Low Vitamin D Status: Definition, Prevalence, Consequences, and Correction

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KEYWORDS

- Vitamin D 25-Hydroxyvitamin D Supplementation
- Deficiency
 Insufficiency

Vitamin D is obtained either by ingestion or cutaneous production. When skin is exposed to ultraviolet B radiation, 7-dehydrocholesterol is converted to vitamin D_3 (cholecalciferol). Dietary sources may provide either vitamin D_3 or vitamin D_2 (ergocalciferol). However, few foods contain appreciable amounts of vitamin D, as such dietary intake is often low. Combining low intake with indoor lifestyle and sun-avoiding behaviors including sunscreen use, it is not surprising that low vitamin D status is endemic. The skeletal health consequences of vitamin D deficiency (calcium malabsorption and skeletal fragility) have long been recognized. More recently it has become appreciated that low vitamin D status leads to muscle weakness, falls, and potentially a multitude of nonskeletal morbidities. This review considers the definition and prevalence, potential health consequences, and approaches to correcting low vitamin D status.

VITAMIN D BACKGROUND AND ASSESSMENT

Vitamin D must be metabolized to become physiologically active. Specifically, vitamin D (either D_2 or D_3) is converted to 25-hydroxyvitamin D (25(OH)D) in the liver and subsequently to the active or "hormonal" form, 1,25-dihydroxyvitamin D (1,25(OH)₂D) in the kidneys.⁹ Measurement of 25(OH)D is the accepted indicator of an individual's vitamin D status.¹⁰ This evaluation is not intuitive, as it would seem logical that measurement of the active form, 1,25(OH)₂D, would be the appropriate measure of an individual's vitamin D status. It is not rare anecdotally to see health

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care providers obtain measurement of $1,25(OH)_2D$ purportedly to evaluate an individual patient's vitamin D status. However, measurement of circulating $1,25(OH)_2D$ does not provide a useful assessment of an individual's vitamin D status, as vitamin D deficiency leads to parathyroid hormone (PTH) elevation, which enhances renal 1α -hydroxylase activity thereby promoting conversion of available 25(OH)D to $1,25(OH)_2D$. As 25(OH)D is present in much higher concentration than $1,25(OH)_2D$ (ng/mL vs pg/mL) given the enhanced conversion induced by PTH elevation, $1,25(OH)_2D$ may be normal even in the setting of low vitamin D status.

The clinical measurement of 25(OH)D has been problematic, with substantial variability present between laboratories. ^{11,12} It is not the purpose of this review to detail approaches to and challenges with vitamin D measurement; this topic is reviewed elsewhere in this issue. Suffice it to say that current evaluations find that clinical 25(OH)D measurement has improved, ¹³ allowing health care providers to have reasonable confidence in clinical 25(OH)D measurements. Moreover, the recent availability of standard reference materials from the National Institute of Standards and Technology seems destined to further improve between-laboratory agreement. However, despite 25(OH)D assay improvements, health care providers must appreciate that assay variability is present for all laboratory results. The analytical imprecision and inaccuracy present in all quantitative medical procedures is due to method, human, and instrument limitations that confound application of rigid diagnostic cutpoint approaches. For example, if one were using a 25(OH)D value of 30 ng/mL to differentiate "low" from "optimal" vitamin D status, it must be recognized that a laboratory result of 29 ng/mL does not differ from 31 ng/mL. ¹⁴

LOW VITAMIN D STATUS: DEFINITION AND PREVALENCE

A spectrum of vitamin D status has been proposed wherein individuals whose serum 25(OH)D value is less than approximately 10 ng/mL are classified as deficient and may sustain impaired bone mineralization (rickets/osteomalacia), while those with a value less than approximately 30 ng/mL are identified as insufficient (**Fig. 1**) and may sustain long-term adverse health consequences. However, the cutpoint values selected, and even the verbiage to describe low vitamin D status, remain controversial. For example, terminology including deficiency, insufficiency, inadequacy, and hypovitaminosis has been variously, and interchangeably, applied to describe low vitamin D status. To avoid what seems to be a nonproductive debate, the terminology "low vitamin D status" is used here. Moreover, as noted above, 25(OH)D assay variability and absence of accepted standards has confounded agreement on a single definition of "low."

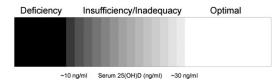


Fig. 1. Spectrum of vitamin D status. The spectrum of low vitamin D status is depicted. At very low vitamin D levels, (25(OH)D of approximately 10 ng/mL or less) calcium malabsorption, osteomalacia/rickets and myopathy occur. Less marked vitamin D deficiency (often referred to as inadequacy or insufficiency) has been associated with a variety of adverse health consequences. Consensus regarding an "optimal" 25(OH)D concentration continues to evolve; however, there seems to be increasing agreement that values greater than approximately 30 to 32 ng/mL are associated with optimal physiologic function.

Recognizing that controversy exists, there seems to be increasing consensus that circulating 25(OH)D values less than approximately 30 to 32 ng/mL indicate less than ideal vitamin D status. 16 These cutpoint values were suggested based on the long-established role of vitamin D to facilitate calcium and phosphorus absorption with deficiency leading to rickets/osteomalacia, 17-19 Thus, less severe vitamin D "deficiency" appears to cause calcium malabsorption, leading to secondary hyperparathyroidism with resulting elevated bone turnover and ultimately, bone loss.²⁰ The 25(OH)D to PTH relationship has been extensively reported, with various studies finding an apparent inflection point at around 20 to 30 ng/mL. 16,21 Moreover, some work demonstrates improved calcium absorption at 25(OH)D levels within what had previously been accepted as the "normal" range.²² However, others have challenged this seemingly cardinal tenant of low vitamin D status by reporting that calcium malabsorption does not occur until severe vitamin D deficiency is present, due to PTH-mediated maintenance of 1,25-dihydroxyvitamin D levels.^{23,24} Finally, it has recently been suggested by Heaney and colleagues²⁵ that the point at which hepatic 25(OH)D production becomes zero order could be used to define the lower end of normal vitamin D status. In this work, the investigators found serum 25(OH)D to rapidly increase as circulating vitamin D₃ (cholecalciferol) increased. When circulating vitamin D₃ exceeds approximately 5.8 ng/mL, the hepatic 25-hydroxylase appears to become saturated and this reaction switches from first order to zero order. Taking this approach would define the lower limit of normal at approximately 35 ng/mL, 25 obviously quite close to the 30 to 32 ng/mL suggested by other end points such as the relationship with PTH.

