Review Article Vitamin D and pulmonary fibrosis: a review of molecular mechanisms

Dandan Ma¹, Lipan Peng²

¹Department of Critical-Care Medicine, Affiliated Jining First People's Hospital, Shandong, China; ²Department of Gastrointestinal Surgery, Shandong Provincial Hospital Affiliated to Shandong University, Jinan 250000, Shandong, China

Received May 31, 2018; Accepted February 18, 2019; Epub September 1, 2019; Published September 15, 2019

Abstract: Pulmonary fibrosis is a serious interstitial disease characterized by initial diffuse alveolar inflammation, fibroblast proliferation, ECM accumulation, and the destruction of normal pulmonary tissues, whose etiology remains unknown and therapeutic options remain limited. The prevalence of Vitamin D deficiency is increasing and has been linked to pulmonary fibrosis. In recent years, many studies focused on the mechanistic pathway of Vitamin D in the prevention of fibrosis. This review highlights the current evidence on the molecular mechanisms of Vitamin D in pulmonary fibrosis. We want to provide new clues to the clinical management of pulmonary fibrosis.

Keywords: Vitamin D, vitamin D receptor, VDR, pulmonary fibrosis

Introduction

Pulmonary fibrosis describes a progressive and irreversible lung disease, with a mean life expectancy of only 3-5 years [1-5], posing a big threat to public health. Common symptoms caused by pulmonary fibrosis such as fatigue, cough, and dyspnea have a major impact on the quality of life (QOL) of patients [6]. Its etiology remains elusive and therapeutic options remain limited. Until now, there is no pharmacological therapy but lung transplantation to change the natural course of the disease.

Pulmonary fibrosis is a wound healing response caused by lung injury and infection. However, chronic exposure to the injury factor such as allergens, toxic chemicals, and radiation leads to dysregulated wound healing response, overlapping inflammation and subsequent pulmonary fibrosis [2]. Finally the scar tissues take the place of normal lung architecture [7]. To date, the potential molecular mechanism of pulmonary fibrosis is still unknown. But it is closely related to the regulation of collagensecreting myofibroblasts proliferation, activation, and differentiation, and inflammation.

Vitamin D has long been regarded as a key player in calcium homeostasis, bone health, electrolyte and blood pressure regulation and immune response [8, 9]. It is provided by the kidney and parathyroid gland endocrine system. To achieve full biologic activity, Vitamin D must be metabolized to the hormonal form 1,25-dihydroxy Vitamin D by the activating hydroxylase the Vitamin D 25-hydroxylase, and 1-αhydroxylase [10]. As is known to all, Vitamin D is distributed not only to the liver but also to all tissues in the human body. For the moment, many of these tissues are now found to contain many hydroxylases that alters Vitamin D into 1.25-dihydroxy Vitamin D, thus the autocrine production of 1,25-dihydroxy Vitamin D in those tissues occurs [11-14]. The biologically active metabolite of 1,25-dihydroxy Vitamin D is thought to exert its principal actions by binding to the Vitamin D receptor (VDR, a ligand-dependent transcription factor) [15].

However, vitamin D is not just a vitamin; the pleiotropic roles of Vitamin D have been highlighted in various diseases. Vitamin D is related to cell proliferation, differentiation, apoptosis, intercellular adhesion, oxidative stress, matrix homeostasis, and regulation of inflammatory

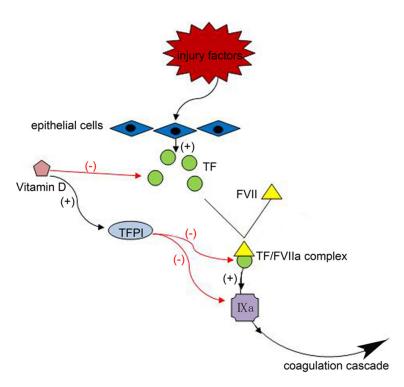


Figure 1. When epithelial cells get stimulated by injury factors, they will release TF, and then together with activated factor VIIa (FVIIa) formed TF/FVIIa complex, which activate factor IX, this is the primary initiator of coagulation cascade. Vitamin D can exert anticoagulant effects in two ways: (1) Vitamin D suppresses TF expression, (2) Vitamin D increases the levels of TFPI.

response [16-18]. The association between serum Vitamin D and pulmonary fibrosis has become a hot topic in recent years. Vitamin D affects the progress of pulmonary fibrosisa variety of ways. This review gives a more detailed and integrated elaboration on the mechanistic pathway of Vitamin D in prevention of pulmonary fibrosis.

