**Vitamin D in combination cancer treatment**

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Published: 2010.07.15

**Abstract**

As a steroid hormone that regulates mineral homeostasis and bone metabolism, 1α, 25-dihydroxycholecalciferol (calcitriol) also has broad spectrum anti-tumor activities as supported by numerous epidemiological and experimental studies. Calcitriol potentiates the anti-tumor activities of multiple chemotherapeutics agents including DNA-damaging agents cisplatin, carboplatin and doxorubicin; antimetabolites 5-fluorouracil, cytarabine, hydroxyurea, cytarabine and gemcitabine; and microtubule-disturbing agents paclitaxel and docetaxel. Calcitriol elicits anti-tumor effects mainly through the induction of cancer cell apoptosis, cell cycle arrest, differentiation, angiogenesis and the inhibition of cell invasiveness by a number of mechanisms. Calcitriol enhances the cytotoxic effects of gamma irradiation and certain antioxidants and naturally derived compounds. Inhibition of calcitriol metabolism by 24-hydroxylase promotes growth inhibition effect of calcitriol. Calcitriol has been used in a number of clinical trials and it is important to note that sufficient dose and exposure to calcitriol is critical to achieve anti-tumor effect. Several trials have demonstrated that safe and feasible to administer high doses of calcitriol through intermittent regimen. Further well designed clinical trials should be conducted to better understand the role of calcitriol in cancer therapy.

Key words: vitamin D, calcitriol, cancer, chemotherapy

**Introduction**

Vitamin D is a steroid hormone that regulates calcium homeostasis, bone metabolism and a variety of other physiological functions (1). Vitamin D can be obtained from ultraviolet light-induced photobiogenesis in the skin or from the diet (1). In the skin, 7-dehydrocholesterol is converted to vitamin D₃, which is hydroxylated to 25(OH)D₃ by 25-hydroxylase in the liver and then to 1,25(OH)₂D₃ (1α, 25-dihydroxycholecalciferol, calcitriol), the hormonally active metabolite, by 1α-hydroxylase in the kidney (1). Calcitriol is mainly catabolized by 24-hydroxylase (CYP24A1) to 1α,24, 25(OH)₃D₃ which loses its bioactivity (1).

Calcitriol acts through both genomic and non-genomic mechanisms (2). In genomic pathways, calcitriol binds to intracellular vitamin D receptor (VDR), which subsequently heterodimerizes with another nuclear receptor retinoid X receptor (RXR). The heterodimer binds to vitamin D response element in target genes and leads to gene transcription regulation (1). In addition, calcitriol has rapid effects that are independent of gene transcription regulation, which are defined as non-genomic effects and not mediated directly through steroid receptor-ligand-DNA interaction. On the other hand, non-genomic actions may indirectly affect gene transcription via the regulation of intracellular signaling pathways that target transcription factors (3). Calcitriol induces a number of non-genomic responses including rapid intestinal absorption of calcium, release
of calcium from intracellular stores, opening of voltage-gated calcium and chloride channels, and the activation of protein kinase C, protein kinase A, phosphatidylinositol-3 kinase (PI3K) and phospholipase C (4).

**Vitamin D compounds in cancer prevention and treatment**

Numerous epidemiological and preclinical studies support a role of vitamin D compounds in cancer prevention and treatment in colorectal, breast, prostate, ovarian, bladder, lung and skin cancers and leukemia (1, 5, 6). Low levels of plasma 25(OH)D3 are associated with higher cancer incidence and mortality in men in colorectal, breast, lung and prostate cancers (7-10). The broad spectrum anti-tumor effects of calcitriol and analogs are mostly based on inhibition of cancer cell proliferation and invasiveness, induction of differentiation and apoptosis, and promotion of angiogenesis.