It is plausible that some of the debate surrounding what value defines "optimal" 25(OH)D status is being confounded by different levels for various tissues and end points; that is, the cutpoint for various nonclassic targets of vitamin D might vary from that for bone.²⁶ Furthermore, it is possible that the 25(OH)D value for "optimal" physiologic functioning might differ between individuals. Having a range of "normal" for virtually all clinically measured biologic parameters is well known by clinicians. As vitamin D is, in essence, an endogenously produced hormone, it is not surprising that between-individual variability and regulation would exist. In this regard, the skin of humans,²⁷ and other animals,²⁸ possesses the ability to regulate cholecalciferol production. Moreover, limited data suggest that variation in vitamin D degradation may exist, in that differences in 24-hydroxylase capacity between individuals may be based on race.²⁹ Data from a study of adults in Hawaii supports between-individual differences despite abundant sun exposure.30 In fact, inspection of serum 25(OH)D concentration in that cohort reveals a virtually normal or Gaussian distribution (Fig. 2). Indeed, other studies of highly UV-exposed adults^{31,32} find some individuals with "low" 25(OH)D despite high UV exposure and a fairly broad range of what it would seem logical to define as "normal," as noted in Fig. 2. Thus, it seems likely that some of these people with "low" 25(OH)D levels could in fact be "optimal" for them. It is clearly accepted that a range exists for multiple physiologic functions that is considered "normal" for healthy adults; it is not known whether the same should apply to 25(OH)D.

Given these data, it is not surprising that an exact 25(OH)D cutpoint to define suboptimal vitamin D status remains somewhat controversial.³³ Despite this, there is increasing agreement that values less than approximately 30 to 32 ng/mL be identified as "low."^{34,35} When this cutpoint is applied, low vitamin D status is extremely common worldwide. For example, recent reports classify 52% to 77% of the studied cohorts as "low" using 30 ng/mL as a cutpoint.^{21,36–38} Even the more restrictive cutpoint of less than 20 ng/mL identifies 18% to 36% as "low" (**Fig. 3**). It is not surprising that studies report a variable prevalence of low vitamin D status as the studied cohorts differ in

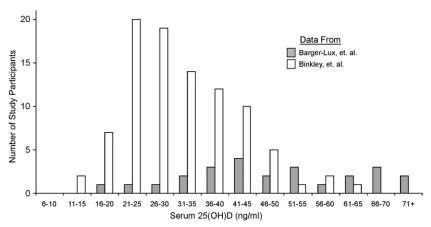


Fig. 2. Distribution of serum 25(OH)D in highly sun-exposed adults. In these 2 studies in which the average total body sun exposure was approximately 11 hours per week, a broad, and somewhat Gaussian, distribution of circulating 25(OH)D is apparent. Note that the study by Barger-Lux and colleagues used a 25(OH)D assay that measures approximately 10% higher than the HPLC assay used in the report by Binkley and colleagues. (*Data from* Binkley N, Novotny R, Krueger D, et al. Low vitamin D status despite abundant sun exposure. J Clin Endocrinol Metab 2007;92:2130–5; and Barger-Lux MJ, Heaney RP. Effects of above average summer sun exposure on serum 25-hydroxyvitamin D and calcium absorption. J Clin Endocrinol Metab 2002;87:4952–6.)

age, sex, race, body mass index, and dietary vitamin D intake. Although it is often assumed that some of the variable prevalence of low vitamin D reflects limited availability of sun exposure due to living at higher latitudes, it is of interest that a recent meta-analysis involving 394 studies comprising more than 32,000 subjects found no

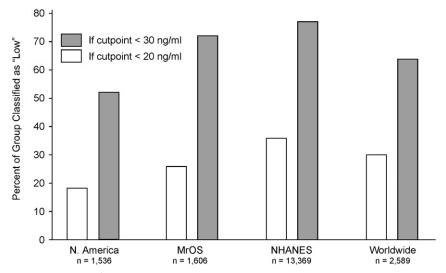


Fig. 3. Prevalence of low vitamin D status in various populations. In these recent cohort studies, low vitamin D status, whether defined as a 25(OH)D level below 20 ng/mL or below 30 ng/mL, is extremely common. (*Data from* Refs.^{21,36–38}.)

influence of latitude on 25(OH)D concentration.³⁹ It seems likely that the absence of an effect based on latitude reflects current human indoor lifestyles, clothing, and sun-avoidance behavior. Moreover, it is probable that these factors are contributing to worsening population vitamin D status. Although measurement issues confound data interpretation, recent National Health and Nutrition Examination Survey data report a decline in mean 25(OH)D concentration from the 1988 to 1994 data collection to that of 2000 to 2004.^{36,40} In summary, despite variation in 25(OH)D methodology, cutpoint selected, and cohort studied, it is clear that low vitamin D status is common worldwide.

LOW VITAMIN D STATUS: CONSEQUENCES Bone

Low vitamin D status has long been associated with osteomalacia/rickets, and a role in osteoporosis pathogenesis via calcium malabsorption and secondary hyperparathyroidism has more recently been suggested. Consistent with an important role of low vitamin D status in osteoporosis, recent meta-analyses find low 25(OH)D to be associated with higher fracture risk. A1-43 In addition, a dose effect was reported, with greater vitamin D intakes and higher achieved 25(OH)D concentrations providing superior fracture reduction benefit. In summary, while one can debate the cutpoint, low vitamin D status leads to adverse bone consequences.