The distinct stages of pulmonary fibrosis and vitamin D

Pulmonary fibrosis is an abnormal wound healing response caused by lung injury and infection, which has four distinct stages: a clotting/coagulation phase, an inflammatory phase, a fibroblast migration/proliferation/activation phase, and a tissue remodeling and resolution phase [7]; pulmonary fibrosis usually occurs if any stage in the tissue repair process is dysregulated.

A clotting/coagulation phase

When epithelial cells are stimulated by injury factors, they release inflammatory mediators

and then activate an antifibrinolytic coagulation cascade. Unfortunately, the excessive activation of the coagulation cascade has been seen throughout the process of pulmonary fibrosis [19, 20]. Recent studies implicated that the coagulation factors are predominantly mediated by protease-activated receptors (PARs), and PARs play a significant role in the pathogenesis of lung fibrosis [19, 21-23]. This family comprises four members (PAR1, PAR2, PAR3, and PAR4) but current evidence suggests PAR1 play a major role in the context of lung injury. PARs mediates tissue factor (TF) [24, 25], then TF together with activated factor VIIa (FVIIa) forms TF/FVIIa complex. The TF/FVIIa is the primary initiator of the coagulation cascade, which activates factor IX, but is blocked by the TF pathway inhibitor (TFPI), a protease inhibitor.

Vitamin D can exert anticoagulant effects by two predominant ways (**Figure 1**): 1) Some experimental work has shown that the Vitamin D had a potent capacity to suppress TF expression, by tumor necrosis factor- α (TNF α , a key activator of TF) [26-29]. Other studies demonstrated a significant positive correlation between Vitamin D and TFPI levels [26], hence Vitamin D could exert anticoagulant effects.

An inflammatory phase

The next phase of wound healing is inflammation: the injured epithelial or endothelial cells release excessive inflammatory mediators, inducing the sequential infiltration of inflammatory cells (neutrophils, macrophages and lymphocytes). Macrophages can release cytokines IL-13, IL-1, growth factors such as active transforming growth factor β (TGF- β), platelet-derived growth factor (PDGF), fibroblast growth factor (FGF) and TNF- α , which promote the inflammatory response and fibrosis [5, 30-33]. These intricate and poorly understood interactions ultimately result in myofibroblast activation and collagen expression, especially TGF- β , a multi-

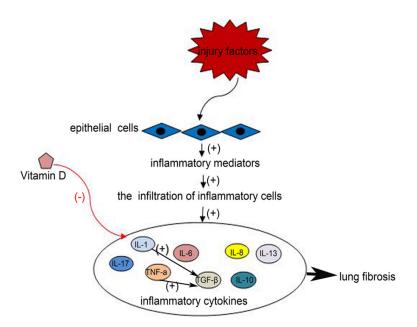


Figure 2. The injured epithelial cells release excessive inflammatory mediators, inducing the sequential infiltration of inflammatory cells. Then inflammatory cells release cytokines (IL-1, IL-6, IL-8, IL-10, IL-13, IL-17, TGF- β , TNF- α), which promote the inflammatory response and fibrosis. Vitamin D can reduce the levels of inflammatory cytokines, preventing the further expansion of the inflammatory response.

functional cytokine that is causally linked to pulmonary fibrosis.

Past research suggests that inflammatory mediators play a role in the initiation and progression of pulmonary fibrosis [34, 35]. However, now more studies are showing that pulmonary fibrosis is the consequence of multiple repeated injuries to the lung epithelium [5, 30]. The concept of pathogenesis for lung fibrosis has been transited inflammatory-driven into an epithelial-driven, and inflammation is not a cause, but a consequence, of pulmonary fibrosis [5, 30].