Calcitriol and analogs are mostly based on inhibition of calcitriol and analogs are mostly based on inhibition of calcium-sensing receptor (23). When added together or immediately after ara-C (cytarabine), calcitriol promotes the accumulation of DNA fragments and cytotoxicity (24). Calcitriol and cytarabine combination has been used in clinic as the minimally intensive chemotherapy, and prolonged remission in elderly patients with acute myeloid leukemia (AML) and myelodysplastic syndrome (MDS) (25, 26). Adenosine deaminase-resistant analog fludarabine synergistically enhances calcitriol-induced differentiation of human monoblastic leukemia U937 cells (27). Hydroxyurea, cytarabine or camptothecin acts synergistically with calcitriol to inhibit human monoblastic leukemia U937 cell growth (28). Hydroxyurea also promotes calcitriol-mediated U937 cell differentiation (28). Gemcitabine is a widely used antimetabolite, and the combination of gemcitabine and cisplatin is the current standard chemotherapy regimen for locally advanced and metastatic bladder cancer (29, 30). Calcitriol enhances caspase-dependent apoptosis and synergistically promotes the anti-proliferative effects of gemcitabine and cisplatin in human bladder cancer model systems T24 and UMUC3 (31). We have shown in vitro in SCC cells that p73 is important in calcitriol antiproliferative effect; we are also examining p73 status in vivo in human transitional cell carcinoma. The expression of p73 protein is lower in human bladder cancer tissue compared with adjacent normal tissue in 3 out of 4 pairs as assessed by immunoblot analysis (31). Calcitriol augments p73 protein level in vivo in human transitional cell carcinoma.
with calcitriol in combination with gemcitabine and cisplatin markedly inhibits T24 tumor growth in nude mice (31). Anti-tumor activity of gemcitabine is also augmented by calcitriol in Capan-1 human pancreatic cancer model system, as suggested by enhanced growth inhibition, apoptosis, inhibition of Akt survival pathway and xenograft tumor growth (32).

Calcitriol potentiates antitumor activity of microtubule-disrupting agents such as paclitaxel (33, 34) and docetaxel (35). This effect is associated with reduced expression level of p21 in prostate cancer cell PC3 (33) or increased Bcl-2 phosphorylation in breast cancer cells (34), and multidrug resistance-associated protein 1 (35), respectively. Calcitriol analog 1,25(OH)2-16-ene-23-yne-19-nor-26,27-F6-D3 (LT) or EB1089 also potentiates antitumor activity of paclitaxel in breast cancer model systems (36). Calcitriol analog ILX 23-7553 additively enhances the antitumor effects of both adriamycin and ionizing irradiation in breast tumor cells MCF-7 through growth inhibition and apoptosis induction (37). These substantial data suggest that the addition of calcitriol to multiple chemotherapy regimens increases the activity of such treatments and potentially a better response rate to the regimens.

VDR forms heterodimer in association with RXR. RXR ligand 9-cis-retinoic acid (9-cis-RA) combined with calcitriol results in delayed tumor progression in the prostate PC3 tumor xenograft model in nude mice (38). This combination treatment results in direct binding of VDR/RXR heterodimer to the promoter region of human telomerase reverse transcriptase (hTERT) which inhibits the expression of hTERT and subsequently leads to decreased telomerase activity in prostate cancer cells (38). Calcitriol analog 20-epi-22oxa-24a,26a,27a-tri-homo-1α,25(OH)2D3 (KH1060) synergizes with 9-cis-RA to inhibit the growth and promote the differentiation of acute promyelocytic leukemia cells NB4 (39) and myeloblastic cells HL-60 (40). The combination leads to increased apoptosis which is accompanied by reduced Bcl-2 expression and increased Bax expression (39).

The antitumor activity of calcitriol may also involve histone deacetylation. Combining histone deacetylase inhibitor sodium butyrate or trichostatin A with calcitriol or its analog LH or 1α,25-(OH)2-16,23E-diene-26,27-hexafluoride-D3 (LT) synergistically suppresses calcitriol or analog-induced growth inhibition in prostate cancer cell lines LNCaP, PC3 and DU145 (41). This effect is mediated by enhanced apoptosis instead of induction of cell cycle arrest (41).

Besides chemotherapy, vitamin D is also used in combination with other types of cancer treatment. Calcitriol or its less calcaemic analogue 19-nor-1α,25-(OH)2D3 acts synergistically with ionizing radiation to inhibit the growth and apoptosis of LNCaP prostate cancer cells and primary tumor cells (42). EB1089 potentiates the antitumor activity of ionizing radiation partially through increased apoptosis in breast cancer model system MCF-7 (43). Breast cancer cells overexpress one of the NF-κB subunits RelB, which promotes cancer cell survival. Calcitriol treatment results in reduced mRNA and protein levels of RelB and its target genes survivin, Bcl-2 and MnSOD, and sensitizes the breast cancer cells Hs578T and NF639 to gamma-irradiation. Overexpression of RelB enhances NF639 cells survival following calcitriol and irradiation treatment (44). Calcitriol pretreatment for 24 h enhances the phototoxic response of human SCC A431 cells to methyl aminolaevulinate-based photodynamic therapy (45).