Muscle Function and Falls

Both genomic and nongenomic effects of vitamin D on muscle have been proposed. A4,45 Regardless of the mechanism, patients with osteomalacia due to vitamin D deficiency develop muscle pain and weakness that is improved with vitamin D therapy. Muscle biopsy in such people reveals atrophy of the fast twitch (type II) fibers. As type II fibers are first to be recruited to avoid falling, this observation may explain the increased falls risk in vitamin D deficient individuals. Of note, randomized prospective studies find vitamin D to reduce risk of falls by more than 20%. It seems likely that reducing falls contributes in a major way to the fracture reduction efficacy observed with vitamin D. Moreover, similar to the relationship observed with fracture, a higher vitamin D dose provides greater reduction in the risk of falls. The 25(OH)D concentration needed to optimize leg function has been explored, with various cutpoints (eg, 16–24 ng/mL) suggested. S5,53 A recent review finds 25(OH)D concentrations less than approximately 16 ng/mL to be associated with substantially poorer leg function, but additionally finds values greater than approximately 36 to 40 ng/mL to be optimal.

Cancer

Vitamin D has antiproliferative and prodifferentiating effects on many cell types.⁵⁴ It has been proposed that these effects are related to local production of 1,25-dihydroxyvitamin D, thus favorably impacting genes affecting cellular proliferation/differentiation and thereby reducing cancer risk.⁷ Consistent with this, an extensive, albeit largely associational, literature exists relating higher latitude, low vitamin D intake, and/or less sunlight exposure to increased risk of, or mortality from, multiple types of cancer.^{55–59} Prospective trials of vitamin D supplementation with cancer as an end point are very limited; the Women's Health Initiative did not demonstrate a reduction in colon cancer risk, perhaps related to the low daily dose (400 IU) of vitamin D used.⁶⁰ However, a smaller prospective study of postmenopausal women found calcium plus vitamin D₃ (1100 IU daily) to reduce overall cancer risk by approximately

60%.⁶¹ To summarize, physiologically logical hypotheses, observational data, and one small randomized trial find low vitamin D status to be associated with higher cancer risk. Additional prospective studies are needed.

Other Conditions

It is likely that vitamin D has immune modulating effects. It has long been recognized that vitamin D deficiency is associated with respiratory infections, which perhaps contributed to the use of cod liver oil in antituberculous therapy. ^{62,63} More recently, it has become appreciated that calcitriol enhances monocyte mycobacterial killing, likely by facilitating production of the antimicrobial protein, cathelicidin. ⁶⁴ Moreover, helper type 1 and 2 cells are vitamin D targets, with vitamin D causing a shift toward an anti-inflammatory profile. ^{65–68} Thus, it is not surprising that low vitamin D status is associated with an increased risk of autoimmune and potentially infectious diseases. ^{69–71} In addition, inflammation is increasingly being recognized as a contributor to the pathogenesis of various diseases, and vitamin D modulates inflammatory cytokine production. ^{72–74}

It has been suggested that endemic low vitamin D status is contributing to the increased prevalence of diabetes mellitus. Multiple potential mechanisms have been proposed, including vitamin D increasing insulin production/secretion. In addition, observational studies associate low vitamin D status with diabetes type 1 and type 2. Prospective studies of vitamin D supplementation are clearly indicated; however, on the whole it appears that low vitamin D status impairs glucose metabolism.

Observational studies report an association between low vitamin D status and cardiovascular disease. ^{81–85} Potential mechanisms include a vitamin D effect on the endothelium, ⁸⁶ vascular smooth muscle, ^{87,88} and/or cardiomyocytes, ⁸⁹ all of which possess the vitamin D receptor. Prospective studies to further evaluate this reported association are needed.

In summary, low vitamin D status has been associated with a variety of diseases, and biologically plausible hypotheses exist to suggest a possible causal role. However, until confirmed by randomized studies, it is wise to be cautious and recognize that association does not prove causation.

WHEN SHOULD VITAMIN D STATUS BE ASSESSED?

Given the multitude of potential adverse health consequences ascribed to low vitamin D status, it is not surprising that screening 25(OH)D measurement has been advocated. Such screening may in fact be appropriate, if it becomes established that low vitamin D status contributes in a causal manner to multiple adverse health outcomes, for example, cardiovascular disease, diabetes, hypertension, and so forth, with which it is currently associated. However, in the absence of randomized trials documenting benefit for these varied outcomes, population-based screening seems premature.

At this time, rather than advocating a population screening approach, it seems reasonable to measure 25(OH)D in those identified as being at high risk of vitamin D deficiency and those for whom a prompt musculoskeletal response to optimization of vitamin D status could be expected. Such groups include those with osteoporosis, a history of falls or with high risk of falls, malabsorption (eg, celiac disease, radiation enteritis, bariatric surgery, and so forth), individuals with liver disease, and those requiring medications known to alter vitamin D status, for example, certain anticonvulsants. Given the relationship of low vitamin D status with cancer, it also seems rational to measure 25(OH)D in those with malignancy. 92

Alternatively, it could be argued that simple treatment of all individuals with vitamin D should be advocated, thereby making 25(OH)D measurement unnecessary. Although this approach is attractive, it is unfortunately problematic in that no expert consensus exists regarding a recommended dose. For example, the National Osteoporosis Foundation recommends 800 to 1000 IU daily, 93 whereas some vitamin D experts suggest values over 2000 IU. 94 Moreover, as discussed later, vitamins D₂ and D₃ appear to not be equally potent in maintaining 25(OH)D.95,96 As such, daily intake of 1000 IU of vitamin D₂ may well not be equal to 1000 IU of vitamin D₃. In addition, vitamin D dosing may differ by age in that older adults likely require higher vitamin D intakes because of to the lower capability of skin to produce vitamin D with advancing age. 97 Similarly, clear differences exist between races, with African Americans requiring higher intakes than Caucasian Americans; Hispanic individuals may have intermediate requirements.98 Some of these differences in required intake may reflect differences in cutaneous melanin content⁹⁹; however, other less well understood between-individual differences in vitamin D absorption and subsequent metabolism may well play a role. In this regard, even among individuals of similar age and race/ethnicity, substantial between-individual variability in response to equal vitamin D intake is noted (Fig. 4). Thus, if a health care provider wishes to assure optimal vitamin D status in an individual patient, it is necessary to obtain a 25(OH)D measurement.