Vitamin D has been shown to cause decline in serum inflammatory cytokine levels (**Figure 2**), including IL-13 [36], IL-17 [36, 37], IL-1, IL-6, IL-8, and TNF- α [38], and may also act directly on CD4+ T cells to promote T-regulatory cells (Tregs) that secrete the anti-inflammatory cytokine IL-10 [36, 37, 39, 40], and prevent the further expansion of the inflammatory response.

Several studies have supported the notion that VD3 could markedly inhibit activation of TGF-β signaling pathways, diminish the up-regulation of fibronectin and collagen expression, and

also inhibit the trans-differentiation of TGF- $\beta1$ and stimulated lung epithelial cells into myofibroblasts [41, 42], and we will elaborate on this in the next section.

Fibroblast migration/proliferation/activation phase

After the inflammation, the wound healing process enters the next phase, where fibroblast hyperplasia and exaggerated ECM deposition is initiated, and more than one mechanism is involved in the fibrosis process.

TGF-β/SMAD signaling pathways: At the very beginning of lung damage, the injured epithelial or endothelial cells release excessive inflammatory mediators that start an antifibrinolytic-coagulation cascade that triggers clotting and

creates an interim ECM. Then it enters the next phase, characterized by a fibroblast migration/proliferation/activation phase, and myofibroblasts plays a significant role in this phase. Myofibroblasts are converted from a variety of sources including settled mesenchymal cells, bone marrow progenitors (also entitled fibrocytes). Epithelial cells go through epithelial-mesenchymal transition (EMT) [7]. EMT of epithelial cells is a major step toward pulmonary fibrosis, and TGF- β is an accepted activator of EMT, and also plays a central role in the proliferation, differentiation, and migration of cells [41, 43-46].

TGF- β expression can be induced by proinflammatory cytokines, such as IL-1 β and TNF- α , and IL-1 β triggers TGF- β gene expression by activating NF-B and AP-1 pathways [47]. TGF- β attaches to cell surface type I and II serine/threonine receptor kinases, leading to phosphorylation of SMAD2 and SMAD3, then the phosphorylation of receptor kinases released into the cytosol, and make up a complex with SMAD4. Followed by into the nucleus, the activated Smad complexes in conjunction with SMAD-binding elements within the genome to play its role, such

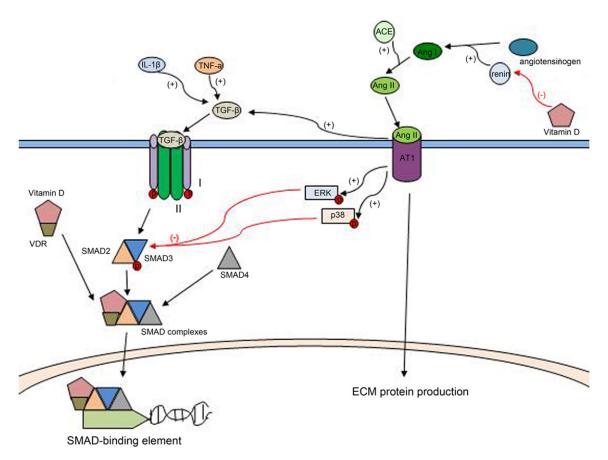


Figure 3. (1) TNF- α , and IL-1 β can induce the expression of TGF- β . TGF- β attaches to cell surface type I and II serine/threonine receptor kinases, leading to phosphorylation of SMAD2 and SMAD3, then makes up a complex with SMAD4. Followed by translocation into the nucleus, the activated SMAD complexes in conjunction with SMAD-binding elements and plays its role. Vitamin D binds a complex with VDR, then the complex directly interacts with SMAD3 and decreases binding of SMAD3 to DNA, at last resulting in the inhibited TGF- β -SMAD signaling pathways. (2) Renin cleaves angiotensinogen to angiotensin (Ang) I, which is further converted to Ang II by ACE. Ang II plays an important role in lung fibrogenesis by the AT1 receptor. Ang II is activated by AT1 receptor, directly stimulates ECM protein production, and elevates the expression of TGF- β , meanwhile it activates SMAD2/3 directly by the ERK/p38 pathway. Vitamin D can reduce the expression of rennin, and regulate TGF- β SMAD3 signaling, Vitamin D causes negative regulation of the RASboth of these two ways.

as regulating the expression of profibrotics [48, 49].

