, IL, USA).



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Table 1 Distribution of vitamin D status in the studied children according to IOM and Endocrine Society criteria

	Deficiency, n (%)	Insufficiency, n (%)	Sufficiency, n (%)
Institute of Medicine	52 (57.1)	32 (35.2)	7 (7.7)
Endocrine Society Practice Guideline	84 (92.3)	7 (7.7)	0 (0)

Vitamin D status based on circulating 25(OH) D concentrations according to:

a. Institute of Medicine [37]: deficiency < 25 nmol/L; insufficiency: 25-50 nmol/L; sufficiency: > 50 nmol/L

b. Endocrine Society Practice Guideline [38]: deficiency < 50 nmol/L; insufficiency: 50-75 nmol/L; sufficiency: > 75 nmol/L

Results

Over 92 and 100% of the participants had undesirable vitamin D status according to Institute of Medicine (IOM) [37] and Endocrine Society Guideline [38], respectively (Table 1). There was no significant difference in anthropometric, biochemical and serum antiviral antibody measures between boys and girls (Table 2). Though circulating 25(OH) D concentrations were higher in boys than in girls, the difference was not statistically significant (27.8 \pm 13.0 vs. 22.6 \pm 13.3 nmol/L, p = 0.068).

Table 3 shows the measures of the studied variables in three BMI categories. The concentrations of anti-ADV36-Ab increased across BMI categories. However, only the difference between categories of normal weight and obesity was statistically significant (2.0 ± 1.6 vs. 6.3 ± 6.8 arb.unit, p = 0.013). This difference disappeared in analysis of covariate after adjustment for 25(OH) D concentrations. Circulating concentrations of 25(OH) D decreased across BMI categories with higher amounts in normal weight than in children with overweight and obesity (31.0 \pm 16.4, 22.5 ± 10.5 and 21.9 ± 9.8 nmol/L, respectively, p =0.004). Anti-ADV36-Abs showed a direct correlation with BMI z-score (r = 0.294, p = 0.005) but a negative correlation with circulating 25(OH) D concentrations (r = -0.236, p = 0.024) (Fig. 2). Serum 25(OH) D concentrations also inversely correlated with BMI z-score (r = -0.287, p = 0.006). Figure 3 shows the trends of serum 25(OH) D and anti-ADV36-Ab levels across BMI quartiles of the studied children.

Logistic regression analysis revealed that for each unit increment of anti-ADV36-Ab, the chance of increase in weight was 8.5 times (OR: 8.5, p = 0.029). Interestingly, when 25(OH) D was introduced into the model, anti-ADV36-Ab was no longer the predictor of weight increment and the chance of increase in weight reduced 5% for each unit increase in 25(OH) D concentration (OR: 0.95, p = 0.012).

Ordinal regression models were applied to estimate the odds ratios (ORs) and 95% confidence intervals (CIs) of obesity by serum 25(OH) D and anti-ADV36-Ab, adjusted for potential confounders. These models were examined using a full likelihood ratio test comparing the fitted location model to a model with varying location parameters (Table 4). The analysis revealed that with each unit increment of anti-ADV36-Ab, the chance of increase in weight status was 2.86 times (95% CI: 1.25 to 6.52, p = 0.012). Interestingly, when 25(OH) D was introduced into the model, anti-ADV36-Ab was no longer the predictor of weight increment and the chance of increase in weight reduced 5% for each unit increase in calcidiol concentration (OR: 0.96, p = 0.017).

Table 2 Comparison of anthropometric measures, lipid profile components, serum 25(OH) D and anti-ADV36-Ab between boys and girls

Variable	Boys (n ₁ = 53)	Girls (n ₂ = 38)	p value	Total (n = 91)
Age (yrs)	9.16 ± 2.7	10.3 ± 3.5	0.078	9.6 ± 3.1
Weight (kg)	42.5 ± 16.9	46.8 ± 22.8	0.308	44.3 ± 19.6
Height (cm)	138.4 ± 18.4	138.2 ± 22.5	0.969	138.3 ± 20.1
BMI (kg/m ²)	21.6 ± 6.1	23.3 ± 6.8	0.215	22.3 ± 6.4
z-score for BMI for age	1.33 ± 0.87	1.30 ± 0.94	0.854	1.3 ± 0.9
Triglyceride (mg/dL)	87.5 ± 42.4	86.9 ± 47.0	0.954	87.3 ± 44.1
Total Cholesterol (mg/dL)	144.3 ± 27.1	148.1 ± 22.7	0.477	145.9 ± 25.3
HDL-C (mg/dL)	53.3 ± 13.4	52.0 ± 11.3	0.627	52.7 ± 12.5
LDL-C (mg/dL)	73.5 ± 20.8	78.8 ± 18.9	0.220	75.7 ± 21.0
25(OH) D (nmol/L)	27.8 ± 13.0	22.6 ± 13.3	0.068	25.7 ± 13.3
Anti-ADV36-Ab (arb.unit)	4.05 ± 5.5	3.80 ± 5.5	0.831	3.94 ± 5.5