APPROACHES TO VITAMIN D REPLETION/SUPPLEMENTATION

Increasing exposure to sunlight would be an effective and free approach to improving vitamin D status. However, this does not seem to be a viable approach, given

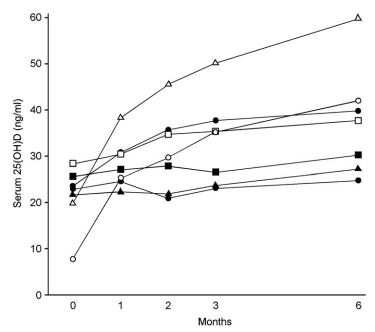


Fig. 4. Variable response to daily vitamin D_3 . In these 7 Caucasian older adults (age 66–88 years), all of whom started the study with a 25(OH)D level less than 30 ng/mL, the variable response to daily administration of 1600 IU vitamin D_3 is apparent. (*From* Binkley N, Gemar D, Woods A, et al. Effect of vitamin D_2 or vitamin D_3 supplementation on serum 25OHD. J Bone Miner Res 2008;23(Suppl 1):S350; with permission.)

widespread sun-avoidance campaigns 100-103 based on the association of UV exposure with skin cancer. 104 Sun avoidance and sunscreen use 55,100,105,106 reduce skin exposure to UV radiation and thereby reduce skin vitamin D production. 2,107 In the face of such pervasive and powerful efforts, advocating sun exposure as a population-based measure to improve vitamin D status faces grave obstacles. Despite this, exposure to sunlight in moderation, perhaps for 15 minutes prior to sunscreen application, seems reasonable and is free. It should be noted that due to differences in skin pigmentation, season, latitude, time of day of sun exposure, and amount of body surface exposed, simple recommendations such as "15 minutes of sun on the hands and face" are overly simplistic and will not assure optimal vitamin D status in all people.

Higher dose vitamin D treatment approaches to the clinical correction of vitamin D deficiency, and when to monitor 25(OH)D status during and following vitamin D treatment/supplementation, have received surprisingly little attention. Various "high-dose" repletion approaches, for example, 50,000 IU 3 times weekly, weekly or monthly, have been evaluated. $^{108-111}$ A recent evaluation of clinical approaches found vitamin D_2 regimens using more than 600,000 IU administered over an average time of 2 months achieved 25(OH)D values greater than 30 ng/mL in 64% of patients, with the highest value being 100 ng/mL. 108 An additional clinical report of "high-dose" vitamin D_2 (50,000 IU once weekly for up to 3 years) achieved a 25(OH)D level above 30 ng/mL in 23 of 24 patients, with the highest reported value being 100 ng/mL. 112

Maintenance of vitamin D status with daily doses from 1000 to 4000 IU have been studied. 96,113,114 As noted above, between-individual variability exists. However, a reasonable clinical "rule of thumb" is that the addition of 1000 IU of vitamin D_3 daily can be expected to increase circulating 25(OH)D by approximately 10 ng/mL.

Although daily vitamin D supplementation is very inexpensive (approximately \$1 per month), available data find daily vitamin D supplementation to be less effective than expected at increasing serum 25(OH)D status, simply due to failure to reliably take the supplements. This finding is hardly surprising in that suboptimal adherence to prescribed therapies for a variety of conditions is well known to clinicians. However, based on the increasing calcium intake of the United States population over time temperated to widespread educational programs), it seems feasible that similar approaches to informing the public about health benefits of vitamin D supplementation could improve endemic low vitamin D status. An alternative, and highly viable, approach is increased availability of vitamin D fortified foods coupled with higher amounts of vitamin D per serving in such food.

How best to monitor 25(OH)D status in individuals receiving vitamin D therapy has not been systematically evaluated. However, as compliance/adherence with many daily therapies (including vitamins) is often poor, monitoring 25(OH)D status 4 to 6 months after initiating treatment in those at high risk (eg, patients with osteomalacia, fragility fractures, or high risk of falls) seems reasonable. Repeat evaluation at earlier time points seems inappropriate, as it takes 3 to 6 months for serum 25(OH)D to plateau following initiation of supplementation.

WHAT IS VITAMIN D TOXICITY?

Both clinicians and patients may express concern that the "high" amounts of vitamin D noted here will lead to toxicity. It is clear that huge doses of vitamin D do lead to hypercalcemia and hypercalciuria. However, there is no clear-cut definition of which level of serum 25(OH)D should be considered "toxic." This doubt has led to variability in the clinical reporting of 25(OH)D results, with some laboratories reporting possibly

toxic levels as being above 80 ng/mL while others include up to 100 ng/mL as being within the reference range. Such variability is not surprising, as recent expert opinion suggests "the serum 25(OH)D concentration that is the threshold for vitamin D toxicity has not been established." However, a review of the published vitamin D toxicity cases finds all reports of hypercalcemia due to vitamin D intoxication to be associated with 25(OH)D concentrations greater than 88 ng/mL. 120

Regarding what constitutes "high" 25(OH)D values, it seems reasonable that highly sun-exposed individuals could be used to assist in the determination of "normal" vitamin D status.³³ When such individuals are evaluated, it appears that the highest attainable 25(OH)D values from cutaneous production are in the 70 to 80 ng/mL range.^{30,31} Thus, the current approach of reporting 80 to 100 ng/mL as the upper limit of normal seems appropriate.

DOES THE EFFECT OF VITAMIN D2 DIFFER FROM THAT OF VITAMIN D3?

Two chemically distinct forms of vitamin D exist: vitamin D₃ (cholecalciferol) is a 27carbon molecule, whereas vitamin D₂ (ergocalciferol) contains 28 carbons and differs from vitamin D₃ by the presence of an additional methyl group and a double bond between carbons 22 and 23. Although clear chemical differences exist, whether vitamin D₂ and vitamin D₃ are equally effective at increasing 25(OH)D and have equivalent physiologic effects remains unclear. At present, these 2 forms are regarded as equal and interchangeable. However, some data suggest that vitamin D₂ is less "potent" at maintaining serum 25(OH)D than is vitamin D₃.95,96,121 Although a recent report challenges this 113 and found D2 and D3 to be equally effective, the vast majority of studies find vitamin D₃ to be somewhat more potent. It seems possible that this reflects lower affinity of vitamin D₂ for vitamin D binding protein in the circulation, leading to more rapid clearance. As such, use of supplements containing vitamin D_3 , rather than vitamin D_2 , seems appropriate. 122 It is unfortunate that vitamin D_2 is the only high-dose prescription vitamin D preparation in the United States and some other countries. Despite its lower potency, use of high-dose vitamin D₂ does increase circulating 25(OH)D concentration.