Studies have revealed that Vitamin D can inhibits the TGF- β -SMAD signaling pathway [41, 50-52], and the details of this process is 1,25(OH) $_2$ D $_3$ binds a complex with VDR, then the complex directly interacts with SMAD3 and decreased binding of SMAD3 to DNA, at last resulting in the inhibited TGF- β -SMAD signal transduction [42, 53].

Ang II-AT1 receptor signaling: The activation of renin-angiotensin system (RAS) has been implicated to induce lung fibrosis both in transgenic animals and in disease models [54-56], and is

recognized one important pathogenic factor in the pathogenesis of lung fibrosis [57]. However, the induction of lung fibrosis by RAS is not due to hypertension, although hypertension is an independent risk factor for lung fibrosis [58]. The RAS consists of angiotensinogen (AGT), an aspartyl protease such as renin or cathepsin D, angiotensin-converting enzyme (ACE). Renin cleaves angiotensinogen to angiotensin (Ang) I, which is further converted to Ang II by angiotensin-converting enzyme (ACE). Ang II plays an important role in lung fibrogenesis by both AT1 and AT2 receptors, which are mediated mainly by the AT1 receptor [57, 59, 60]. Ang II activated by AT1 receptor, directly stimulates ECM protein production, and expression of TGF-β and

connective tissue growth factor (CTGF) [61, 62], both of which could activate the fibrotic cascade and contribute to the development of lung fibrosis. Importantly, Ang II can activate SMAD2/3 directly by the ERK/p38 pathway [63] (64), and regulate TGF-β/SMAD3 signaling at multiple levels.

Vitamin D plays its antifibrotic role through a negative regulation of the RAS (**Figure 3**). Research has shown that hypovitaminosis D has been the other face of RAS activation [64], and VDR knockout mice caused the overexpression of renin, resulting in more angiotensinogen transformed into angiotensin II. In summary, vitamin D could reduce over-activation of RAS, causing an antifibrotic effect. Besides, 1,25(OH)₂D₃ binds a complex with VDR, and the complex directly interacts with SMAD3, which restrains the ERK/p38 pathway. Vitamin D plays its negative regulation of the RAS both of these two ways.

Discussion

A large number of studies have displayed the prominent role Vitamin D has in fibrosis disease. Vitamin D affects the progress of clotting/coagulation phase, inflammation phase, and fibroblast migration/proliferation/activation phase of pulmonary fibrosis, in many ways, and then plays its antifibrotic role.

However, the progress of pulmonary fibrosis is complex and results from different factors. More importantly, a variety of pathways could be responsible for lung fibrosis, not only TGF- β / SMAD signaling and Ang II-AT1 receptor signaling, but also other ways, such as NF- κ B signaling [65, 66], WNT and β -catenin signaling pathways [67], and so on; which limits the antifibrotic role of Vitamin D.

Nevertheless, the evidence reviewed in this paper indicates a crucial role of Vitamin D in lung fibrosis.

Acknowledgements

This work was supported by State Key Development Program of Shandong Province, China, under Grant number 2017GSF221003.

Disclosure of conflict of interest

None.

Address correspondence to: Lipan Peng, Department of Gastrointestinal Surgery, Shandong Provincial Hospital Affiliated to Shandong University, Jinan 250000, Shandong, China. Tel: +86-150-53102101; E-mail: 15053102101@163.com