Non-specific cyclooxygenase (COX) inhibitors acetyl salicylic acid or indomethacin in combination with calcitriol markedly induce differentiation of leukemia cell lines into monocytes and cell cycle arrest at G1 phase (46). Cell differentiation is dependent on phosphorylation of Raf1 (46). The combination of ibuprofen, a non-steroidal anti-inflammatory drug (NSAID), and calcitriol results in greater growth inhibition and G1 cell cycle arrest in human prostate cancer LNCaP cells compared to either agent alone (47). A calcitriol analog 22-oxa-1α,25-(OH)2D3, when used together with vitamin K2, promotes leukemia cells HL-60 differentiate into monocytes as examined by morphology and cell surface CD14 expression in a synergistic nature (48). This combination also induces cell cycle arrest at G0/G1 phase; however, it suppresses apoptosis compared to vitamin K2 alone (48). Carnosic acid, a plant-derived polyphenolic antioxidant, enhances the monocytic differentiation effects of calcitriol in human myeloid leukemia cells HL60 (49). Decreased intracellular reactive oxygen species, increased intracellular glutathione, and activation of Raf-1/MEK1/ERK1/2 pathway are observed with the combination treatment (49). Bryostatin-1, a marine bryozoan-derived natural compound, has antitumor activities in both solid and lymphoid tumors (50). Bryostatin-1 synergizes with calcitriol to induce monocytic differentiation of NB4 cells (50, 51), which is associated with G1 phase cell cycle arrest, decreased cell growth and increased plastic adhesion (50). 25(OH)D3, when used together with iron deprivation agents including iron chelators or transferrin receptor antibody A24, induces the differentiation of myeloid leukemia cell lines and primary myeloblasts from AML patients into monocytes/macrophages (52). These effects are dependent on the increased level of
reactive oxygen species and the activation of JNK MAPK pathway (52).

Interaction of calcitriol with certain agents results in enhanced calcitriol anti-tumor activity. Administration of dexamethasone (Dex), which reduces calcitriol-induced hypercalcemia, prior to calcitriol inhibits SCC cell proliferation compared to calcitriol alone (53). Combined treatment with Dex and calcitriol reduces SCC xenograft tumor growth. These findings are associated with the observations that Dex enhances VDR expression in SCC cells and VDR ligand binding activities in tumor cell extracts and kidneys but decreases that in intestinal mucosa (53). Further studies show that the combination of calcitriol and Dex apoptosis and cell cycle arrest at G0/G1 in SCC cells (54). This combination also suppresses the activation of Akt and ERK1/2 pathways (54).

The effect of calcitriol is modulated by its metabolizing enzymes. The primary vitamin D3 inactivating enzyme CYP24A1, a mitochondrial cytochrome P450, induces calcitriol degradation and thereby inhibits calcitriol biological activity. The broad spectrum cytochrome P450 inhibitor ketoconazole (KTZ) or a specific CYP24A1 inhibitor RC2204, which effectively inhibits the expression and enzyme activity of CYP24A1 in PC3 cells and mice kidney tissue, synergistically inhibits the anti-proliferative effect of calcitriol in human prostate PC3 cells (55). Dex is administered together with KTZ to minimize calcitriol-mediated hypercalcemia. Enhanced apoptosis is observed which does not involve caspase 3 activation but the translocation of apoptosis inducing factor (AIF) to the nucleus. Calcitriol and ketoconazole/Dex combination enhances the growth inhibition observed with calcitriol alone in PC3 xenograft tumor mouse model (55). KTZ also potentiates the anti-proliferative effect of calcitriol or its analog EB1089 in prostate cancer cells (56). An imidazole derivative liarozole inhibits CYP24 activity in prostate cancer cells DU145 and thus sensitizes these cells to calcitriol-mediated growth inhibition, which is associated with increased VDR expression (57).

RRR-alpha-vitamin E succinate (VES), one of the most effective vitamin E forms, induces VDR expression in prostate cancer cells (58). Pretreatment with VES synergistically enhances calcitriol-mediated growth inhibition of prostate cancer cells in vitro and reduces the rate of prostate cancer xenograft tumor growth (58), which allows for a low-dose calcitriol to be administered. A glutathione-depleting compound, menadione, sensitizes breast cancer cells MCF-7 to calcitriol-mediated growth inhibition, which may be caused, at least in part, by the increased oxidative stress, as shown by enhanced ROS production (59).