SUMMARY

Low vitamin D status is extremely common worldwide due to low dietary intake and low skin production. Suboptimal vitamin D status contributes to many conditions, including osteomalacia/rickets, osteoporosis, falls, and fractures. It is possible or even likely that low vitamin D status increases risk for a multitude of other conditions. Although consensus does not exist, it appears that circulating 25(OH)D concentrations greater than 30 to 32 ng/mL are needed for optimal health. To achieve this, daily intakes of at least 1000 IU of D_3 daily are required, and it is probable that substantially higher amounts are required to achieve such values on a population basis. It seems premature to recommend widespread screening for 25(OH)D measurement. Targeted measurement in those at increased risk for vitamin D deficiency and those most likely to have a prompt positive response to supplementation is appropriate. Widespread optimization of vitamin D status likely will lead to prevention of many diseases with attendant reduction of morbidity, mortality, and expense.

REFERENCES

1. Holick MF. Environmental factors that influence the cutaneous production of vitamin D. Am J Clin Nutr 1995;61:638S-45S.

- 2. Holick MF. The photobiology of vitamin D and its consequences for humans. Ann N Y Acad Sci 1985:453:1–13.
- 3. Looker AC, Dawson-Hughes B, Calvo MS, et al. Serum 25-hydroxyvitamin D status of adolescents and adults in two seasonal subpopulations from NHANES III. Bone 2002;30:771–7.
- 4. Rucker D, Allan JA, Fick GH, et al. Vitamin D insufficiency in a population of healthy western Canadians. Can Med Assoc J 2002;166:1517–24.
- 5. Nesby-O'Dell S, Scanlon KS, Cogswell ME, et al. Hypovitaminosis D prevalence and determinants among African American and white women of reproductive age: Third National Health and Nutrition Examination Survey, 1988-1994. Am J Clin Nutr 2002;76:187–92.
- Calvo MS, Whiting SJ. Prevalence of vitamin D insufficiency in Canada and the United States: importance to health status and efficacy of current food fortification and dietary supplement use. Nutr Rev 2003;61:107–13.
- 7. Holick MF. Vitamin D deficiency. N Engl J Med 2007;357:266-81.
- 8. Binkley N. Does low vitamin D status contribute to "age-related" morbidity? J Bone Miner Res 2007;22:V55–8.
- 9. DeLuca HF. The vitamin D story: a collaborative effort of basic science and clinical medicine. FASEB J 1988;2:224–36.
- Standing Committee on the Scientific Evaluation of Dietary Reference Intakes Food and Nutrition Board, Institute of Medicine. Dietary reference intakes for calcium phosphorus, magnesium, vitamin D and fluoride. Washington, DC: National Academy Press; 1997.
- 11. Lips P, Chapuy MC, Dawson-Hughes B, et al. An international comparison of serum 25-hydroxyvitamin D measurements. Osteoporos Int 1999;9: 394–7.
- 12. Binkley N, Krueger D, Cowgill C, et al. Assay variation confounds hypovitaminosis D diagnosis: a call for standardization. J Clin Endocrinol Metab 2003;89: 3152–7.
- 13. Binkley N. Vitamin D: clinical measurement and use. J Musculoskelet Neuronal Interact 2006;6:338–40.
- Binkley N, Krueger D, Engelke JA, et al. What is your patient's vitamin D status?
 Clinical consideration of variability in a 25(OH)D measurement. J Bone Miner Res 2008;23(Suppl 1):S351.
- 15. Heaney RP. Functional indices of vitamin D status and ramifications of vitamin D deficiency. Am J Clin Nutr 2004;80(Suppl):1706S–9S.
- 16. Dawson-Hughes B, Heaney RP, Holick MF, et al. Estimates of optimal vitamin D status. Osteoporos Int 2005;16:713–6.
- 17. McCollum EV, Simmonds N, Becker JE, et al. An experimental demonstration of the existence of a vitamin which promotes calcium deposition. J Biol Chem 1922;53:293–312.
- 18. Steenbock H. The induction of growth promoting and calcifying properties in a ration by exposure to light. Science 1924;60:224–5.
- 19. DeLuca HF. Overview of general physiologic features and functions of vitamin D. Am J Clin Nutr 2004;80(Suppl):1689S–96S.
- 20. Lips P. Vitamin D deficiency and secondary hyperparathyroidism in the elderly: Consequences for bone loss and fractures and therapeutic implications. Endocr Rev 2001;22:477–501.
- 21. Holick MF, Siris ES, Binkley N, et al. Prevalence of vitamin D inadequacy among postmenopausal North American women receiving osteoporosis therapy. J Clin Endocrinol Metab 2005;90:3215–24.