References

- [1] Raghu G, Collard HR, Egan JJ, Martinez FJ, Behr J, Brown KK, Colby TV, Cordier JF, Flaherty KR, Lasky JA, Lynch DA, Ryu JH, Swigris JJ, Wells AU, Ancochea J, Bouros D, Carvalho C, Costabel U, Ebina M, Hansell DM, Johkoh T, Kim DS, King TE Jr, Kondoh Y, Myers J, Müller NL, Nicholson AG, Richeldi L, Selman M, Dudden RF, Griss BS, Protzko SL, Schünemann HJ; ATS/ERS/JRS/ALAT Committee on Idiopathic Pulmonary Fibrosis. An official ATS/ERS/JRS/ALAT statement: idiopathic pulmonary fibrosis: evidence-based guidelines for diagnosis and management. Am J Respir Crit Care Med 2011; 183: 788-824.
- [2] Sundarakrishnan A, Chen Y, Black LD, Aldridge BB and Kaplan DL. Engineered cell and tissue models of pulmonary fibrosis. Adv Drug Deliv Rev 2018; 129: 78-94.
- [3] Ahluwalia N, Shea BS and Tager AM. New therapeutic targets in idiopathic pulmonary fibrosis. Aiming to rein in runaway wound-healing responses. Am J Respir Crit Care Med 2014; 190: 867-78.
- [4] Wolters PJ, Blackwell TS, Eickelberg O, Loyd JE, Kaminski N, Jenkins G, Maher TM, Molina-Molina M, Noble PW, Raghu G, Richeldi L, Schwarz MI, Selman M, Wuyts WA and Schwartz DA. Time for a change: is idiopathic pulmonary fibrosis still idiopathic and only fibrotic? Lancet Respir Med 2018; 6: 154-160.
- [5] Liu YM, Nepali K and Liou JP. Idiopathic pulmonary fibrosis: current status, recent progress, and emerging targets. J Med Chem 2017; 60: 527-553.
- [6] Selman M, King TE, Pardo A. Idiopathic pulmonary fibrosis: prevailing and evolving hypotheses about its pathogenesis and implications for therapy. Ann Intern Med 2001; 134: 136-51
- [7] Wynn TA. Integrating mechanisms of pulmonary fibrosis. J Exp Med 2011; 208: 1339-50.
- [8] Demay MB. Mechanism of vitamin D receptor action. Ann N Y Acad Sci 2006; 1068: 204-13.
- [9] Reid IR, Bolland MJ and Grey A. Effects of vitamin D supplements on bone mineral density: a systematic review and meta-analysis. Lancet 2014; 383: 146-155.
- [10] Hollis BW and Wagner CL. Clinical review: the role of the parent compound vitamin D with respect to metabolism and function: why clinical

- dose intervals can affect clinical outcomes. J Clin Endocrinol Metab 2013; 98: 4619-28.
- [11] Flanagan JN, Young MV, Persons KS, Wang L, Mathieu JS, Whitlatch LW, Holick MF, Chen TC. Vitamin D metabolism in human prostate cells: implications for prostate cancer chemoprevention by vitamin D. Anticancer Res 2006; 26: 2567-2572.
- [12] Hosseinpour F and Wikvall K. Porcine microsomal vitamin D(3) 25-hydroxylase (CYP2D25). Catalytic properties, tissue distribution, and comparison with human CYP2D6. J Biol Chem 2000; 275: 34650-5.
- [13] Schuessler M, Astecker N, Herzig G, Vorisek G, Schuster I. Skin is an autonomous organ in synthesis, two-step activation and degradation of vitamin D(3): CYP27 in epidermis completes the set of essential vitamin D(3)-hydroxylases. Steroids 2001; 66: 399-408.
- [14] Zhu J and DeLuca HF. Vitamin D 25-hydroxylase-four decades of searching, are we there yet? Arch Biochem Biophys 2012; 523: 30-6.
- [15] Haussler MR, Whitfield GK, Haussler CA, Hsieh JC, Thompson PD, Selznick SH, Dominguez CE, Jurutka PW. The nuclear vitamin D receptor: biological and molecular regulatory properties revealed. J Bone Miner Res 1998; 13: 325-349.
- [16] Prietl B, Treiber G, Pieber TR and Amrein K. Vitamin D and immune function. Nutrients 2013; 5: 2502-21.
- [17] Samuel S and Sitrin MD. Vitamin D's role in cell proliferation and differentiation. Nutr Rev 2008; 66: S116-24.
- [18] Norman PE and Powell JT. Vitamin D and cardiovascular disease. Circ Res 2014; 114: 379-93.
- [19] Park YS, Park CM, Lee HJ, Goo JM, Chung DH, Lee SM, Yim JJ, Kim YW, Han SK and Yoo CG. Clinical implication of protease-activated receptor-2 in idiopathic pulmonary fibrosis. Respir Med 2013; 107: 256-62.
- [20] Imokawa S, Sato A, Hayakawa H, Kotani M, Urano T, Takada A. Tissue factor expression and fibrin deposition in the lungs of patients with idiopathic pulmonary fibrosis and systemic sclerosis. Am J Respir Crit Care Med 1997; 156: 631-6.
- [21] Lin C, Borensztajn K and Spek CA. Targeting coagulation factor receptors-protease-activated receptors in idiopathic pulmonary fibrosis. J Thromb Haemost 2017; 15: 597-607.
- [22] Borensztajn K, Stiekema J, Nijmeijer S, Reitsma PH, Peppelenbosch MP and Spek CA. Factor Xa stimulates proinflammatory and profibrotic responses in fibroblasts via protease-activated receptor-2 activation. Am J Pathol 2008; 172: 309-20.
- [23] Wygrecka M, Kwapiszewska G, Jablonska E, von Gerlach S, Henneke I, Zakrzewicz D, Guen-