Genistein, an isoflavone found in soybeans and a number of plants, in combination with calcitriol, inhibits cell growth in human prostate LNCaP cells, which is dependent on increased expression of p21 and associated with increased VDR expression (60). Another study shows that genistein and calcitriol further reduce prostate DU145 cell proliferation compared to either agent alone (61). The mechanisms for this effect may involve the induction of mRNA expression and enzyme activity of CYP24 by genistein which leads to prolonged half-life of calcitriol (61). Increased expression of VDR protein, VDR transcriptional activity, and the expression of VDR target genes are also observed in the combination treatment group (61). A medicinal herb ginseng (Panax ginseng C.A. Meyer, Araliaceae) promotes calcitriol-induced differentiation of leukemia cells HL-60 into monocytes as assessed by expression levels of CD14 and CD11b (62). This effect may be mediated by the ERK1/2 and PKC, but not PI3K pathway (62).

Phosphorylated prolactin (PRL) antagonizes the proliferation promoting effect of unmodified PRL. Molecular mimicry of naturally phosphorylated human PRL at the major phosphorylation site S179, S179D (PRL), sensitizes relatively vitamin D-insensitive prostate cancer cells DU-145 and PC3 to calcitriol-mediated anti-proliferative effect and apoptosis, which is associated with increased VDR and p21 expression (63). Secreted protein acidic and rich in cysteine (SPARC; osteonectin, BM-40), a family member of matricellular proteins including thrombospondins, tenasin, and osteopontin, may serve as a tumor suppressor (64). Compared with parental MIP101 colorectal cancer cells, calcitriol markedly reduces cell growth and enhances calcitriol alone- or calcitriol+fluorouracil-induced apoptosis in SPARC-overexpressing MIP101 cells (64). Calcitriol treatment also suppresses the phosphorylation of Akt and the expression of Bcl-2 family member BAD (64).

Calcitriol has been utilized in a number of clinical studies, either alone or in combination with Dex or cytotoxic agents, which has been reviewed (65, 66). It is important to emphasize that anti-tumor activity of calcitriol is dependent on its dose and exposure according to preclinical studies. Exposure to high concentrations of calcitriol is necessary to achieve anti-tumor results. Calcitriol or its analog Ro23-7553 delays SCC xenograft tumor growth in a dose dependent manner (67). Calcitriol of 2.5 μg/mouse administered twice a week results in a markedly stronger tumor suppression compared with once a week regimen in human pancreatic cancer model system Capan-1 (32). Pharmacokinetic (PK) studies indicate that calcitriol of 0.125 μg/mouse results in a
C\textsubscript{max} > 10.0 ng/ml and AUC > 40.0 ng h/mL in normal mice (68), which exceeds the concentration needed for calcitriol anti-tumor activity in vitro.

Although multiple clinical studies have been conducted with calcitriol or its analogs, the anti-tumor results are largely disappointing. This may be due to the fact that calcitriol or its analogs has been used at much lower doses than maximum tolerated dose (MTD) with the concern of dose-limiting hypercalcemia (69, 70). We and others demonstrate that sufficient doses of calcitriol to achieve exposure similar to those seen in preclinical models can be safely administered by high dose intermittent regimen (once weekly or QDx3 weekly) (69, 71-73). A recent phase I clinical trial demonstrates that the MTD of calcitriol (i.v.) is 74 μg/week when administered with gefitinib (70). The C\textsubscript{max} of calcitriol at the MTD is 6.68 ± 1.42 ng/ml (16 ± 3.40 nmol/L), which is much higher than the dose needed to elicit anti-tumor effect in preclinical studies (70). The area under the curve (AUC) of calcitriol at the MTD is 35.65 ± 8.01 ng h/mL (70). In comparison, 75 μg of DN101, a weekly oral formulation of calcitriol, results in a lower C\textsubscript{max} (3.8 nmol/L) but similar AUC (38.4 ng h/mL) (74). These results show that high doses of calcitriol can be administered alone or in combination with other agents to elicit or enhance the anti-tumor effects.

**Summary**

In summary, calcitriol has shown potential in enhancing the antitumor activities of a variety of cytotoxic or differentiating agents. The combination treatment studies with calcitriol do provide evidence and support for the continued study of calcitriol in cancer chemotherapies.

**Conflict of Interest**

The authors have declared that no conflict of interest exists.

**References**


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