- 22. Heaney RP, Dowell MS, Hale CA, et al. Calcium absorption varies within the reference range for serum 25-hydroxyvitamin D. J Am Coll Nutr 2003;22:142–6.
- 23. Need AG, Nordin BEC. Misconceptions—vitamin D insufficiency causes malabsorption of calcium. Bone 2008;42:1021–4.
- Need AG, O'Loughlin PD, Morris HA, et al. Vitamin D metabolites and calcium absorption in severe vitamin D deficiency. J Bone Miner Res 2008; 23:1859–63.
- 25. Heaney RP, Armas LAG, Shary JR, et al. 25-hydroxylation of vitamin D_3 : relation to circulating vitamin D_3 under various input conditions. Am J Clin Nutr 2008;87: 1738–42.
- 26. Bischoff-Ferrari HA, Giovannucci E, Willett WC, et al. Estimation of optimal serum concentrations of 25-hydroxyvitamin D for multiple health outcomes. Am J Clin Nutr 2006;84:18–28.
- Holick MF, MacLaughlin JA, Doppelt SH. Regulation of cutaneous previtamin D₃
 photosynthesis in man: skin pigment is not an essential regulator. Science 1981;
 211:590–3.
- 28. Ferguson GW, Gehrmann WH, Karsten KB, et al. Ultraviolet exposure and vitamin D synthesis in a sun-dwelling and a shade-dwelling species of Anolis: are there adaptations for lower ultraviolet B and dietary vitamin D₃ availability in the shade? Physiol Biochem Zool 2005;78:193–200.
- 29. Awumey EMK, Mitra DA, Hollis BW, et al. Vitamin D metabolism is altered in Asian Indians in the Southern United States: a clinical research center study. J Clin Endocrinol Metab 1998;83:169–73.
- 30. Binkley N, Novotny R, Krueger D, et al. Low vitamin D status despite abundant sun exposure. J Clin Endocrinol Metab 2007;92:2130–5.
- 31. Barger-Lux MJ, Heaney RP. Effects of above average summer sun exposure on serum 25-hydroxyvitamin D and calcium absorption. J Clin Endocrinol Metab 2002:87:4952–6.
- 32. Tangpricha V, Turner A, Spina C, et al. Tanning is associated with optimal vitamin D status (serum 25-hydroxyvitamin D concentration) and higher bone mineral density. Am J Clin Nutr 2004;80:1645–9.
- 33. Hollis BW. Circulating 25-hydroxyvitamin D levels indicative of vitamin D sufficiency: Implications for establishing a new effective dietary intake recommendation for vitamin D. J Nutr 2005;135:317–22.
- 34. Hollis BW. Assessment of vitamin D status and definition of a normal circulating range of 25-hydroxyvitamin D. Curr Opin Endocrinol Diabetes Obes 2008;15: 489–94.
- 35. Kuchuk NO, Pluijm SMF, van Schoor NM, et al. Relationships of serum 25-hydroxyvitamin D to bone mineral density and serum parathyroid hormone and markers of bone turnover in older adults. J Clin Endocrinol Metab 2009;94:1244–50.
- 36. Ginde AA, Liu MC, Camargo CA. Demographic differences and trends of vitamin D insufficiency in the US population, 1988-2004. Arch Intern Med 2009;169:626–32.
- 37. Lips P, Hosking D, Lippuner K, et al. The prevalence of vitamin D inadequacy amongst women with osteoporosis: an international epidemiological investigation. J Intern Med 2006;260:245–54.
- 38. Orwoll E, Nielson CM, Marshall LM, et al. Vitamin D deficiency in older men. J Clin Endocrinol Metab 2009;94:1214–22.
- 39. Hagenau T, Vest R, Gissel TN, et al. Global vitamin D levels in relation to age, gender, skin pigmentation and latitude: an ecologic meta-regression analysis. Osteoporos Int 2009;20:133–40.

- Looker AC, Pfeiffer CM, Lacher DA, et al. Serum 25-hydroxyvitamin D status of the US population: 1988-1994 compared with 2000-2004. Am J Clin Nutr 2008; 88:1519–27.
- 41. Cauley JA, La Croix A, Wu L, et al. Serum 25-hydroxyvitamin D concentrations and risk for hip fractures. Ann Intern Med 2008;149:242–50.
- 42. Bischoff-Ferrari HA, Willett WC, Wong JB, et al. Fracture prevention with vitamin D supplementation: a meta-analysis of randomized controlled trials. JAMA 2005; 293:2257–64.
- 43. Bischoff-Ferrari HA, Willett WC, Wong JB, et al. Prevention of nonvertebral fractures with oral vitamin D and dose dependency. Arch Intern Med 2009;169:551–61.
- 44. Pfeifer M, Begerow B, Minne HW. Vitamin D and muscle function. Osteoporos Int 2002;13:187–94.
- 45. Janssen HCJP, Samson MM, Verhaar HJJ. Vitamin D deficiency, muscle function, and falls in elderly people. Am J Clin Nutr 2002;75:611–5.
- 46. Mingrone D, Greco AV, Castagneto M, et al. A woman who left her wheelchair. Lancet 1999;353:806.
- 47. Skaria J, Katiyar BC, Srivastava TP, et al. Myopathy and neuropathy associated with osteomalacia. Acta Neurol Scand 1975;51:37–58.
- 48. Glerup H, Mikkelsen K, Poulsen L, et al. Hypovitaminosis D myopathy without biochemical signs of osteomalacic bone involvement. Calcif Tissue Int 2000; 66:419–24.
- 49. Flicker L, Mead K, MacInnis RJ, et al. Serum vitamin D and falls in older women in residential care in Australia. J Am Geriatr Soc 2003;51:1533–8.
- 50. Bischoff-Ferrari HA, Dawson-Hughes B, Willett WC, et al. Effect of vitamin D on falls: a meta-analysis. JAMA 2004;291:1999–2006.
- 51. Dawson-Hughes B, Harris SS, Krall EA, et al. Effect of calcium and vitamin D supplementation on bone density in men and women 65 years of age or older. N Engl J Med 1997;337:670–6.
- 52. Broe KE, Chen TC, Weinberg J, et al. A higher dose of vitamin D reduces the risk of falls in nursing home residents: a randomized, multiple-dose study. J Am Geriatr Soc 2007;55:234–9.
- Wicherts IS, van Schoor NM, Boeke AJP, et al. Vitamin D status predicts physical performance and its decline in older persons. J Clin Endocrinol Metab 2007;92: 2058–65.
- 54. Bikle D. Nonclassic actions of vitamin D. J Clin Endocrinol Metab 2009;94: 26-34
- 55. Garland CF, Garland FC, Gorham ED, et al. The role of vitamin D in cancer prevention. Am J Public Health 2006;96:252–61.
- Gorham ED, Garland CF, Garland FC, et al. Optimal vitamin D status for colorectal cancer prevention A quantitative meta analysis. Am J Prev Med 2007; 32:210–6.
- 57. Giovannucci E, Liu Y, Rimm EB, et al. Prospective study of predictors of vitamin D status and cancer incidence and mortality in men. J Natl Cancer Inst 2006;98:451–9.
- 58. Feskanich D, Ma J, Fuchs CS, et al. Plasma vitamin D metabolites and risk of colorectal cancer in women. Cancer Epidemiol Biomarkers Prev 2004;13: 1502–8.
- 59. Garland CF, Gorham ED, Mohr SB, et al. Vitamin D and prevention of breast cancer: pooled analysis. J Steroid Biochem 2007;103:708–11.
- 60. Wactawski-Wende J, Kotchen JM, Anderson GL, et al. Calcium plus vitamin D supplementation and the risk of colorectal cancer. N Engl J Med 2006;354: 684–96.