- ther A, Preissner KT and Markart P. Role of protease-activated receptor-2 in idiopathic pulmonary fibrosis. Am J Respir Crit Care Med 2011; 183: 1703-14.
- [24] Ruf W, Mueller BM. Tissue factor signaling. Thromb Haemost 1999; 82: 175-182.
- [25] Schaffner F and Ruf W. Tissue factor and protease-activated receptor signaling in cancer. Semin Thromb Hemost 2008; 34: 147-53.
- [26] Kasthuri RS, Glover SL, Boles J and Mackman N. Tissue factor and tissue factor pathway inhibitor as key regulators of global hemostasis: measurement of their levels in coagulation assays. Semin Thromb Hemost 2010; 36: 764-71.
- [27] Koyama T, Shibakura M, Ohsawa M, Kamiyama R, Hirosawa S. Anticoagulant effects of 1alpha,25-dihydroxyVitamin D3 on human myelogenous leukemia cells and monocytes. Blood 1998; 92: 160-167.
- [28] Ohsawa M, Koyama T, Yamamoto K, Hirosawa S, Kamei S and Kamiyama R. 1alpha,25-dihydroxyvitamin D(3) and its potent synthetic analogs downregulate tissue factor and upregulate thrombomodulin expression in monocytic cells, counteracting the effects of tumor necrosis factor and oxidized LDL. Circulation 2000; 102: 2867-2872.
- [29] Martinez-Moreno JM, Herencia C, Montes de Oca A, Munoz-Castaneda JR, Rodriguez-Ortiz ME, Diaz-Tocados JM, Peralbo-Santaella E, Camargo A, Canalejo A, Rodriguez M, Velasco-Gimena F and Almaden Y. Vitamin D modulates tissue factor and protease-activated receptor 2 expression in vascular smooth muscle cells. FASEB J 2016; 30: 1367-76.
- [30] Noble PW. Idiopathic pulmonary fibrosis. New insights into classification and pathogenesis usher in a new era therapeutic approaches. Am J Respir Cell Mol Biol 2003; 29: S27-31.
- [31] Khalil N and O'Connor R. Idiopathic pulmonary fibrosis: current understanding of the pathogenesis and the status of treatment. CMAJ 2004; 171: 153-60.
- [32] Selman M, Pardo A. The epithelial/fibroblastic pathway in the pathogenesis of idiopathic pulmonary fibrosis. Am J Respir Cell Mol Biol 2003; 29: S93-7.
- [33] Plataki M, Koutsopoulos AV, Darivianaki K, Delides G, Siafakas NM, Bouros D. Expression of apoptotic and antiapoptotic markers in epithelial cells in idiopathic pulmonary fibrosis. Chest 2005; 127: 266-74.
- [34] Bringardner BD, Baran CP, Eubank TD and Marsh CB. The role of inflammation in the pathogenesis of idiopathic pulmonary fibrosis. Antioxid Redox Signal 2008; 10: 287-301.
- [35] Mura M, Belmonte G, Fanti S, Contini P, Pacilli AM, Fasano L, Zompatori M, Schiavina M, Fabbri M. Inflammatory activity is still present in