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Table 3 Comparison of anthropometric measures, lipid profile components, serum 25(OH) D and anti-ADV36-Ab among children in three weight categories

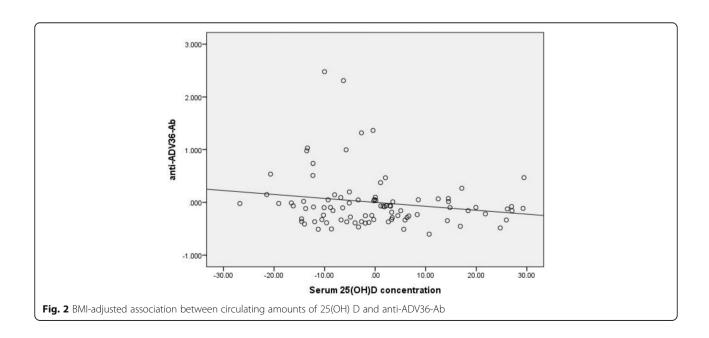
Variable	Normal Weight (n ₁ = 33)	Overweight (n ₂ = 33)	Obesity (n ₃ = 25)	p value
Age (yrs)	9.8 ± 3.1	9.7 ± 3.0	9.3 ± 3.3	0.820
Weight (kg)	34.8 ± 13.1^{a}	46.0 ± 16.2^{b}	53.3 ± 25.3^{b}	0.001
Height (cm)	138.3 ± 19.7	142.0 ± 15.1	133.5 ± 25.7	0.289
BMI (kg/m ²)	17.4 ± 1.7 ^a	22.5 ± 4.1^{b}	$28.7 \pm 7.5^{\circ}$	< 0.001
Triglyceride (mg/dL)	74.9 ± 36.4	90.8 ± 43.0	98.8 ± 52.1	0.104
Total Cholesterol (mg/dL)	144.1 ± 24.0	143.3 ± 26.5	151.8 ± 25.5	0.401
HDL-C (mg/dL)	55.0 ± 10.2	51.8 ± 13.5	51.0 ± 14.0	0.428
LDL-C (mg/dL)	74.1 ± 20.2	73.3 ± 19.3	80.9 ± 20.8	0.308
25(OH) D (nmol/L)	31.0 ± 16.4^{a}	22.5 ± 10.5 ^b	21.9 ± 9.8^{b}	0.004
Anti-ADV36-Ab (arb.unit)	2.0 ± 1.6^{a}	4.1 ± 6.4^{b}	6.3 ± 6.8^{b}	0.013

Numbers in a row not sharing a common superscript are significantly different

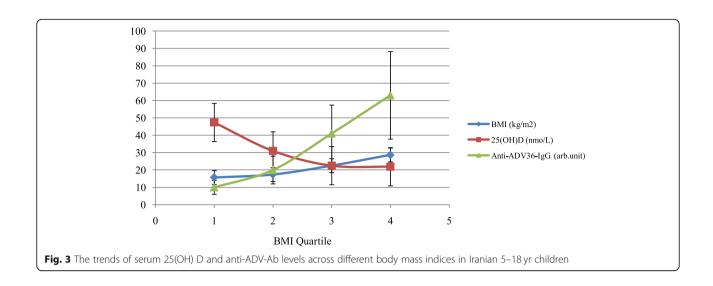
Discussion

High occurrence of undesirable vitamin D status in our subjects is in accord with our previous reports [39, 40]. We found an increase in the amount of anti-ADV36-Ab across BMI categories in the studied children indicating a possible effect of subclinical infection in adipogenesis. Several factors have been introduced as the contributors in childhood obesity mostly in the first 1000 days of life and then in the obesogenic environment [41]. Meanwhile, some reports have shown an association between obesity and ADV36 infection [12, 13, 22]. In contrast, some studies failed to show the lipogenic effect of ADV-36 infection [19].