- 61. Lappe JM, Travers-Gustafson D, Davies KM, et al. Vitamin D and calcium supplementation reduces cancer risk: results of a randomized trial. Am J Clin Nutr 2007;85:1586–91.
- 62. Martineau AR, Honecker FU, Wilkinson RJ, et al. Vitamin D in the treatment of pulmonary tuberculosis. J Steroid Biochem Mol Biol 2007;103:793–8.
- 63. Russell B. The history of lupus vulgaris: its recognition, nature, treatment and prevention. Proc R Soc Med 1954;48:127–32.
- 64. Liu PT, Stenger S, Li H, et al. Toll-like receptor triggering of a vitamin D-mediated human antimicrobial response. Science 2006;311:170–3.
- 65. van Etten E, Mathieu C. Immunoregulation by 1,25-dihydroxyvitamin D₃: Basic concepts. J Steroid Biochem Mol Biol 2005;97:93–101.
- 66. Bemiss CJ, Mahon BD, Henry A, et al. Interleukin-2 is one of the targets of 1,25-dihydroxyvitamin D_3 in the immune system. Arch Biochem Biophys 2002;402: 249–54.
- 67. Mahon BD, Wittke A, Weaver V, et al. The targets of vitamin D depend on the differentiation and activation status of CD4 positive T cells. J Cell Biochem 2003;402:922–33.
- 68. Cantorna MT, Humpal-Winter J, DeLuca HF. In vivo upregulation on interleukin-4 is one mechanism underlying the immunoregulatory effects of 1,25-dihydroxyvitamin D₃. Arch Biochem Biophys 2000;377:135–8.
- 69. Peterlik M, Cross HS. Vitamin D and calcium deficits predispose for multiple chronic diseases. Eur J Clin Invest 2005;35:290–304.
- 70. Froicu M, Weaver V, Wynn TA, et al. A crucial role for the vitamin D receptor in experimental inflammatory bowel diseases. Mol Endocrinol 2003;17: 2386–92.
- Cantorna MT, Hayes CE, DeLuca HF. 1,25-dihydroxyvitamin D₃ reversibly blocks the progression of relapsing encephalomyelitis, a model of multiple sclerosis. Proc Natl Acad Sci U S A 1996;93:7861–4.
- 72. Boonstra A, Barrat FJ, Craine C, et al. 1 alpha 25-dihydroxyvitamin D_3 has a direct effect on naive CD4+ T cells to enhance the development of TH2 cells. J Immunol 2001;167:4974–80.
- 73. Willheim M, Thien R, Schrattbauer K, et al. Regulatory effects of 1 alpha 25 dihydroxyvitamin D₃ on cytokine production of human peripheral blood lymphocytes. J Clin Endocrinol Metab 1999;84:3739–44.
- 74. Rigby WF, Denome S, Fanger MW. Regulation of lymphokine production and human T-lymphocyte activation by 1,25 dihydroxyvitamin D₃: Specific inhibition at the level of messenger RNA. J Clin Invest 1987;79:1659–64.
- 75. Kadowaki S, Norman AW. Demonstration that the vitamin D metabolite $1,25(OH)_2$ -vitamin D_3 and not $24R,25(OH)_2$ -vitamin D_3 is essential for normal insulin secretion in the perfused rat pancreas. Diabetes 1985;34: 315–20.
- 76. Norman AW, Frankel JB, Heldt AM, et al. Vitamin D deficiency inhibits pancreatic secretion of insulin. Science 1980;209:823–5.
- 77. Rabinovitch A, Suarez-Pinzon WL, Sooy K, et al. Expression of calbindin-D28K in a pancreatic islet b-cell line protects against cytokine-induced apoptosis and necrosis. Endocrinology 2001;142:3649–55.
- 78. Mathieu C, Gysemans C, Guilietti A, et al. Vitamin D and diabetes. Diabetologia 2005;48:1247–57.
- 79. Zipitis CS, Akobeng AK. Vitamin D supplementation in early childhood and risk of type 1 diabetes: a systematic review and meta-analysis. Arch Dis Child 2008; 93:512–7.