Vitamin D and pulmonary fibrosis

- the advanced stages of idiopathic pulmonary fibrosis. Respirology 2005; 10: 609-14.
- [36] Brown SD, Calvert HH and Fitzpatrick AM. Vitamin D and asthma. Dermatoendocrinol 2012; 4: 137-45.
- [37] Paul G, Brehm JM, Alcorn JF, Holguin F, Aujla SJ and Celedon JC. Vitamin D and asthma. Am J Respir Crit Care Med 2012; 185: 124-32.
- [38] Stubbs JR, Idiculla A, Slusser J, Menard R and Quarles LD. Cholecalciferol supplementation alters calcitriol-responsive monocyte proteins and decreases inflammatory cytokines in ESRD. J Am Soc Nephrol 2010; 21: 353-61.
- [39] Herr C, Greulich T, Koczulla RA, Meyer S, Zakharkina T, Branscheidt M, Eschmann R and Bals R. The role of vitamin D in pulmonary disease: COPD, asthma, infection, and cancer. Respir Res 2011; 12: 31.
- [40] Finklea JD, Grossmann RE and Tangpricha V. Vitamin D and chronic lung disease: a review of molecular mechanisms and clinical studies. Adv Nutr 2011; 2: 244-53.
- [41] Fischer KD, Agrawal DK. Erratum to: vitamin D regulating TGF-β induced epithelial-mesenchymal transition. Respir Res 2015; 16: 139.
- [42] Jiang F, Yang Y, Xue L, Li B and Zhang Z. 1α,25-dihydroxyvitamin D3 attenuates TGF-β-induced pro-fibrotic effects in human lung epithelial cells through inhibition of epithelial-mesenchymal transition. Nutrients 2017; 9.
- [43] Sun X, Chen E, Dong R, Chen W and Hu Y. Nuclear factor (NF)-κB p65 regulates differentiation of human and mouse lung fibroblasts mediated by TGF-β. Life Sci 2015; 122: 8-14.
- [44] Abramovitch S, Sharvit E, Weisman Y, Bentov A, Brazowski E, Cohen G, Volovelsky O and Reif S. Vitamin D inhibits development of liver fibrosis in an animal model but cannot ameliorate established cirrhosis. Am J Physiol Gastrointest Liver Physiol 2015; 308: G112-20.
- [45] Sporn MB, Roberts AB, Wakefield LM, Assoian RK. Transforming growth factor-beta: biological function and chemical structure. Science 1986; 233: 532-4.
- [46] Massague J. The transforming growth factorbeta family. Annu Rev Cell Biol 1990; 6: 597-641.
- [47] Lee KY, Ito K, Hayashi R, Jazrawi EP, Barnes PJ, Adcock IM. NF-kappaB and activator protein 1 response elements and the role of histone modifications in IL-1beta-induced TGF-beta1 gene transcription. J Immunol 2006; 176: 603-15.
- [48] Bonventre JV. Antifibrotic vitamin D analogs. J Clin Invest 2013; 123: 4570-3.
- [49] Huang LS and Natarajan V. Sphingolipids in pulmonary fibrosis. Adv Biol Regul 2015; 57: 55-63.