Subclinical viral infections may contribute in metabolic derangements commonly seen in obesity and metabolic syndrome [42]. In a study on Hispanic males and females, the associations among ADV36 seropositivity, adiposity and indicators of glycemic status were examined initially and after almost 10 years. Seropositive subjects showed greater adiposity both at the baseline and after 10 years but they had lower concentrations of fasting serum insulin compared with seronegative subjects. The researchers concluded that ADV36 infection may exert its adipogenic effect even long after the initial infection but may also have a modulatory effect on glycemic control [15]. Improvement of glycemic status in



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ADV-36 seropositive subjects has been ascribed to the induction of hepatic mitochondrial function [43].

Some meta-analytical studies confirmed the higher risk of adiposity and weight gain in adults due to ADV36 infection [17, 22]. Notwithstanding, we found this association may be modulated by vitamin D status. The inverse relationship of adiposity and circulating 25(OH) D that was observed in this study and other studies [44] may be the key and already missing link between ADV36 infection and obesity. Vitamin D has anti-inflammatory [6], antioxidant [45] and antiviral properties [46]. The exact mechanism of vitamin D antiviral function remains to be clarified but it may pertain to up-regulation of antimicrobial peptides, including cathelcidins (LL-37) and β -defensin 2 [47, 48].

Viruses causing chronic infection with low-grade inflammation including ADV36 may comprise a small part of the host's metagenome or virome. The final upshot of the relation between the virome and the host could be damaging, harmless or even symbiotic [49] depending on the host's immunocompetence status. Vitamin D functions as an immunomodulator in several ways by affecting both innate and adaptive immunity [50, 51].

Table 4 Odds Ratio (95% CI) from ordinal regression models for association of weight status with anti-ADV36-Ab

	Variables	OR	95% CI	Wald χ ²	p value
Model 1	anti-ADV36-Ab	2.86	1.25 to 6.52	6.24	0.012
Model 2	anti-ADV36-Ab	2.27	0.99 to 5.2	3.79	0.052
	25(OH)D	0.961	0.93 to 0.99	5.72	0.017
Model 3 ^a	anti-ADV36-Ab	2.17	0.94 to 5.0	3.35	0.067
	25(OH)D	0.95	0.92 to 0.98	6.64	0.010

^a Model 3. Adjusted for sex and age

Vitamin D deficiency may, therefore, turn the relation between ADV36 virome and the affected child to damaging by rendering ADV36-induced adipogenesis.

The escalating trends of BMI in children have reached a steady high state in many developed countries but have hastened in certain parts of Asia [52]. These data together with emerging evidence for the relationship between ADV36 infection and obesity have been provocative enough that even vaccination against adenoviral infection has been proposed to induce herd immunity against obesity and its related metabolic derangements [53]. Our findings, however, strengthen the possibility that improvement of vitamin D status of children through supplementation and food fortification might work as well as, and even better than, a true vaccine. Some recent studies support this notion [54–56].

Some limitations in this study are acknowledged. The specificity of EIA method to detect neutralizing anti-ADV-Abs has been questioned [57]. The results of this study need to be confirmed by further studies in other populations with a larger sample size and a more specific method.

Conclusion

To the best of our knowledge, this the first report of the relation of vitamin D status with anti-ADV36-Ab in children with obesity. We propose that in the future human studies on the association of ADV-36 infection and adiposity, vitamin D status of the subjects must also be taken into consideration. This observational study sets the stage for conducting a randomized controlled trial that would clearly define this proposed relationship.

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Abbreviations

ADV-36: Adenovirus-36; ANOVA: Analysis of variance; Anti-ADV-36-Ab: Anti-adenovirus 36 antibody; Arb.unit: Arbitrary unit; BMI: Body mass index; CI: Confidence interval; EIA: Enzyme immunoassay; HDL-C: High-density lipoprotein-cholesterol; IOM: Institute of Medicine; LDL-C: Low-density lipoprotein-cholesterol; NFNS: National Food and Nutrition Surveillance Program; NNFTRI: National Nutrition and Food Technology Research Institute; 25(OH)D: 25-hydroxycalciferol; OR: Odds ratio; SD: Standard deviation; UNICEF: United Nations Children's Fund; VDR: Vitamin D receptor

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Authors' contributions

TN and BN designed this part of NFNS and performed all laboratory works. Statistical analyses were done by BN who also prepared the preliminary manuscript. TN finalized the manuscript with the intellectual aid of BH. All authors have read and approved the manuscript.

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Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Ethics approval and consent to participate

All parents or legal guardians of the participants signed a written informed consent. The ethical issues of this study were approved by the Ethical Committee of National Nutrition and Food Technology Research Institute, Shahid Beheshti University of Medical Sciences (code: ir.sbmu.nnftri.rec.1396.170).

Consent for publication

Not applicable

Competing interests

The authors declare that they have no competing interests.

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