- 80. Pittas AG, Lau J, Hu FB, et al. The role of vitamin D and calcium in type 2 diabetes. A systematic review and meta-analysis. J Clin Endocrinol Metab 2007;92:2017–29.
- 81. Grimes DS, Hindle E, Dyer T. Sunlight, cholesterol and coronary heart disease. QJM 1996;89:579–89.
- 82. Voors AW, Johnson WD. Altitude and arteriosclerotic heart disease mortality in white residents of 99 of the 100 largest cities in the United States. J Chronic Dis 1979;32:157–62.
- 83. Rostand SG. Ultraviolet light may contribute to geographic and racial blood pressure differences. Hypertension 1997;30:150–6.
- 84. Scragg R, Jackson RD, Holdaway IM, et al. Myocardial infarction is inversely associated with plasma 25-hydroxyvitamin D_3 levels: A community based study. Int J Epidemiol 1990;19:559–63.
- 85. Poole KE, Loveridge N, Barker PJ, et al. Reduced vitamin D in acute stroke. Stroke 2006;37:243–5.
- 86. Merke J, Milde P, Lewicka S, et al. Identification and regulation of 1,25-dihydroxyvitamin D_3 receptor activity and biosynthesis of 1,25-dihydroxyvitamin D_3 : studies in cultured bovine aortic endothelial cells and human dermal capillaries. J Clin Invest 1989;83:1903–15.
- 87. Somjen D, Weisman Y, Kohen F, et al. 25-hydroxyvitamin D_3 -1 alpha hydroxylase is expressed in human vascular smooth muscle cells and is upregulated by parathyroid hormone and estrogenic compounds. Circulation 2005;111:1666–71.
- 88. Merke J, Hofmann W, Goldschmidt D, et al. Demonstration of 1,25 (OH) $_2$ vitamin D $_3$ receptors and actions in vascular smooth muscle cells in vitro. Calcif Tissue Int 1987;41:112–4.
- 89. Holick MF. High prevalence of vitamin D inadequacy and implications for health. Mayo Clin Proc 2006;81:353–73.
- 90. Holick MF. Too little vitamin D in premenopausal women: why should we care? Am J Clin Nutr 2002;76:3-4.
- 91. Giovannucci E. Can vitamin D reduce total mortality? Arch Intern Med 2007;167: 1709–10.
- 92. Anonymous. Vitamin D deficiency: information for cancer patients. New York: The Bone and Cancer Foundation; 2008.
- 93. Anonymous. Clinician's guide to prevention and treatment of osteoporosis. Washington, DC: National Osteoporosis Foundation; 2008.
- 94. Heaney RP. Barriers to optimizing vitamin D₃ intake for the elderly. J Nutr 2006; 136:1123–5.
- 95. Armas LAG, Hollis BW, Heaney RP. Vitamin D₂ is much less effective than vitamin D₃ in humans. J Clin Endocrinol Metab 2004;89:5387–91.
- 96. Binkley N, Gemar D, Woods A, et al. Effect of vitamin D₂ or vitamin D₃ supplementation on serum 25OHD. J Bone Miner Res 2008;23(Suppl 1):S350.
- 97. MacLaughlin JA, Holick MF. Aging decreases the capacity of human skin to produce vitamin D₃. J Clin Invest 1985;76:1536–8.
- 98. Weaver CM, Fleet JC. Vitamin D requirements: current and future. Am J Clin Nutr 2004;80:1735S–9S.
- 99. Matsuoka LY, Wortsman J, Haddad JG, et al. Racial pigmentation and the cutaneous synthesis of vitamin D. Arch Dermatol 1991;127:536–8.
- Anonymous. National coalition for sun safety. Available at: http://www.aad.org/ public/sunsafetydb.htm. Accessed February 10, 2010.
- 101. Task Force on Community Preventive Services. Recommendations to prevent skin cancer by reducing exposure to ultraviolet radiation. Am J Prev Med 2004;27:467–70.

- Kirsner RS, Parker DF, Brathwaite N, et al. Sun protection policies in Miami-Dade county public schools: opportunities for skin cancer prevention. Pediatr Dermatol 2005;22:513–9.
- 103. Anonymous. Saving your skin from sun damage. Am Fam Physician 2006;74: 815–6.
- 104. Randle HW. Suntanning: differences in perceptions throughout history. Mayo Clin Proc 1997;72:461–6.
- 105. Skin cancer primary prevention and education initiative 2006 Sun safety at school: What you can do. Center for Disease Control; Guidelines for School. Available at: www.cdc.gov/cancer/skin/pdf/sunsafety_v0908.pdf. Accessed February 10, 2010.
- 106. US Environmental Protection Agency. SunWise Program. Available at: www.epa. gov/sunwise. Accessed February 10, 2010.
- Matsuoka LY, Wortsman J, Hanafin N, et al. Chronic sunscreen use decreases circulating concentrations of 25-hydroxyvitamin D. Arch Dermatol 1988;124: 1802–4.
- 108. Pepper KJ, Judd SE, Nanes MS, et al. Evaluation of vitamin D repletion regimens to correct vitamin D status in adults. Endocr Pract 2009;15:95–103.
- 109. Przybelski R, Agrawal S, Krueger D, et al. Rapid correction of low vitamin D status in nursing home residents. Osteoporos Int 2008;19:1621–8.
- 110. Malabanan A, Veronikis IE, Holick MF. Redefining vitamin D insufficiency. Lancet 1998;351:805–6.
- 111. Geller JL, Adams JS. Vitamin D therapy. Current Osteoporos Rep 2008;6:5-11.
- 112. Ramamurthy R, Przybelski R, Gemar D, et al. Long-term high-dose vitamin D supplementation in the elderly is both safe and efficacious. J Clin Densitom 2009;12:375–6.
- 113. Holick MF, Biancuzzo RM, Chen TC, et al. Vitamin D_2 is as effective as vitamin D_3 in maintaining circulating concentrations of 25-hydroxyvitamin D. J Clin Endocrinol Metab 2008;93:677–81.
- 114. Heaney RP, Davies KM, Chen TC, et al. Human serum 25-hydroxycholecalciferol response to extended oral dosing with cholecalciferol. Am J Clin Nutr 2003;77: 204–10.
- 115. The Record Trial Reporting Group. Oral vitamin D_3 and calcium for secondary prevention of low-trauma fractures in elderly people (Randomized Evaluation of Calcium or vitamin D, RECORD): a randomized-placebo-controlled trial. Lancet 2005;365:1621–8.
- 116. Wactawski-Wende J, Jackson RD, LaCroix AZ, et al. Calcium plus vitamin D supplementation and the risk of fractures. N Engl J Med 2006;354:669–83.
- 117. Chesnut CHI. Treating osteoporosis with bisphosphonates and addressing adherence. Drugs 2006;66:1351–9.
- 118. Briefel RR, Johnson CL. Secular trends in dietary intake in the United States. Annu Rev Nutr 2004;24:401–31.
- 119. Vieth R. Vitamin D toxicity, policy and science. J Bone Miner Res 2007;22(Suppl 2):V64–8.
- 120. Vieth R. Vitamin D supplementation, 25-hydroxyvitamin D concentrations and safety. Am J Clin Nutr 1999;69:842–56.
- 121. Trang HM, Cole DEC, Rubin LA, et al. Evidence that vitamin D₃ increases serum 25-hydroxyvitamin D more efficiently than does vitamin D₂. Am J Clin Nutr 1998; 68:854–8.
- 122. Houghton LA, Vieth R. The case against ergocalciferol (vitamin D2) as a vitamin supplement. Am J Clin Nutr 2006;84:694–7.