- [50] Artaza JN and Norris KC. Vitamin D reduces the expression of collagen and key profibrotic factors by inducing an antifibrotic phenotype in mesenchymal multipotent cells. J Endocrinol 2009; 200: 207-21.
- [51] Wang L, Yuan T, Du G, Zhao Q, Ma L and Zhu J. The impact of 1,25-dihydroxyvitamin D3 on the expression of connective tissue growth factor and transforming growth factor-beta1 in the myocardium of rats with diabetes. Diabetes Res Clin Pract 2014; 104: 226-33.
- [52] Kabel AM, Abd Elmaaboud MA, Atef A and Baali MH. Ameliorative potential of linagliptin and/ or calcipotriol on bleomycin-induced lung fibrosis: in vivo and in vitro study. Environ Toxicol Pharmacol 2017; 50: 216-226.
- [53] Ito I, Waku T, Aoki M, Abe R, Nagai Y, Watanabe T, Nakajima Y, Ohkido I, Yokoyama K, Miyachi H, Shimizu T, Murayama A, Kishimoto H, Nagasawa K and Yanagisawa J. A nonclassical vitamin D receptor pathway suppresses renal fibrosis. J Clin Invest 2013; 123: 4579-94.
- [54] Wang J, Chen L, Chen B, Meliton A, Liu SQ, Shi Y, Liu T, Deb DK, Solway J and Li YC. Chronic activation of the renin-angiotensin system induces lung fibrosis. Sci Rep 2015; 5: 15561.
- [55] Jiang JS, Lang YD, Chou HC, Shih CM, Wu MY, Chen CM and Wang LF. Activation of the reninangiotensin system in hyperoxia-induced lung fibrosis in neonatal rats. Neonatology 2012; 101: 47-54.
- [56] Lang YD, Hung CL, Wu TY, Wang LF and Chen CM. The renin-angiotensin system mediates hyperoxia-induced collagen production in human lung fibroblasts. Free Radic Biol Med 2010; 49: 88-95.
- [57] Uhal BD, Li X, Piasecki CC and Molina-Molina M. Angiotensin signalling in pulmonary fibrosis. Int J Biochem Cell Biol 2012; 44: 465-8.
- [58] Shi Y, Liu T, Yao L, Xing Y, Zhao X, Fu J and Xue X. Chronic vitamin D deficiency induces lung fibrosis through activation of the renin-angiotensin system. Sci Rep 2017; 7: 3312.
- [59] Konigshoff M, Wilhelm A, Jahn A, Sedding D, Amarie OV, Eul B, Seeger W, Fink L, Gunther A, Eickelberg O and Rose F. The angiotensin II receptor 2 is expressed and mediates angiotensin II signaling in lung fibrosis. Am J Respir Cell Mol Biol 2007; 37: 640-50.
- [60] Marshall RP, McAnulty RJ, Laurent GJ. Angiotensin II is mitogenic for human lung fibroblasts via activation of the type 1 receptor. Am J Respir Crit Care Med 2000; 161: 1999-2004.
- [61] Finckenberg P, Inkinen K, Ahonen J, Merasto S, Louhelainen M, Vapaatalo H, Müller D, Ganten D, Luft F and Mervaala E. Angiotensin II induces connective tissue growth factor gene expression via calcineurin-dependent pathways. Am J Pathol 2003; 163: 355-66.

Vitamin D and pulmonary fibrosis

- [62] Marshall RP, Gohlke P, Chambers RC, Howell DC, Bottoms SE, Unger T, McAnulty RJ and Laurent GJ. Angiotensin II and the fibroproliferative response to acute lung injury. Am J Physiol Lung Cell Mol Physiol 2004; 286: L156-64.
- [63] Mirkovic K and de Borst MH. Beyond the RAAS: dissecting the antifibrotic effects of vitamin D analogues. Lab Invest 2012; 92: 1666-9.
- [64] Ferder M, Inserra F, Manucha W and Ferder L. The world pandemic of vitamin D deficiency could possibly be explained by cellular inflammatory response activity induced by the reninangiotensin system. Am J Physiol Cell Physiol 2013; 304: C1027-39.
- [65] Hou J, Ma T, Cao H, Chen Y, Wang C, Chen X, Xiang Z and Han X. TNF-alpha-induced NF-kappaB activation promotes myofibroblast differentiation of LR-MSCs and exacerbates bleomycin-induced pulmonary fibrosis. J Cell Physiol 2018; 233: 2409-2419.

- [66] Kandhare AD, Bodhankar SL, Mohan V and Thakurdesai PA. Effect of glycosides based standardized fenugreek seed extract in bleomycin-induced pulmonary fibrosis in rats: decisive role of Bax, Nrf2, NF-kappaB, Muc5ac, TNF-alpha and IL-1beta. Chem Biol Interact 2015; 237: 151-65.
- [67] Moon RT, Kohn AD, De Ferrari GV and Kaykas A. WNT and beta-catenin signalling: diseases and therapies. Nat Rev Genet 2004; 5: 691-701.